CBI Primer-Bl

An online tool for designing target-specific PCR primer pairs (with internal probes) https://www.ncbi.nlm.nih.gov/tools/primer-blast/ National Center for Biotechnology Information • National Library of Medicine • National Institutes of Health • Department of Health and Human Services

Scope and Access

Primer-BLAST [1] is a PCR primer design and specificity checking tool from NCBI. It picks primers using the Primer3 algorithm [2] and then uses BLAST [3] to screen for primers specific to the input template. Similar to other BLAST searches, you can limit a Primer-BLAST search to specific taxa or a custom st of sequences specified by Entrez queries. It presents candidate primers along with their alignment to targets. Primer-BLAST is a web only application accessible through the "Specialized BLAST" section of the BLAST homepage (https://blast.ncbi.nlm.nih.gov/) or directly at https://www.ncbi.nlm.nih.gov/tools/primer-blast/.



Accepted Inputs

Accepted Inputs	Primer-BLAST		A tool for finding specific	primers	For details on "Primers
The Primer-BLAST search page		Finding primers s	pecific to your PCR template (us	sing Primer3 and BLAST).	sequences, read our
(right) defaults to single template	Primers for target on one	template Primers co	ommon for a group of sequ	ences 🔸	blogpost [5].
input form. This contains multiple	PCR Template	Retrieve recent res	ults Publication Tips for fin	iding specific primers	Save search parameters Reset page
sections. The top one (A) takes	Enter accession, gi, or FASTA	sequence (A refseq record is prefe	rred) 😮 🔹 Clear	Range 🕄	Clear
your input and allows the				From	То
adjustment to a basic set of		КВ		Forward primer	
parameters. Given a template	Or, upload FASTA file	Choose File No file chos	en		
alone (B), Primer-BLAST will find	Daiman Dammatana				
a set of primer pairs optimal for	Primer Parameters			Primer s	sequences
PCR amplification. Primer-	(5'->3' on plus strand)		Clear	C should b	be entered here
BLAST also accepts existing	(5'- >3' on minus strand)		Clear	in the 5'	to 3' orientation.
primers (C) and supports other	PCR product size	70 Max			
combinations of input: 1) a primer	# of primers to return	10			
pair with its template, 2) a		Min Opt	Max Max T _m differ	rence	
template with a single primer,	Primer melting temperatures (T _m)	57.0 60.0	63.0 3		
and 3) a pair of primers alone. In	Exon/intron selection	4	Second sector is a second sector set		
the case 1), Primer-BLAST	Exon junction span	No preference	e template input is required for opti	ons in the section 🔮	
validates the primer pair for the	Exon junction match	Min 5' match Min 3' match M	lax 3' match		
template sequence and performs		7 4	8	ale film of state of state to set as 🖉	
a specificity check if this option is	Intron inclusion	Primer pair must be separated	ases that must anneal to exons at I by at least one intron on the corre	sponding genomic DNA 😮	
selected. In the case 2), Primer-	Intron length range	Min Max	`		Clicking the guestion
BLAST finds candidate primers					mark icon next to a
that work with the input primer	Primer Pair Specificity C	Checking Parameters	G		parameter to see the
and reports their target-	Search mode	Automatic	specific to the intended PCR temp	late 😈	help information.
specificity. In case 3) with primer	Database	Refseq mRNA	•	✓ 8	
pairs alone, Primer-BLAST finds	Exclusion	Exclude predicted Refseq tran	scripts (accession with XM, XP pre	fix) Exclude uncultured/envir	ronmental sample sequences 💡
the amplification target and	Organism	Homo sapiens		Add organism	
provides primer template	Entrez query (optional)	Enter an organism name (or organ	ism group name such as enterobad	cteriaceae, rodent s), taxonomy id	or select from the suggestion list as you type. 🔮
alignments.	Primer specificity stringency	Primer must have at least 2	total mismatches to uninter	nded targets, including	
Automatic		at least 2 💌 mismatches w	ithin the last 5 💌 bps at th	ne 3' end. 😮	
Automatic	Max target amplican size	Ignore targets that have 6 🛩	or more mismatches to the p	rimer. 😮	
User guided		4000		•••••••••••••••••••••••••••••••••••••••	
No user guidance		Allow primer to amplify mRNA	splice variants (requires refseq m	quence as PCR template i	input) 🧐
With a RefSeg mRNA accession	Get Primers	🗌 🔲 Show results in a new window 🗹	Use new graphic view 😯	$\overline{\mathbf{\nabla}}$	
as a template Primer-BLAST can	+ Advanced parameters	[Defect mDNA		
take exon junctions into			Reiseq mRivA	•	
consideration through options give	n in the "Exon/int	ron	Refseq mRNA	ive generate	
selection" section (D). There you c	an set Primer-BL	AST to have	Genomes for select	ive genomes ted organisms (prim	ary reference assembly only)
candidate primers span or not spar	n splicing junction	ns. or ianore	nr	ica organisms (prin	iary reference assertiony only)
those junctions (E). You can also a	ctivate intron inc	lusion using	Refseq RNA (refsec	q_rna)	
the checkbox (F).			Custom	M	
		-			
In the Primer Specificity Checkina	Parameters secti	on (G), vou can	select different	databases	harlay
using the pull-down menu (H), rest	rict the search to	a different orga	nism by selectir	ng from the	balley
suggested list upon typing (I), adju	st the stringency	of the specificity	checking throu	igh	pariey (taxid:112009)

using the pull-down menu (H), re suggested list upon typing (I), ac parameters listed below the database (\mathbf{J}) , and check the box (**K**) to generates primer pairs that amplify all known transcript variants for the same gene. You can also adjust the search mode (L) to increase the chance of finding specific primers when the input template is highly similar to other targets in the database, and use the "Custom" database (M) option to upload a custom set of sequences (accessions or FASTA) for use as the specificity checking database.

domesticated barley (taxid:112509)

two-rowed barley (taxid:112509)

barleys (taxid:4512)

Advanced Parameters for Primer-BLAST

Clicking the "Advanced Parameters" link (A) toggles open the section with infrequently adjusted parameters. The first section (B) contains parameters for BLAST that specify the exhaustiveness of specificity checking. The second section (C) contains parameters specific to the selected primers and their PCR products (D): such as, the Tm of the PCR product, the primer length, the primer GC content, and GC clamps at the 3'-end of the primer. It also contains settings on PCR buffer conditions (E) since they can greatly affect the primer Tm calculation. Note that, in favor of search speed, Primer-BLAST does not use thermodynamic alignment features by default (F). This section also allows you to instruct Primer-BLAST to take SNPs mapped to template into consideration during primer picking (Human RefSeq accession required) by checking the checkbox (G)

required, by checking t		D	Advanced parameters		
			Primer Pair Specificity (Checking Parame	ters B
Primer Parameters			Max number of Blast target sequences	50000 🔻 🔞	
PCR Product Tm	Min Opt	Max	Blast expect (E) value	30000 🔻 🙆	
	Min Opt	Max	Blast word size	7 • 0	
Primer Size	15 20	25	Max primer pairs to screen		
	Min Max	20	Max targets to show (for	500 • 🕑	
Primer GC content (%)	20.0 80.0		designing new primers)	20	Θ
GC clamp	0 0		Max targets to show (for pre designed primers)	1000	0
Max Poly-X	5 0		Max targets per sequence	100	0
Max 3' Stability	9 0	Ļ			
Max GC in primer 3' end	5				
Secondary Structure	Use Thermodynamic Oli	go Alignment 🗆 Use	Thermodynamic Template Alignment ((warning: search F	You can pick internal probe
Alignment Methods	Primer Pair				for real-time PCR by
TH: Max Template	40.00 70.00	(For thermodynam	ic alignment model only)		activating and adjusting
Mispriming	Δην 3'	_ ` _ `	, , , , , , , , , , , , , , , , , , ,		options given in the third
TH: Max Self	45.0 35.0	(For thermodynam	ic alignment model only)		section (H). An option of
Complementarity	Any 3'](,,	,,,		Use new graphic view (I),
TH: Max Pair	45.0 35.0	(For thermodynam	ic alignment model only)		Primor BLAST to croate a
Complementarity					visually informative and
TH: Max Primer Hairpin	24.0 (For thermodyn	amic alignment mod	ei oniy)		interactive graphical
Max Template Mispriming	12 00 24 00	(For old secondary	structure alignment model only)		summary of the result using
	Any 3'		structure alignment model only		the embedded Graphical
Max Self Complementarity	8.00 3.00	(For old secondary	structure alignment model only)		Sequence Viewer [4].
Max Pair Complementarity	Any 3' 8 00 3 00	(For old second	Internal hybridization oligo		
Excluded regions			Hybridization oligo		
Overlap impetiene			hybridization ongo	Min Opt	Max
Overlap Junctions			Hyb Oligo Size	18 20	27
	5' side overlaps 3' side o	overlaps		Min Opt	Max
	/ 4 Minimal number of nucleotic	les that the left or t	Hyb Oligo tm	57.0 60.0	63.0
Concentration of monovalent	50.0		Hyb Oligo GC%	20.0 50	80.0
cations Concentration of divalent					
cations	1.5		Cat Brimana		
Concentration of dNTPs	0.6		Get primersy	Show results in a n	ew window 🖤 Use new graphic view 🥥 🤇 📘
Sall correction formula	SantaLucia 1998				
parameters	SantaLucia 1998 🔻	0	J		
Annealing Oligo	50.0 💿		\checkmark		
SNP handling G	Primer binding site may	not contain known SN	IP 😡		
Repeat filter	Automatic 🔻 🔞				
	Avoid repeat region for prim	er selection by filtering	g w ▶ NCBI/ Primer-BLAST: Maki	ing primers specific	to your PCR template. more
Low complexity filter	Avoid low complexity reg	on for primer selectio	Status	Running	K L Check Cancel
			Current time	23 June 2014, 1	6:10:54
			Time since submission	42 sec	
			Progress Message		
			Frogress wessage		

Submitting a Search

Click the "Get Primers" button (J) to submit the search. The browser tracks the progress of the submitted job via an intermediate polling page (K) and displays the result when it becomes available. You can manually check it by using the "Check" link (L).

432066

4000

Primer-BLAST Results: the Graphical Summary The Primer-BLAST displays results by breaking them into several sections: the search Search parameters and other details summary, the graphical overview, and a tabular list of primer pairs with their properties Number of Blast hits analyzed Entrez query plus alignments to the annealing sites on different targets. The summary section (A) Min total mismatches reiterates the template, an informational message with additional details on the primers Min 3' end mismatches Defined 3' end region length returned, and a "Search Summary" link (B) with detailed search statistics. Mismatch threshold to ignore targets 6 Misprimed product size deviation Max number of Blast target Primer-BLAST » Joi sequences yXwVUC3TR9qlVckWkRzFiBfYiQNTHk5DA Δ Primer-BLAST Results



For the template sequence submitted in RefSeq accession format, NM 000410 in this case, the Graphical Sequence Viewer provides much more information. Specifically, it displays:

- A clear overview of the results in the context of the target sequence, by showing the exon boundaries of the template plus its annotated protein product, pulled out from the feature table of this record (D),
- The candidate primer pairs, their predicted products, and exact locations on the template (E),
- The properties of a specific primer pair, viewable in the hovering activated popup (F),
- The sequence-level details of the annealing site through the "Zoom to Sequence" option (G) in the right-click menu
- The highlighted relationship of suggested primers with other features through the "Configure page" dialog box (H) activated by clicking the "Tracks" button, with the example shown being one of the known SNPs (I) mapped to one of the primer's annealing site on this human mRNA template.

Identity Alignment length Seq. start Seq. stop

1

2515

G

Primer-BLAST Results: Primer Pairs and Their Alignment to Targets

Detailed primer reports			The "Detailed Primer
You can re-search for specific primers by accepting some of the unintended targets, check the box(es) is search for specific primers Submit ?	next to the ones you ad	ccept and try again to re-	Reports" section (A) contains the details for returned primer pairs. Each primer
Primer pair 1 E			pair is in its own subsection
Sequence (5'->3) Template strand Length Start Stor GC% Self Forward primer TGATCATGAGAGTCGCCGTG Plus 20 195 214 59.90 55.00 6.00 Reverse primer ACAGCCAAGGTTATCCAGCC Minus 20 827 808 60.03 55.00 4.00 Product length 633 C Start Start	complementarity Se	elf 3' complementarity 00 00	(B), with a summary of basic properties along with alignments to their intended target (C) and to potentially unintended targets (D).
Product length = 633 Forward primer 1 TGATCATGAGAGTCGCCGTG 20 Template 195 214			In the example pair of
Reverse primer 1 ACAGCCAAGGTTATCCAGCC 20 Template 827 808			transcript variant 1 (NM 000410) also amplify
Products on potentially unintended templates NM_001384164.1 Homo sapiens homeostatic iron regulator (HFE), transcript variant 13, mRNA			variants 13 and 6 (under D). Alignments, which are
product length = 633 Forward primer 1 TGATCATGAGAGTCGCCGTG 20 Template 195 214			considered unintended in automatic mode. Checking
Reverse primer 1 ACAGCCAAGGTTATCCAGCC 20 Template 827 808			intended targets in re-search through Submit button (E).
> NM_139006.3 Homo sapiens homeostatic iron regulator (HFE), transcript variant 6, mRNA			
product length = 591 Forward primer 1 TGATCATGAGAGTCGCCGTG 20 Template 195 214			
	Primer Pair Sp	ecificity Checking Parar	neters
Reverse primer 1 ACAGCCAAGGTTATCCAGCC 20 Template 785	Specificity check	Enable search for primer p	pairs specific to the intended PCR template 🔞
Template 785 766	Search mode Database	User guided	F 😡
	Organism	No user guidance	~

More on "User guided" Mode and "Custom" Database NCBI/ Primer-BLAST: Finding primers specific to your PCR template (using Primer3 and BLAST).

The "User guided" (F) search mode allows you to instruct Primer-BLAST whether certain targets that are highly similar to the input template should be considered as intended target upon job submission (G).

The Custom database option (H) allows you to provide your own input dataset for specificity checking. System constraints limit the size of sequence files to 300 MB. For sequences from

the NCBI Nucleotide database, you can use their accessions or GI's to specify a larger custom dataset.

References

1. Ye J, Coulouris G, Zaretskaya I, Cutcutache I, Rozen S, Madden TL. (2012) Primer-BLAST: a tool to design targetspecific primers for polymerase chain reaction. BMC Bioinformatics, 13:134.

Primer Pair Specific	city Checking Parameters
Specificity check	Enable search for primer pairs specific to the intended PCR template
Search mode	Automatic 🔹
Database	Custom •
Organism	Refseq mRNA Refseq representative genomes
	nr bacteriace Refseg RNA (refseg rna)
Exclusion (opional)	Genome (reference assembly from selected organisms)
Entrez query (optional)	Custom J

Enter an organism name (or organism group name such as enterobacteriaceae, rodents taxonomy id or select from the suggestion list as you type.

NM 000249.3 Homo sapiens mutL homolog 1 (MLH1), transcript variant 1, mRNA

Your PCR template is highly similar to the following sequence(s) from the search database. To increase the chance

of finding specific primers, please review the list below and select all sequences (within the given sequence ranges)

PREDICTED: Homo sapiens mutL homolog 1 (MLH1), 99.8% 2520

2. Rozen, S and Skaletsky, HJ (2000) Primer3 on the WWW for general users and for biologist programmers. In: Krawetz S, Misener S (eds) Bioinformatics Methods and Protocols: Methods in Molecular Biology. Humana Press, Totowa, NJ, pp 365-386.

Input PCR template

Accession

Submit

XM 005265164.1

Range

that are intended or allowed targets Select: All None Selected:0

Title

1 - 2662

transcript variant X3, mRNA

Show results in a new window

- 3. Altschul, SF, Madden, TL, Schäffer, AA, Zhang, J, Zhang, Z, Miller, W and Lipman, DJ (1997) "Gapped BLAST and PSI-BLAST: a new generation of protein database search programs." Nucleic Acids Res. 25:3389-3402.
- The Graphical Sequence Viewer Factsheet: https://ftp.ncbi.nih.gov/pub/factsheets/Factsheet Graphical SV.pdf. 4.
- NCBI Insight Blogpost: Primer-BLAST now designs primers for a group of related sequences. https://go.usa.gov/ 5. xuJcq

Technical Assistance

Please send you feedback, questions and bug reports to blast-help@ncbi.nlm.nih.gov