

SUPPLEMENTARY INFORMATION

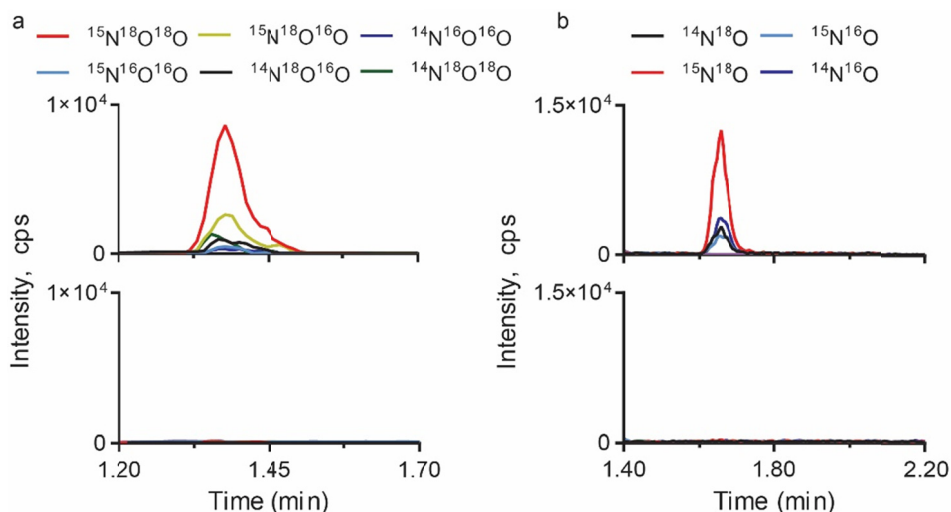
Convergence of Biological Nitration and Nitrosation via Symmetrical Nitrous Anhydride

Dario A. Vitturi¹, Lucia Minarrieta², Sonia R. Salvatore¹, Edward M. Postlethwait³, Marco Fazzari^{1,4}, Gerardo Ferrer-Sueta^{5,6}, Jack R. Lancaster Jr.^{1,7,8}, Bruce A. Freeman^{1,#} and Francisco J. Schopfer^{1,#}.

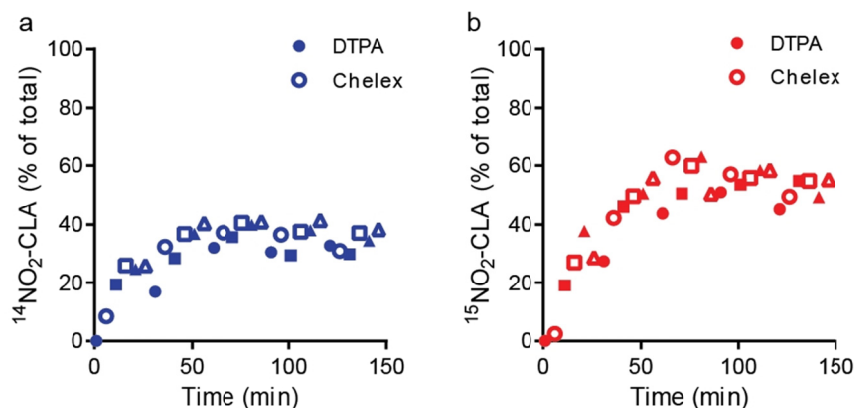
Departments of ¹Pharmacology and Chemical Biology, ⁷Medicine and ⁸Surgery. University of Pittsburgh. Pittsburgh, PA, USA. ²Cátedra de Inmunología, ⁵Laboratorio de Físicoquímica Biológica, and ⁶Center for Free Radical and Biomedical Research. Universidad de la República, Montevideo, Uruguay. ³Department of Environmental Health Sciences, University of Alabama at Birmingham, Birmingham, AL, USA. ⁴Fondazione Ri.MED, Palermo, Italy.

[#]To whom correspondence may be addressed: Bruce A. Freeman, Ph.D. or Francisco J. Schopfer, Ph.D., Department of Pharmacology & Chemical Biology, Thomas E. Starzl Biomedical Science Tower E1340, 200 Lothrop St, University of Pittsburgh, Pittsburgh, PA 15213, USA; Tel: 648-9319; Fax: (412) 648-2229; Email: freerad@pitt.edu or fjs2@pitt.edu.

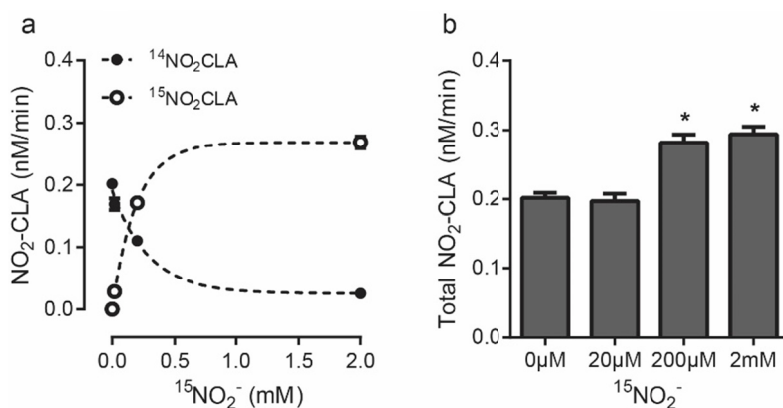
SUPPLEMENTARY RESULTS



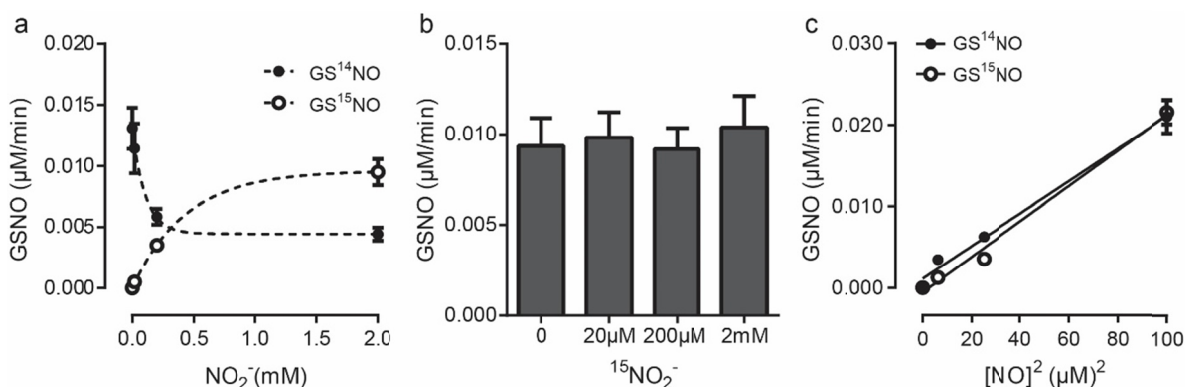
Supplementary Figure 1. NO_2^- does not mediate CLA nitration and GSH nitrosation in the absence of $\bullet\text{NO}$. **a**, CLA (20 μM) and $^{15}\text{N}^{18}\text{O}_2^-$ (2 mM) were incubated for 100 min in 20 mM BisTris + 100 μM DTPA pH 7.0 and 25 $^\circ\text{C}$ in the presence (top panel) or absence (bottom panel) of 25 μM MNO. Representative LC-MS/MS traces show no formation of $\text{NO}_2\text{-CLA}$ isotopologues in the absence of $\bullet\text{NO}$ donor. **b**, GSH (20 μM) and $^{15}\text{N}^{18}\text{O}_2^-$ (2 mM) were incubated for 120 min in 20 mM BisTris + 100 μM DTPA pH 7.0 and 25 $^\circ\text{C}$ in the presence (top panel) or absence (bottom panel) of 2.5 μM MNO. Representative traces demonstrate that GSNO formation was not detected in the absence of $\bullet\text{NO}$ donor.



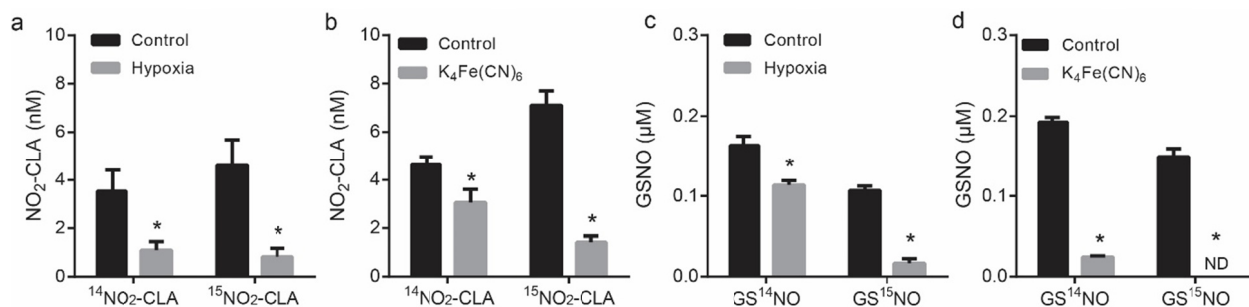
Supplementary Figure 2. NO_2^- dependent CLA nitration is independent of metal catalysis. Identical yields and distribution of $^{14}\text{NO}_2\text{-CLA}$ (a) and $^{15}\text{NO}_2\text{-CLA}$ (b) formation from CLA (20 μM) nitration in the presence of 200 μM $^{15}\text{NO}_2^-$ using either DTPA (100 μM) or Chelex treatment to remove adventitious metals from the reaction buffer (20 mM BisTris pH 7.0).



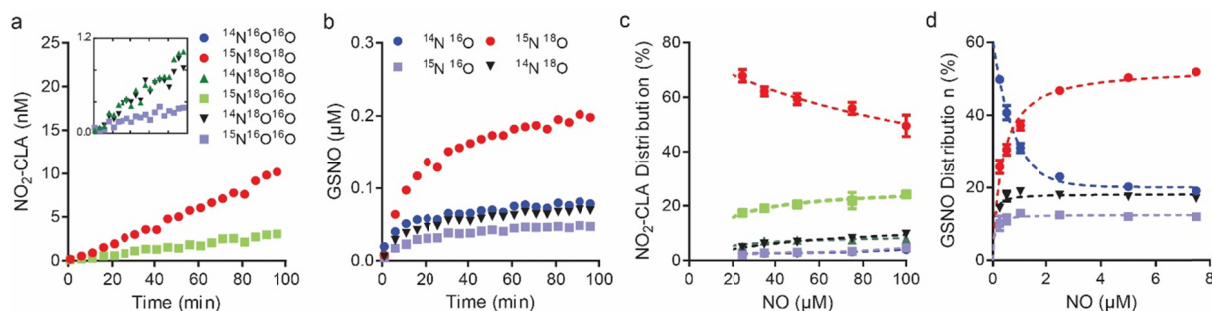
Supplementary Figure 3. $^{15}\text{NO}_2^-$ participates in CLA nitration in the absence of cellular components. **a**, Plot of initial rates of $\text{NO}_2\text{-CLA}$ formation showing reciprocal modulation of the ^{14}N and ^{15}N containing products by $^{15}\text{NO}_2^-$. Data are means \pm SE ($n=4$), with non-linear fits provided for illustrative purposes. **b**, The global initial rate of $\text{NO}_2\text{-CLA}$ formation ($^{14}\text{NO}_2\text{-CLA} + ^{15}\text{NO}_2\text{-CLA}$) is moderately increased by $^{15}\text{NO}_2^-$ concentration versus no-nitrite controls as determined by one way ANOVA and Bonferroni's multiple comparison test. Data are means \pm SD ($n=4$), * $p < 0.0001$.



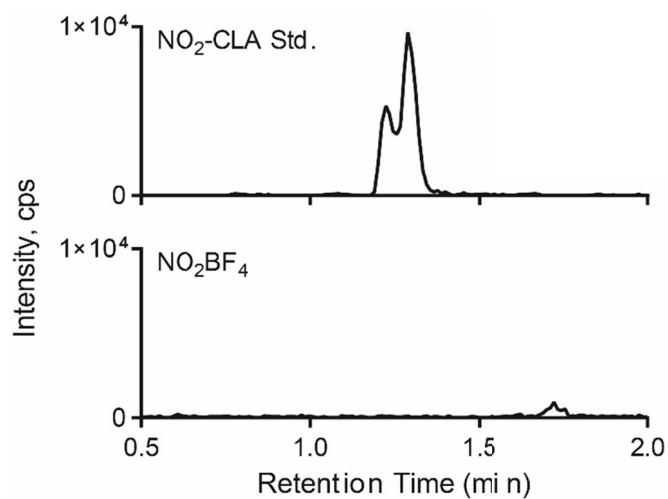
Supplementary Figure 4. $^{15}\text{NO}_2^-$ is incorporated into S-nitrosoglutathione in the presence of $\bullet\text{NO}$. **a**, Initial GS^{14}NO and GS^{15}NO formation rates plotted versus $^{15}\text{NO}_2^-$ concentration illustrating reciprocal modulation of ^{14}N - and ^{15}N -containing products. Data are means \pm SD ($n=4$), fits are provided for illustrative purposes. **b**, No effect of $^{15}\text{NO}_2^-$ in the global initial rate of GSNO formation as determined by one way ANOVA. Data are means \pm SD ($n=4$). **c**, Initial rates of GS^{14}NO and GS^{15}NO formation are linearly dependent on $[\bullet\text{NO}]^2$ ($R^2 > 0.99$), consistent with a process limited by $\bullet\text{NO}$ autoxidation (20 μM GSH, 200 μM $^{15}\text{NO}_2^-$). Data are means \pm SD ($n=4$).



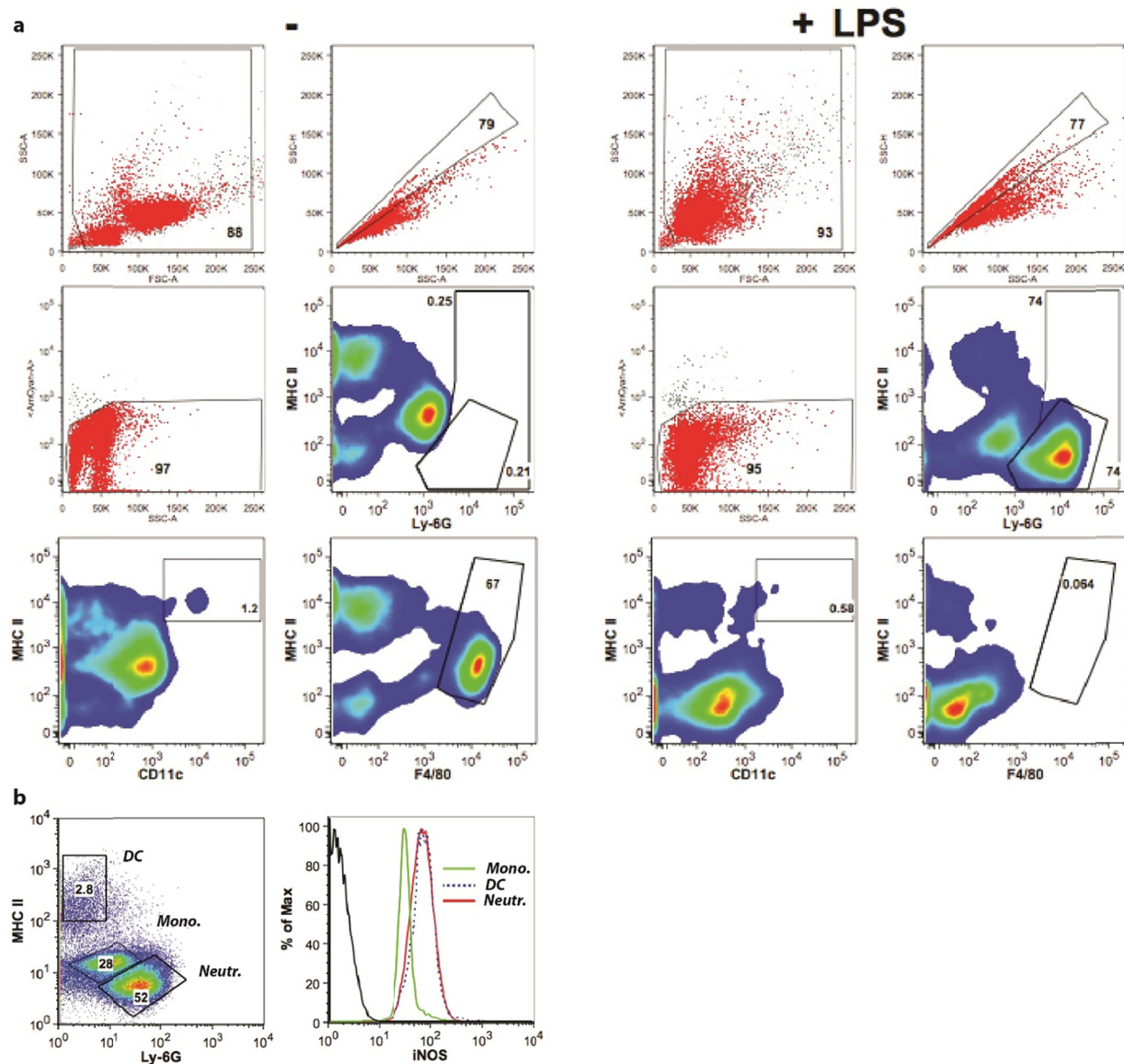
Supplementary Figure 5. $\bullet\text{NO}_2$ is required for nitrite-dependent $^{15}\text{NO}_2\text{-CLA}$ and GS^{15}NO formation. a-b, Formation of $^{14}\text{NO}_2\text{-CLA}$ and $^{15}\text{NO}_2\text{-CLA}$ (25 μM MNO, 200 μM $^{15}\text{NO}_2^-$, 90 minutes) is inhibited both under hypoxic conditions (a) and by 1mM $\text{K}_4\text{Fe}(\text{CN})_6$ (b), * $p < 0.001$ versus corresponding control. c-d, GS^{14}NO and GS^{15}NO formation (2.5 μM MNO, 200 μM $^{15}\text{NO}_2^-$, 90 minutes) is inhibited by low oxygen tension (c) and 1mM $\text{K}_4\text{Fe}(\text{CN})_6$ (d), * $p < 0.0001$ versus corresponding control. Data are mean \pm SD (n=4) of independent experiments. Two-way ANOVA and Bonferroni's multiple comparison were used to test statistical significance.



Supplementary Figure 6. NO_2^- incorporation into GSNO and $\text{NO}_2\text{-CLA}$ is associated with symN_2O_3 formation. a, Time-dependent generation of mixed $^{14}\text{N}/^{15}\text{N}$ and $^{18}\text{O}/^{16}\text{O}$ $\text{NO}_2\text{-CLA}$ isotopologues from the reaction of 2 mM $^{15}\text{N}^{18}\text{O}_2^-$ with of 25 μM MNO. Insert shows additional low-abundance $\text{NO}_2\text{-CLA}$ isotopologues. Data are representative traces generated by combining time-staggered replicates (n=4). b, Formation of mixed $^{14}\text{N}/^{15}\text{N}$ and $^{18}\text{O}/^{16}\text{O}$ GSNO from 2.5 μM MNO and 2 mM $^{15}\text{N}^{18}\text{O}_2^-$. Traces represent four time-staggered replicates. c, $\text{NO}_2\text{-CLA}$ distribution versus $\bullet\text{NO}$ concentration in the presence of 2 mM $^{15}\text{N}^{18}\text{O}_2^-$. Data are mean \pm SD (n=4), fits are provided for illustrative purposes. d, Percent GSNO distribution in the presence of 2 mM $^{15}\text{N}^{18}\text{O}_2^-$ nitrite, with increased formation of isotopically-scrambled products at higher $\bullet\text{NO}$ concentrations. Data are mean \pm SD (n=4), fits are provided for illustrative purposes.

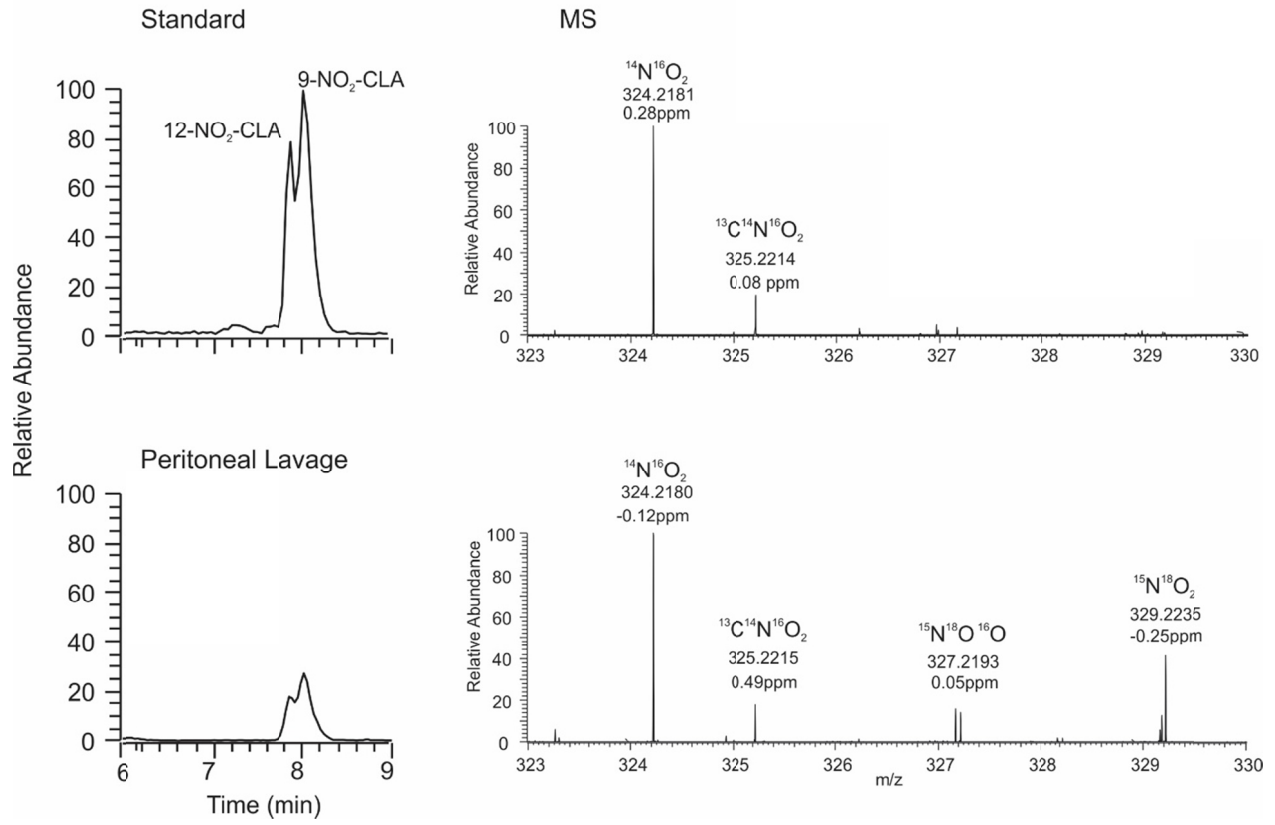


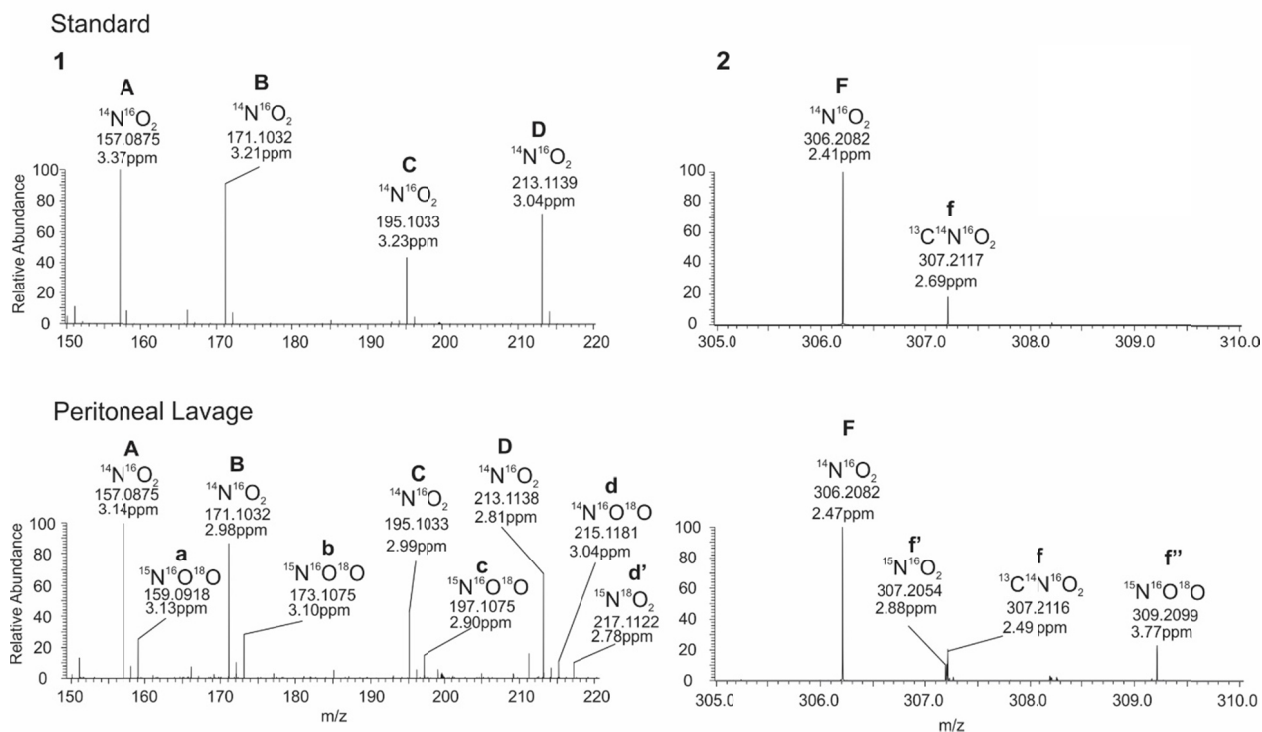
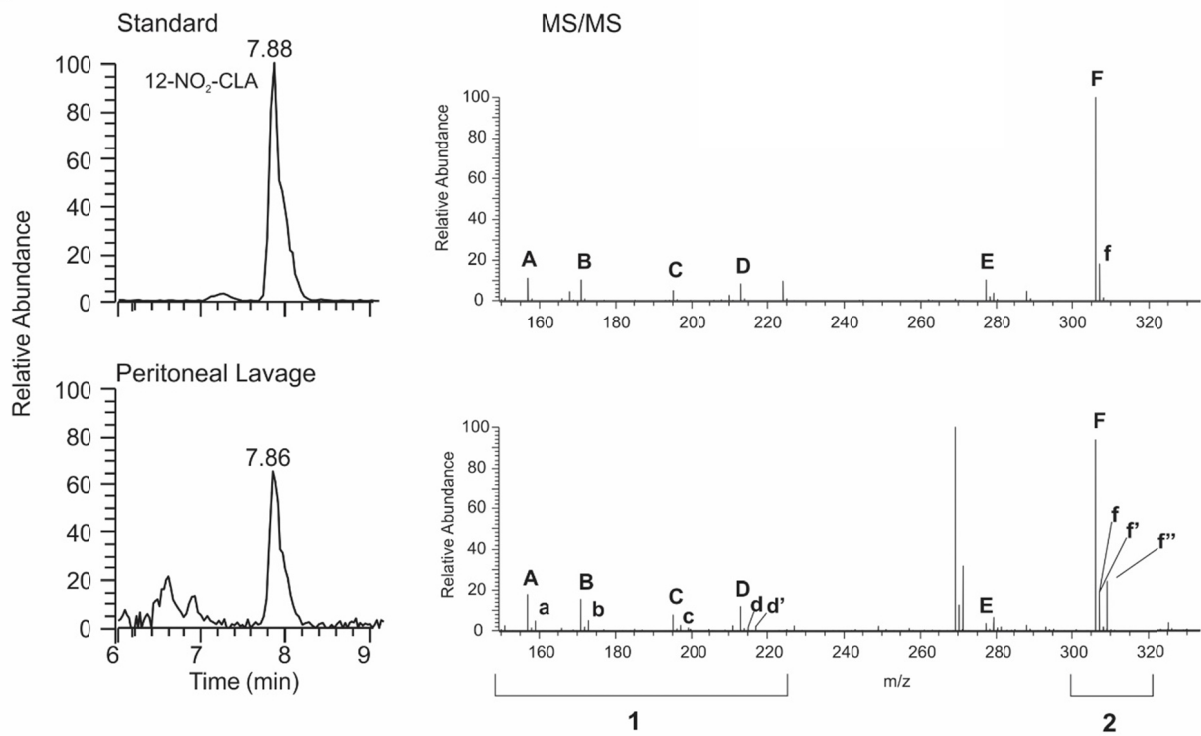
Supplementary Figure 7. NO₂⁺ does not mediate NO₂-CLA CLA nitration. Representative LC-MS/MS traces showing no CLA (20 μM) nitration by NO₂BF₄ (50 μM) following 90 min incubation in 20 mM BisTris + 100 μM DTPA pH 7.0 and 25 °C (bottom panel). A NO₂-CLA standard is provided in the top panel for reference.

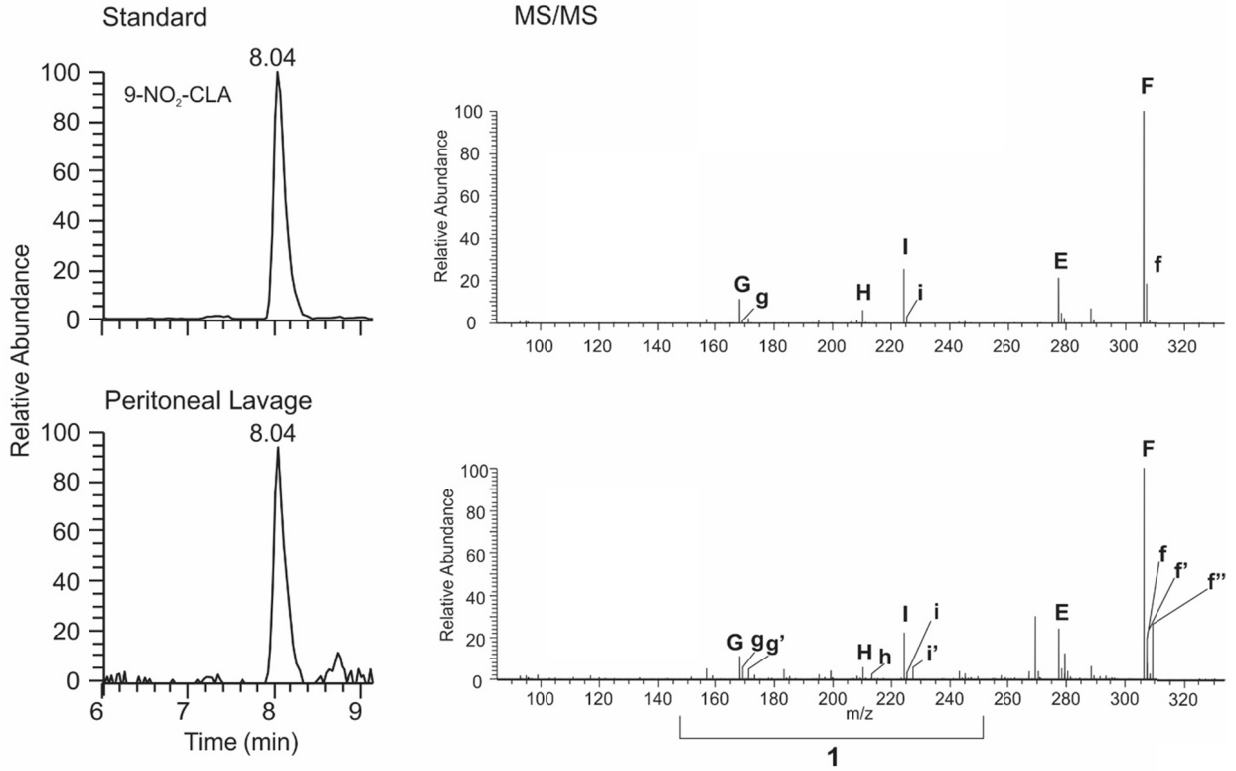
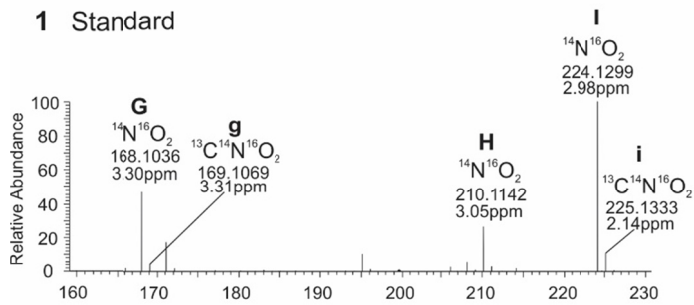
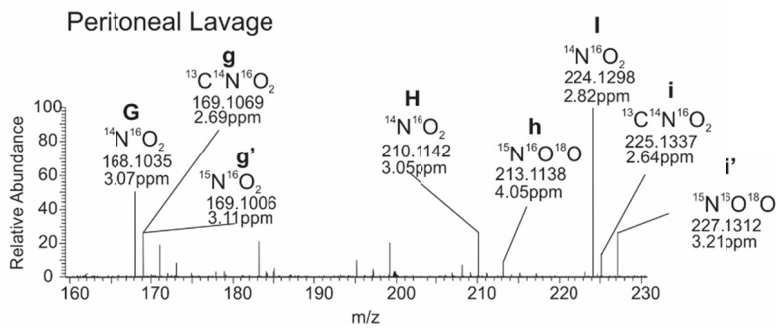


Supplementary Figure 8. Distribution of peritoneal cell subpopulations during LPS-induced inflammation. **a**, Intraperitoneal LPS injection in an incomplete Freund adjuvant emulsion leads to resident macrophage depletion (F480⁺, MHCII^{INT}) and massive recruitment of neutrophils (Ly6G^{HI}, CD11b⁺) and inflammatory monocytes (Ly6G^{INT}, CD11b⁺) 18 hours after treatment. Dendritic cells are also present (CD11c⁺, MHCII^{HI}) in the peritoneal cavity. **b**, Distribution of iNOS expression among inflammatory cell subpopulations in LPS-treated mice.

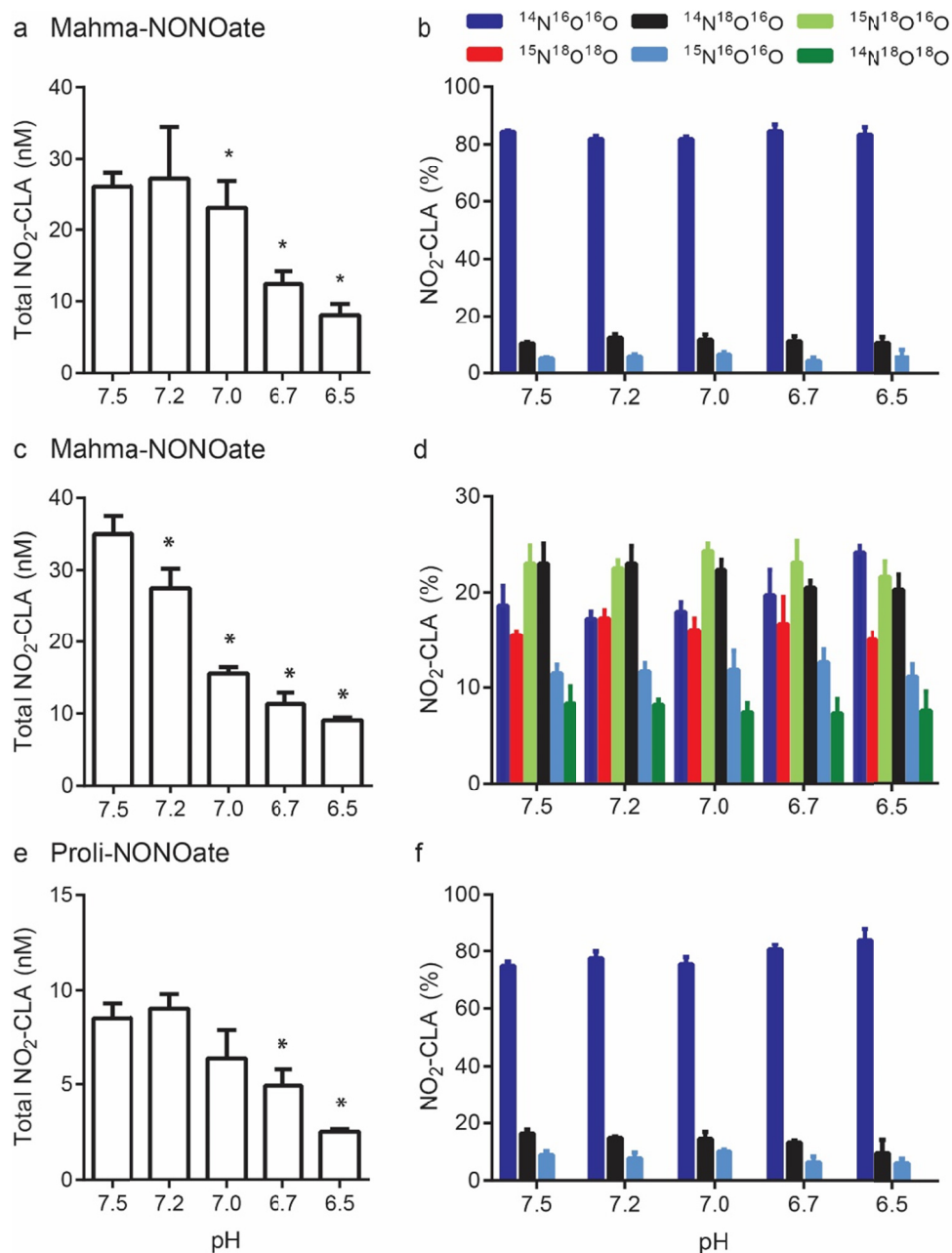
a



b

c**1 Standard****Peritoneal Lavage**

Supplementary Figure 9. High resolution MS/MS analysis of $^{15}\text{N}^{18}\text{O}_2^-$ -derived $\text{NO}_2\text{-CLA}$ isotopologues generated during acute peritoneal inflammation. **a**, LC-MS/MS analysis revealed two positional isomers for each $\text{NO}_2\text{-CLA}$ isotopologue, i.e. 9- $\text{NO}_2\text{-CLA}$ and 12- $\text{NO}_2\text{-CLA}$. An identical chromatographic profile and retention times were observed for both synthetic standards (upper panel) and peritonitis plus $^{15}\text{N}^{18}\text{O}_2$ supplementation samples (lower panel). **b-c**, Individual high resolution MS/MS analysis of the positional 9- $\text{NO}_2\text{-CLA}$ (**b**) and 12- $\text{NO}_2\text{-CLA}$ (**c**) isomers and their corresponding containing isotopologues showed the expected losses of H_2O (Δ mass = 18.0106) and H_2^{18}O (Δ mass = 20.0148) from the NO_2 group to form product ions with m/z 306.2082, 307.2054, and 309.2099 (peaks F, f' and f''). The naturally-occurring ^{13}C containing product ions were also resolved and were distinct from the ^{15}N products at a 60000 resolution (Panel b, peak f. Panel c, peaks g-i). Specific product ions derived from carbon chain fragmentation of the isotopologues further confirmed the incorporation of labeled NO_2^- and the formation of nitroalkene isomers identical to those observed in chemical reaction systems and *in vitro* cell culture models. In this regard, fragmentation of the isotopologues of 12- $\text{NO}_2\text{-CLA}$ generated the same pattern of product ions as the synthetic standard and as previously reported (References 13 and 14 in the main text). In addition to the ^{13}C isotopes observed in both the synthetic 12- $\text{NO}_2\text{-CLA}$ and the peritoneal inflammation sample, easily recognizable products ions containing isotopically-labeled O atoms were identified and confirmed at a 3 ppm resolution (Panel b, peaks A-D, a-d and d'). As previously reported, product ions stemming from carbon chain fragmentation of 12- $\text{NO}_2\text{-CLA}$ lose their nitrogen as the result of a cyclization reaction, thus no ions containing ^{15}N are observed (Figure 5b). The same approach was used to characterize 9- $\text{NO}_2\text{-CLA}$ isomers, where the fragmentation of this isomer is characterized by retaining the N atom. Biologically-generated 9- $\text{NO}_2\text{-CLA}$ shows the same fragmentation pattern as synthetic 9- $\text{NO}_2\text{-CLA}$ with the addition of the labeled ^{15}N and ^{18}O atoms confirmed at high resolution at the 3 ppm level (Panel c, peaks G-I, g'-i'). Corresponding structures and measured m/z for product ions detected in this figure are provided in Supplementary Table 2.



Supplementary Figure 10. pH dependence of CLA nitration by symN₂O₃. CLA nitration by 20 μM (a-b) and 200 μM (c-d) ¹⁵N¹⁸O₂⁻ was determined after 90 min incubations with 20 mM BisTris +100 μM DTPA and 25 μM mahma-NONOate at the indicated pH values. e-f, CLA nitration in the presence of 20 μM was induced by addition of 25 μM of the fast-releasing NO-donor proli-NONOate (half-life < 2 s) under the same conditions as before. For all panels, data are mean ± SD (n=4). Total CLA nitration was reduced at lower pH values as determined by one-way ANOVA analysis. * p < 0.05 versus pH = 7.5.

Supplementary Table 1. Exogenous and endogenous nitrite levels in peritoneal lavages 1 h post CLA/ $^{15}\text{N}^{18}\text{O}_2^-$ injection.

Nitrite Injected	Peritoneal Nitrite (nmoles)					
	$[^{14}\text{N}] \text{NO}_2^-$		$[^{15}\text{N}] \text{NO}_2^-$		Total	% $[^{15}\text{N}]$
	Mean	SD	Mean	SD		
20 nmoles	3.61	± 0.71	0.18	± 0.15	3.78	4.7 %
200 nmoles	3.27	± 1.97	0.30	± 0.14	3.57	8.4 %

Supplementary Table 2. Structures, measured m/z, atomic composition and corresponding precursors for product ions generated from 9- and 12-NO₂-CLA isotopologues isolated from peritoneal lavages.

	Fragment Exp	Fragment Theoretical	Fragment Composition	Fragment Structure	NO ₂ -FA
A	157.0875	157.087	C ₈ H ₁₃ O ₃ ⁻		12-NO₂-CLA Exp Mass: 324.2180 Exact Mass: 324.2180 or 12-¹⁵N¹⁸O¹⁶O-CLA Exp Mass: 327.2193 Exact Mass: 327.2193
B	171.1032	171.1027	C ₉ H ₁₅ O ₃ ⁻		
C	195.1033	195.1027	C ₁₁ H ₁₅ O ₃ ⁻		
D	213.1138	213.1132	C ₁₁ H ₁₇ O ₄ ⁻		
G	168.1035	168.103	C ₉ H ₁₄ NO ₂ ⁻		9-NO₂-CLA Exp Mass: 324.2180 Exact Mass: 324.2180
H	210.1142	210.1136	C ₁₁ H ₁₆ NO ₃ ⁻		
I	224.1298	224.1292	C ₁₂ H ₁₈ NO ₃ ⁻		
a	159.0918	159.0913	C ₈ H ₁₃ ¹⁸ O ¹⁶ O ₂ ⁻		12-¹⁵N¹⁸O¹⁶O-CLA Exp Mass: 327.2193 Exact Mass: 327.2193 or 12-¹⁵N¹⁸O₂-CLA Exp Mass: 329.2234 Exact Mass: 329.2236
b	173.1075	173.1069	C ₉ H ₁₅ ¹⁸ O ¹⁶ O ₂ ⁻		
c	197.1075	197.1069	C ₁₁ H ₁₅ ¹⁸ O ¹⁶ O ₂ ⁻		
d	215.1182	215.1175	C ₁₁ H ₁₇ ¹⁸ O ¹⁶ O ₃ ⁻		
d'	217.1122	217.1217	C ₁₁ H ₁₇ ¹⁸ O ₂ ¹⁶ O ₂ ⁻		
g'	169.1006	169.100	C ₉ H ₁₄ ¹⁵ NO ₂ ⁻		9-¹⁵N¹⁸O¹⁶O-CLA Exp Mass: 327.2193 Exact Mass: 327.2193 or 9-¹⁵N¹⁸O₂-CLA Exp Mass: 329.2234 Exact Mass: 329.2236
h	213.1138	213.1148	C ₁₁ H ₁₆ ¹⁵ N ¹⁸ O ¹⁶ O ₂ ⁻		
i'	227.1312	227.1305	C ₁₂ H ₁₈ ¹⁵ N ¹⁸ O ¹⁶ O ₂ ⁻		