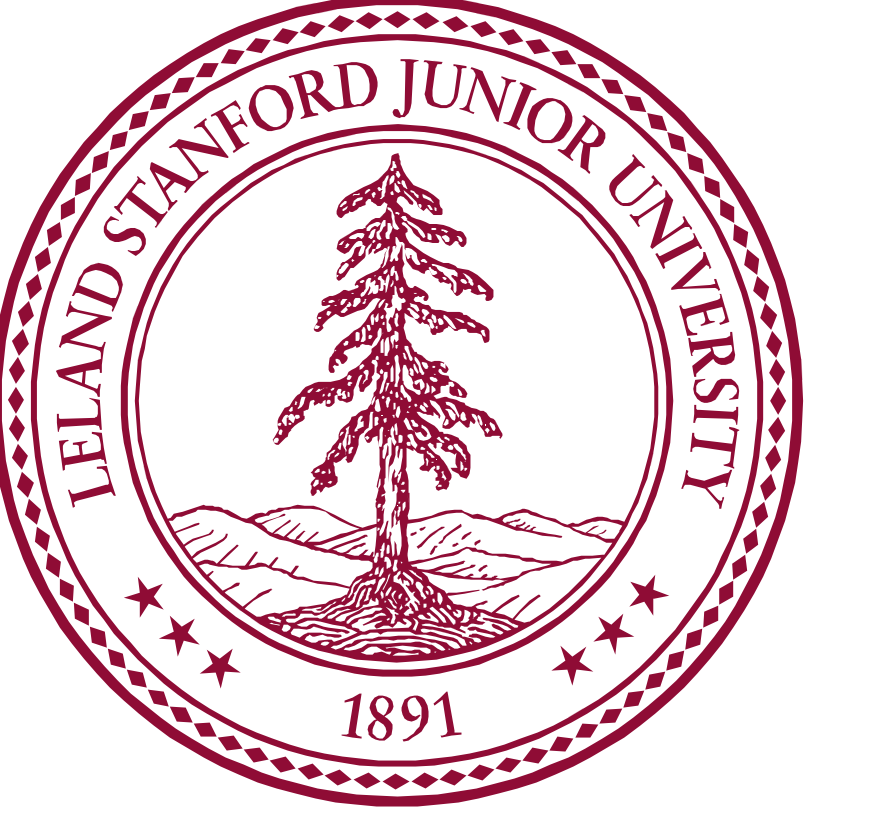




Low Infectivity among Asymptomatic Patients with a Positive SARS-CoV-2 Admission Test at a Tertiary Care Center, 2020–2022



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Introduction

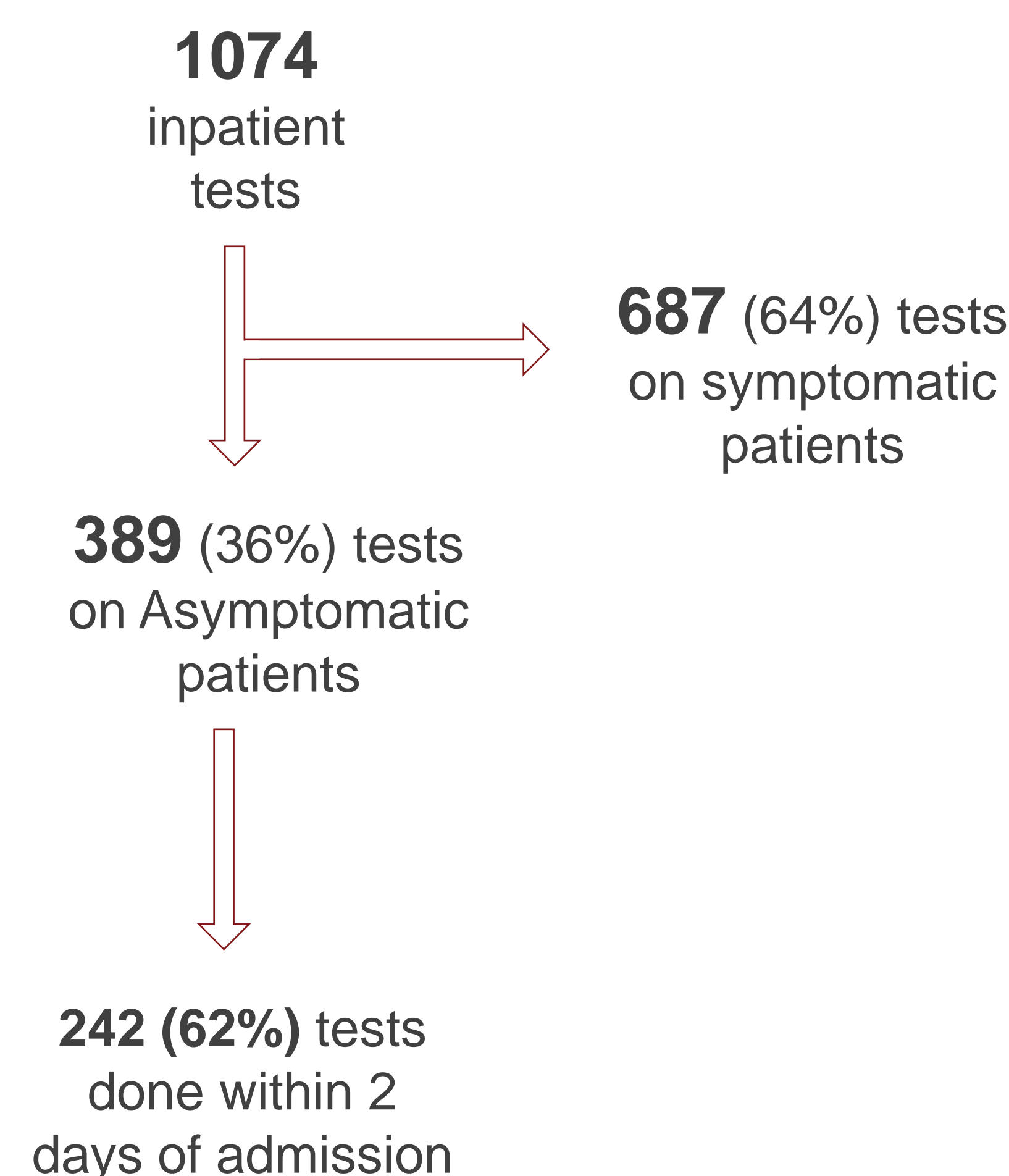
- Many hospitals have implemented admission SARS-CoV-2 testing to evaluate the need for transmission-based precautions.
- A positive test in an asymptomatic patient may represent:
 - 1) Active infection, signifying infectiousness.
 - 2) False positivity.
 - 3) Past infection with prolonged viral shedding, indicating non-infectiousness.
- We used a strand-specific SARS-CoV-2 real-time reverse polymerase chain reaction (rRT-PCR) to assess infectivity among asymptomatic patients with a positive SARS-CoV-2 PCR admission test.
- A positive minus strand PCR signified ongoing viral replication.

Objectives

- Identify the need for admission testing for asymptomatic patients.
- Describe the characteristics of asymptomatic patients who had a positive SARS-CoV-2 PCR with negative minus strand within 2 days of admission.

Methods

- We used a 2-step rRT-PCR specific to the minus strand of the SARS-CoV-2 envelope gene.
- We reviewed records of patients with a positive SARS-CoV-2 PCR who were also tested for the strand-specific SARS-CoV-2 PCR within two days of admission at Stanford Health Care during July 2020–April 2022.
- We restricted our analysis to each patient's first test. We calculated the percent of detectable minus strand-specific tests among asymptomatic patients over time and performed descriptive statistics for age, sex, immunocompromised state.



Results

Patient and test characteristics

- A total of 848 admitted patients had strand-specific SARS-CoV-2 assays performed.
- Out of 532 patients with a strand-specific assay done within 2 days of admission, 242 (45%) were asymptomatic.
- Among asymptomatic patients, the mean age was 56 years (range: 19-99 years), 133 were male (55%), 51 had immunocompromising conditions (21%), and 30 were admitted for a surgical procedure (12%).
- Among the 50 tests sent on immunocompromising patients, immunosuppressive conditions were as follows:
 - Solid organ transplant: 12 tests (24%)
 - CAR-T, BMT or active chemotherapy: 15 tests (30%)
 - Biologic agents or steroids: 4 tests (8%)
 - HIV/AIDS, Primary immunodeficiency, & other immunocompromising conditions: 19 tests (38%)
- A total of 21 had detectable minus strand-specific assays (9%, range: 4–25% per quarter).

Figure 1. Demographic data and ICU admission

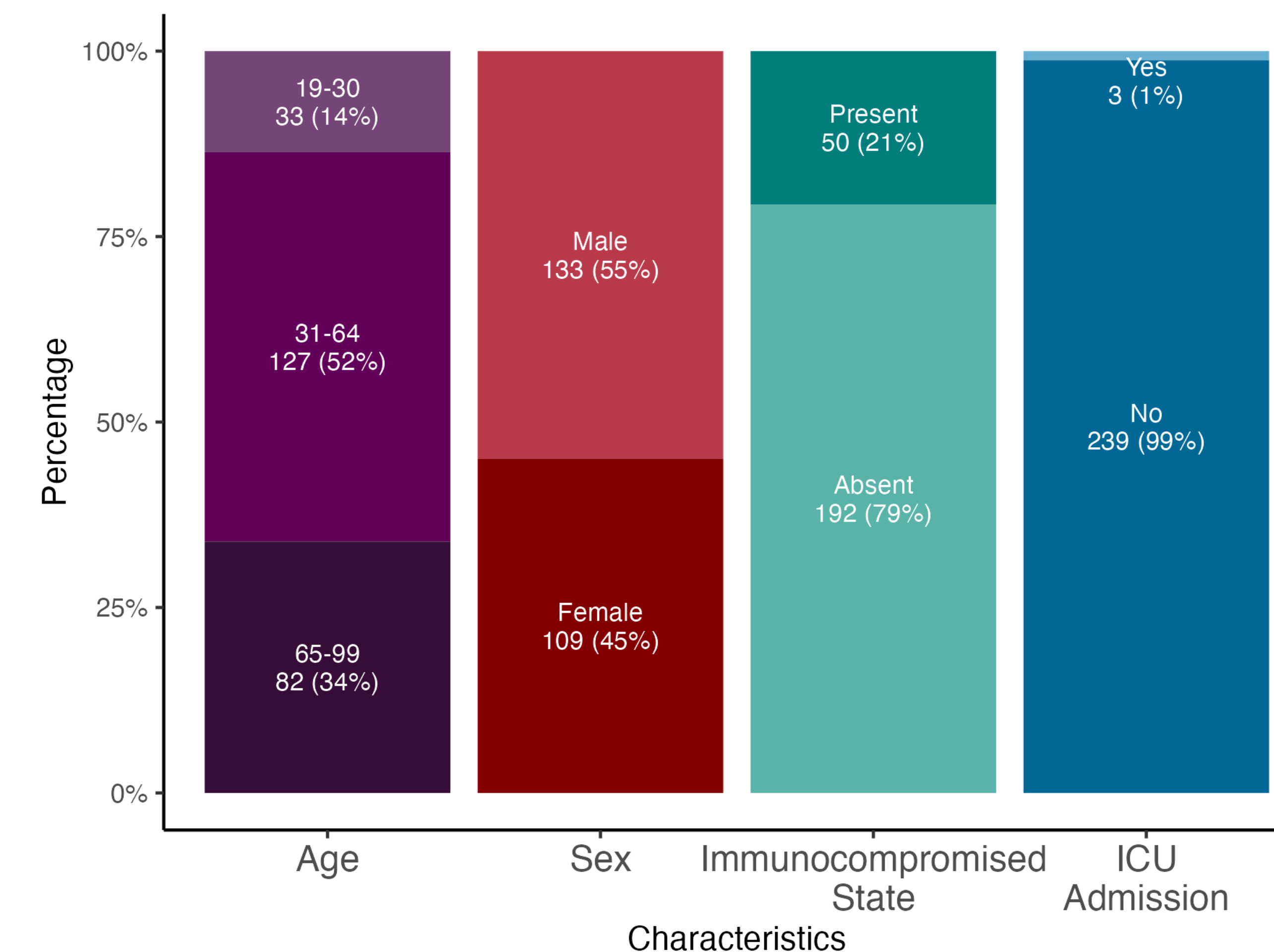
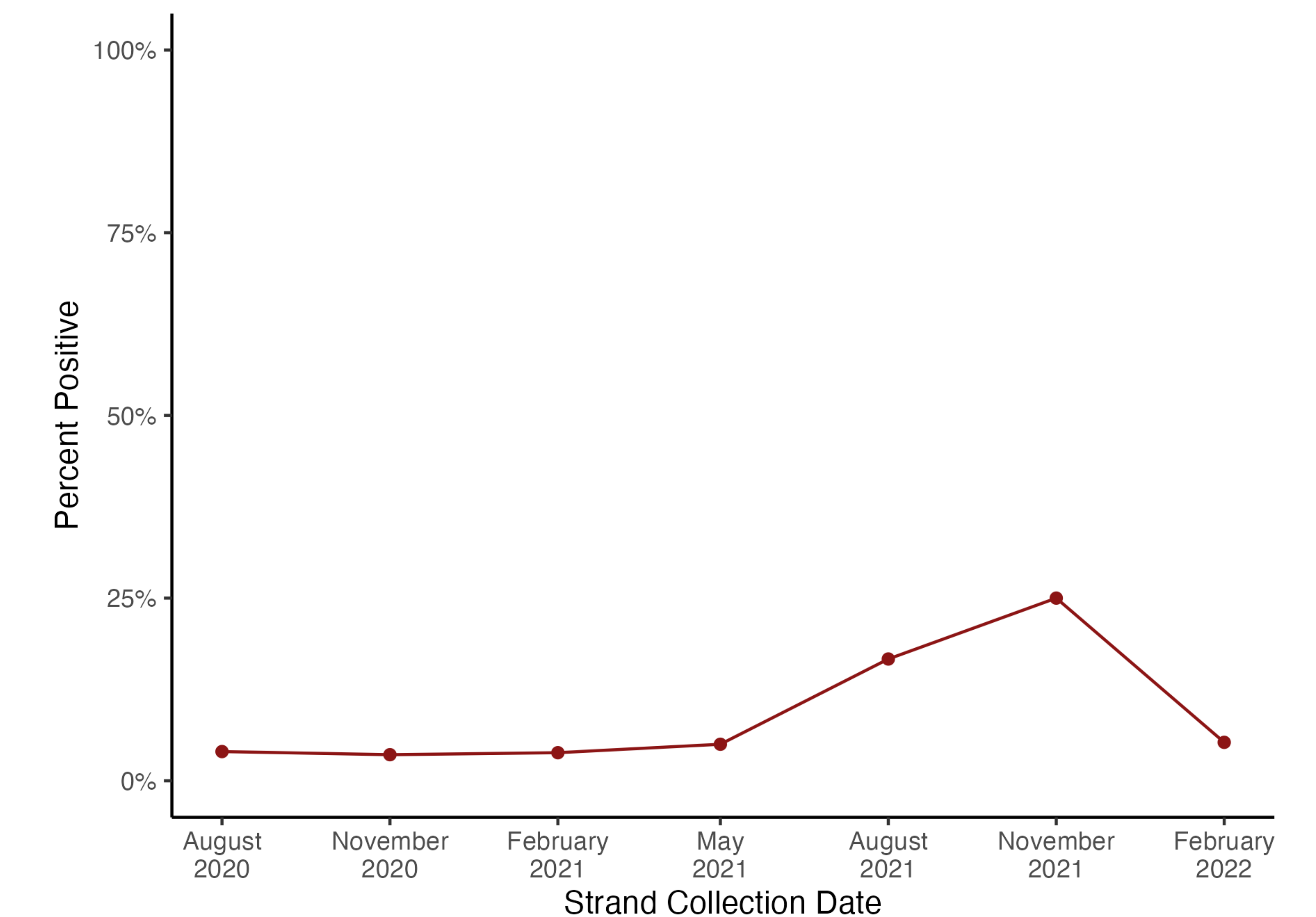


Table 1. Percent positivity of asymptomatic tests done within 2 days of admission per quarter

Asymptomatic tests/quarter	Minus strand percent positivity
August 2020-October 2020	4%
November 2020-January 2021	4%
February 2021-April 2021	4%
May 2021-July 2021	5%
August 2021-October 2021	17%
November 2021-January 2022	25%
February 2022- April 2022	5%

Figure 2. Minus strand-specific SARS-CoV-2 assay percent positivity per quarter among asymptomatic patients tested within 2 days of admission. The peak positivity in November 2021–January 2022 quarter coincided with the omicron surge in our County



Discussion

- The majority of patients with positive SARS-CoV-2 PCRs on admission were found to be non-infectious based on strand-specific rRT-PCR testing.
- Only 4-25% of asymptomatic inpatients had a detectable minus strand.
- Our findings agree with prior studies which have identified persistently positive SARS-CoV-2 PCRs without the presence of culturable live virus.
- Positive SARS-CoV-2 PCR in asymptomatic individuals often leads to delays in medical care, highlight the importance of confirming infectiousness.

Conclusion(s)

- Most asymptomatic patients tested for SARS-CoV-2 on admission were not infectious.
- Hospitals using SARS-CoV-2 PCR admission testing may need to reevaluate the continued use of this practice.

References

1. Hogan CA, Huang C, Sahoo MK, Wang H, Jiang B, Sibai M, Holubar M, Mathew R, Zehnder J, Pinsky BA. Strand-Specific Reverse Transcription PCR for Detection of Replicating SARS-CoV-2. *Emerg Infect Dis.* 2021 Feb;27(2):632-635.
2. Ridgway JP, Shah NS, Robicsek AA. Prolonged shedding of severe acute respiratory coronavirus virus 2 (SARS-CoV-2) RNA among patients with coronavirus disease 2019 (COVID-19). *Infect Control Hosp Epidemiol.* 2020 Oct;41(10):1235-1236.
3. Bullard J, Dust K, Funk D, Strong JE, Alexander D, Garnett L, Boodman C, Bello A, Hedley A, Schiffman Z, Doan K, Bastien N, Li Y, Van Caeselele PG, Poliquin G. Predicting Infectious Severe Acute Respiratory Syndrome Coronavirus 2 From Diagnostic Samples. *Clin Infect Dis.* 2020 Dec 17;71(10):2663-2666.
4. Larsen CG, Bub CD, Schaffler BC, Walden T, Intravia JM. The impact of confirmed coronavirus disease 2019 (COVID-19) infection on ambulatory procedures and associated delays in care for asymptomatic patients. *Surgery.* 2021 Jun;169(6):1340-1345.

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