Hindawi Stem Cells International Volume 2021, Article ID 5593584, 16 pages https://doi.org/10.1155/2021/5593584



# Review Article

# **Characteristics and Developments in Mesenchymal Stem Cell Therapy for COVID-19: An Update**

Lu Sang ,<sup>1,2</sup> Xiaoqin Guo ,<sup>1,2</sup> Jie Shi ,<sup>1,2</sup> Shike Hou ,<sup>1,2</sup> Haojun Fan ,<sup>1,2</sup> and Qi Lv ,<sup>1,2</sup>

<sup>1</sup>Institute of Disaster Medicine, Tianjin University, Tianjin, China <sup>2</sup>Tianjin Key Laboratory of Disaster Medicine Technology, Tianjin, China

Correspondence should be addressed to Shike Hou; housk86@163.com, Haojun Fan; fanhaojun999@126.com, and Qi Lv; lvqi68@163.com

Received 18 January 2021; Revised 23 April 2021; Accepted 30 April 2021; Published 8 June 2021

Academic Editor: Christian Morsczeck

Copyright © 2021 Lu Sang et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The outbreak of coronavirus disease 2019 (COVID-19) has so far resulted in over a hundred million people being infected. COVID-19 poses a threat to human health around the world. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has been confirmed as the pathogenic virus of COVID-19. SARS-CoV-2 belongs to the  $\beta$ -coronavirus family of viruses and is mainly transmitted through the respiratory tract. It has been proven that SARS-CoV-2 mainly targets angiotensin-converting enzyme II (ACE2) receptors on the surface of various cells in humans. The main clinical symptoms of COVID-19 include fever, cough, and severe acute respiratory distress syndrome (ARDS). Current evidence suggests that the damage caused by the virus may be closely related to the induction of cytokine storms in COVID-19. No specific drugs or measures have yet to be shown to cure COVID-19 completely. Cell-based approaches, primarily mesenchymal stem cells (MSCs), have been identified to have anti-inflammatory and immune functions in COVID-19. Clinical studies about using MSCs and its derivatives—exosomes for COVID-19 treatment—are under investigation. Here, we review the current progress of the biological characteristics, clinical manifestations, and cell-based treatment development for COVID-19. Providing up-to-date information on COVID-19 and potential MSC therapies will help highlight routes to prevent and treat the disease.

#### 1. Introduction

Coronavirus is named after the spinous proteins resembling coronae on the surface of the viral envelope. Coronaviruses are widely found in nature. They are respiratory viruses with a range of hosts, including humans, vertebrates, and invertebrates. The occurrence of coronavirus in recent years indicates that it has become one of the major threats to human health [1-6]. The structural characteristics of the coronaviruses and their numerous hosts make them more susceptible to mutations. This brings great difficulty in developing effective preventative and therapeutic options for coronaviruses and their associated diseases [7, 8]. Since December 2019, several cases of viral pneumonia were detected in Wuhan, China. The pneumonia was confirmed to be caused by a novel coronavirus. The first 41 confirmed cases of COVID-19 developed symptoms in December 2019 [8]. According to the World Health Organization (WHO), more than a hundred million cases of COVID-19 patients have been confirmed worldwide [9], including numerous infected health workers. COVID-19 is highly contagious, is susceptible to dyspnea, and presents difficulties when dealing with complex symptoms and complications [10]. In this review, we focus on the biological and clinical characteristics, pathogenic mechanisms, and the current status of treatment, with special focus on MSCs and MSC-derived exosome treatments for COVID-19. This up-to-date account may help to contribute to the further understanding of COVID-19 and to finally achieve improved treatment.

#### 2. SARS-COV-2

2.1. Discovery of SARS-CoV-2. On the 12th of January 2020, the WHO temporarily named the coronavirus novel coronavirus 2019, or "2019-nCoV," and the National Health Commission of the People's Republic of China named the

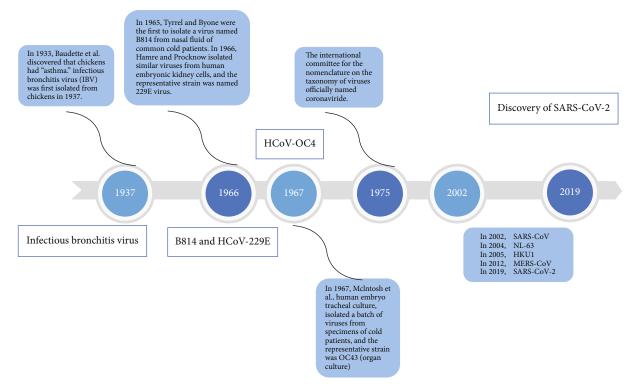


FIGURE 1: The discovery process of human coronaviruses.

disease novel coronavirus pneumonia or "NCP" on the same day. On the 11th of February 2020, the WHO formally named the disease as coronavirus disease 2019 (COVID-19). Meanwhile, the International Committee on Viral Classification formally named the coronavirus causing the disease, as coronavirus type 2 of severe acute respiratory syndrome (SARS-CoV-2) [11].

Coronaviruses are single-stranded RNA viruses with a capsule (25-31 kb) [12, 13]. They can cause respiratory, intestinal, liver, and central nervous system diseases [14]. Approximately 20% of common cold infections in humans (less common in children) are caused by coronavirus [15-17]. Since 1965, a total of 7 human coronaviruses have been identified to cause human disease. The discovered human coronaviruses include 229E, OC43, NL63, HKU1, SARS-CoV, Middle East respiratory syndrome coronavirus (MERS-CoV), and SARS-CoV-2 [14, 18, 19]. The recently discovered SARS-CoV-2 is different from SARS-CoV and is considered a novel coronavirus with prominent capacity for human infection [11]. The study on the human routes of infection and transmission of SARS-CoV-2 is important for the effective prevention and treatment of COVID-19. The discovery process of human coronaviruses is shown in Figure 1 [20–22].

2.2. Pathogenic Characteristics of SARS-CoV-2. The coronavirus family includes the  $\alpha$ ,  $\beta$ ,  $\gamma$ , and  $\delta$  subfamilies. The  $\alpha$  and  $\beta$  subfamilies mainly infect mammals, whereas the  $\gamma$  and  $\delta$  subfamilies are infectious in birds and fish [23]. The coronaviruses, including SARS-CoV-2, SARS-CoV, and MERS-CoV, are  $\beta$  species [11, 24, 25]. Negative staining electron micrographs showed that the SARS-CoV-2 particle was generally a pleomorphic sphere with a diameter of about 60–

140 nm [22]. Viruses have very distinctive protein spikes of about 9 to 12 nm that make the outer surface of the viruses look like coronae. The key spike (S) proteins bind to human ACE2 receptors and use them as entry point to infect host cells [25]. Studies showed that the similarity of the genome sequences between SARS-CoV-2 and SARS-CoV was approximately 80% [11, 21, 22]. The similarity between SARS-CoV-2 and SARS-like coronaviruses collected from bats in China (bat-SL-CoVZC45 and bat-SL-CoVZXC21) was shown to be 88% [11]. However, later sequencing of the novel coronavirus found in the United States and Italy was not consistent with the earlier sequencing of the novel coronavirus found in China. Therefore, it was proposed that it was highly likely that the novel coronavirus had mutated before February 2020 [26].

2.3. Source of Infection. Whether animals are the source of infection has been doubted since the outbreak of a novel coronavirus. SARS-CoV was transmissible through civets, while MERS-CoV was transmissible through camels [27–29]. Studies indicated that SARS-CoV-2 may also be derived from wild animals. Nevertheless, the source of infection was determined to be the seafood market in Wuhan. The natural hosts of SARS-CoV-2 are believed to be bats and/or turtles, snakes, and pangolins, but the exact source of COVID-19 lacks ample evidence [30].

The identification of "patient zero" can help to determine the source of infection, the mode of transmission, whether he/she came into contact with animals, and how he/she came into contact with those animals. The current infection source is mainly human patients that have been infected by SARS-CoV-2. Asymptomatic infected persons may also become

infection sources, as the virus has contagious effects during the incubation period [31, 32]. COVID-19 patients discharged from the hospital might show positive nucleic acid testing in the reexamination, whether these patients are still infectious remains to be determined [33]. It is very likely that the disease will persist within the global population, much like the common cold. Furthermore, since SARS-CoV-2 belongs to the plus-strand RNA virus family, it is unstable and likely to mutate, which makes it extremely difficult to prevent and control spreading.

2.4. Routes of Transmission. SARS-CoV-2 has a definite human-to-human transmission ability and is mainly transmitted by respiratory droplets and close contact [24, 25, 34–40]. Exposure to high-concentration aerosol for a long time, which is called aerosol transmission, may encourage spread in relatively closed environments [41]. As feces and urine in SARS-CoV-2 nucleic acid tests were shown to be positive for the viral RNA, it is likely to indicate the presence of live viruses in human waste, thus environmental pollution may also be a source of infection [42]. SARS-CoV-2 has the ability to replicate in conjunctival tissues. Indeed, COVID-19 was shown to cause infection through the conjunctiva and also the excreta [43, 44]. Recently, cases in China identified a mutated strain, D614G, which promotes the infectivity of SARS-CoV-2 and enhances viral transmission [45]. Effective methods to eliminate infection sources and transmission of the virus to protect vulnerable groups require urgent address.

# 3. Clinical Characteristics of COVID-19

The general population is susceptible to SARS-CoV-2, but the infection rate in children is relatively low. It remains to be further studied whether this relates to the higher proportion of lymphocytes in children [35, 46]. Respiratory failure is the main reason for aggravation of SARS-CoV-2 and SARS-CoV infections. Overall, the incidence rate is higher in men than in women and the case fatality rate is more than three times higher in women [35, 47]. Women may have a less susceptible advantage.

The incubation period for COVID-19 is generally three to seven days but, maximumly, may take up to fourteen days [48]. It has been reported that the median time from the first symptom to dyspnea is about five days and to ARDS is eight days [34]. Most patients have symptoms of fever, chills, fatigue, dry cough, and muscle pain; a few are accompanied by nasal obstruction, running nose, pharynx galgia and diarrhea, which may or may not be accompanied by pneumonia [25]. Severe and critically ill patients present low to moderate fever over the course of the disease, whereas there have been reported cases of no presentation of significant fever [25]. Severe patients usually have dyspnea and/or hypoxemia one week after onset; critically ill patients may rapidly progress to ARDS, septic shock, difficulty to correct metabolic acidosis, bleeding, and coagulation disorders [34]. The data showed that 80% of these patients were mild and 2% to 5% progressed from mild to severe [49-51]. The initial clinical course of the respiratory disease can be complicated by interstitial pneumonia, 10% to 15% of the patients evolving toward ARDS, who then require mechanical ventilation [52].

The target of SARS-CoV-2 is ACE2 receptors and cells rich in this receptor site are vulnerable to viral attack [30]. The reason that SARS-CoV-2 can spread systemically throughout the blood circulation in the body is that ACE2 are expressed in almost all endothelial cells and smooth muscle cells in organs. All tissues and organs that express ACE2 are susceptible to becoming novel coronavirus and host immune cell battlegrounds. ACE2 receptors are enriched on the surface of the type II alveolar epithelial cells (AT2) and capillary endothelial cells [53]. The intestine is the largest and most complex immune organ in the body; 70% to 80% of the immune cells are in the lymphatic tissues of the intestine. More importantly, ACE2 enzyme presents in abundance on enterocytes of the small intestine and some patients first present with gastrointestinal discomfort [53, 54]. COVID-19 also attacks the heart, lungs, kidneys, and testicles [55-58]. Additionally, facial pain and nasal obstruction are the most common symptoms and the effects related to smell and taste disorders are significantly greater in women than men [59]. Moreover, the disease is easily dangerous, progresses quickly, and can cause multiple organ failure—especially for those patients with preexisting diseases (e.g., diabetes, hypertensive, coronary heart disease, and kidney disease) [60–66]. The prognosis is good in most patients, with relatively mild symptoms in children and few patients progressing to a critical condition [46]. Of all patients, the elderly parts have a particularly poor prognosis [60, 61].

#### 4. COVID-19 Pathogenic Mechanisms

Studies have found that SARS-CoV-2, like SARS-CoV, enters into host cells through the combination of the viral spike (S) proteins with the host cell receptor ACE2 [11, 67, 68]. The virus then replicates and spreads in large numbers to trigger an immune response, attracting a large numbers of white blood cells and antibodies to clear the virus. Taken together, SARS-CoV-2-infected patients showed circulating elevated levels of proinflammatory cytokines (such as interferon-y (IFN- $\gamma$ ), interleukin- (IL-) 1 $\beta$ , IL-6, and IL-12) and chemokines (CXCL10 and CCL2), which are associated with pulmonary inflammation and extensive lung involvement [55]. COVID-19 was characterized by a diminished innate immune response, with reduced expression of genes involved in Toll-like receptors (TLRs) and interleukin signaling, chemokine binding, neutrophil degranulation, and interactions with lymphoid cells [69]. The affinity between SARS-CoV-2 S proteins and the host cell ACE2 is 10- to 20-fold greater than that of SARS-CoV, which may be the primary reason for its greater infectivity [70, 71]. Increased cohesiveness between SARS-CoV-2 S proteins and the host cell ACE2 is evident, with the binding force of SARS-CoV-2 being shown to be approximately 10–20 times higher than that of SARS-CoV, hence the greater intensity of infectivity. As shown by previous data in the literature, the receptor ACE2 is the medium for SARS-CoV-2 to enter the host cells and the serine protease TMPRSS2 is responsible for S protein priming. The S proteins present on the capsid of the virus and bind

to the ACE2 receptor on the cells following priming [72]. However, the molecular mechanisms by which SARS-CoV-2 causes the disease and how host-pathogen interactions and host immune responses occur are still relatively unknown.

Most studies speculate that the "cytokine storm" is the cause of severe disease and death. SARS-CoV-2 causes an excessive immune response in the body and induces an inflammatory storm, almost immediately after infection [73]. In the first confirmed patients with severe COVID-19, a large proportion of patients developed cytokine storm syndrome (CSS). That means many immune-active molecules can cause cytokine storm, and subsequently causes ARDS [10, 55, 74]. The representative components involved in cytokine storm development mainly include TNF- $\alpha$ , interferons, interleukins, colony-stimulating factors, and chemokines.

Expressions of inflammatory factors in mild and severe COVID-19 patients are different [54, 55, 75, 76]. The pathological results of COVID-19 patients showed that SARS-CoV-2 mainly attacked the lungs, presenting diffuse alveolar injury and lung hyaline membrane formation, which was consistent with ARDS [77]. Monocytes, especially lymphocytes, were observable in lung tissues. The general pathological features were similar to those of SARS and MERS, but the degree of fibrosis is less severe, and the inflammation is more intense. Furthermore, CD4+ and CD8+ T lymphocytes in peripheral blood were significantly reduced. However, the increased proportions of highly stimulated CCR4+CCR6+ Th17 subset cells and CD8+ T cells were found to contain high concentrations of cytotoxic particles; both revealed dysregulation of the T lymphocytes and hyperactivated status of lymphocytes [77]. This implied that the abnormal activation and imbalance of T lymphocyte subsets were the key to the process of CSS of COVID-19. Patients who have hyperinflammation suffered more risk of mortality [74]. These abnormal immune responses can lead to long-term lung damage and fibrosis in intensive care survivors and may lead to dysfunction and reduced quality of life. Data was reported that showed that 26% COVID-19 patients developed ARDS and more than 90% of nonsurvivors had developed ARDS. Furthermore, a proportion of patients may develop irreversible pulmonary fibrosis [78]. Therefore, it is important to treat this condition early and aggressively to prevent more severe infections that require therapeutic interventions, which can inhibit excessive inflammation and prevent organ damage and long-term dysfunction in severe disease cases.

### 5. Current Therapy for COVID-19

In general, COVID-19 is an acute self-limiting disease [79, 80]. A proportion of patients can be asymptomatic or self-healing after being infected with the virus and can show no symptoms of discomfort. The main clinical treatment for COVID-19 is antiviral therapy supplemented by supportive therapies such as oxygen therapy and mechanical ventilation. Once severe pneumonia or respiratory failure occurs, these comprehensive supportive therapies must be taken into account. There are no available specific treatments for reducing mortality or morbidity until now. In addition, previous

studies have reported that convalescent plasma therapy has also shown some therapeutic effect and therefore mainly can help COVID-19 and respiratory failure patients to stop viral shedding and help prolong survival but does not reduce mortality in patients with severe end-stage disease [81]. Lung transplantation is considered when traditional medicine therapy, mechanical ventilation, and extracorporeal membrane oxygenation (ECOM) cannot improve lung function [82]. Lung transplantation is not a routine treatment for COVID-19, but it is currently the only effective clinical treatment for end-stage pulmonary disease. Moreover, abnormalities in host immunity and inflammatory response are the underlying pathogenesis of SARS-CoV-2 and are associated with high mortality from COVID-19. Therefore, treating severe cases of COVID-19 requires not only suppressing virus replication but also preventing and reversing cytokine storms. Researchers found that remdesivir [83, 84], darunavir, and arbidol could inhibit SARS-CoV-2 virus in vitro [85]. Zhao et al. found that glucocorticoid can significantly reduce the toxic symptoms of pulmonary infection in severe SARS patients but it can also reduce the ability to fight infection and cause serious sequelae, such as osteonecrosis of the femoral head [86, 87]. This result suggested that glucocorticoids should be used with careful consideration. China's experience advocates that the treatment must be integrated with traditional Chinese medicine and Western medicine, giving full play to the advantages of Chinese medicine, for example, Lian-Hua Qing-Wen [88]. It is worth noting that the use of renin-angiotensin-aldosterone system (RAS) inhibitors could increase ACE2 levels, which may in turn increase the risk of COVID-19 infection. Gurwitz proposed that inhibitors of RAS, especially angiotensin receptor 1 (AT1R) blockers, may play a therapeutic role in host responses to the virus in COVID-19 patients [89]. A clinical trial (NCT04318418) from Italy is aimed at retrospectively examining whether COVID-19 patients receiving or not receiving AC-I or ARB treatment are at higher or lower risk of developing severe COVID-19 [90]. Due to the important role of inhibitors of RAS, particularly ARBs in cardiovascular and renal diseases, attention should be paid to whether patients with preexisting diseases coexisting with COVID-19 should take ACE inhibitors or angiotensin-receptor blockers [91, 92].

China has successfully developed a candidate vaccine of BBIBP-CorV with effective protection against SARS-CoV-2 and recently entered phase III clinical trials [93]. According to data available to the Chinese public, 15 COVID-19 vaccines have entered clinical trials in China, among which 6 have entered phase III clinical trials [94]. Biomedical companies including Pfizer and BioNTech also announced the completion of phase III clinical trials of the BNT162b2 vaccine. Their final data from phase III clinical trials showed that the vaccine demonstrated over 90% effectiveness. On the 11th of December 2020, the US Food and Drug Administration (FDA) approved applications from Pfizer and BioNTech for Emergency Use Authorization (EUA) for COVID-19 vaccines. The rapidly advancing mRNA vaccine, mRNA-1273 from Moderna in the United States, ended phase III clinical trials and has since entered clinical use. The vaccine was

declared to have an effectiveness of 94.5% during phase III clinical trials and was effective in reducing the likelihood of infection progressing to severe disease. The Moderna vaccine will be the second vaccine in the United States made available to help prevent COVID-19. Moreover, the Moderna vaccine induced an immune response in the elderly that was similar to that seen in younger participants, providing a new hope that the vaccine will be effective for those considered to be at high risk of severe illness due to novel coronavirus transmission.

Severe and critical cases lack specific treatments. The current principle of treatment is to treat complications of the disease, prevent secondary infection, and provide functional support to at-risk organs. Blood purification and artificial ECMO technological advancements have led to their use as lifesaving treatments for ARDS and refractory respiratory failure. Nevertheless, the potential compound immune damage associated with an extracorporeal circuit initiation during ECMO needs to be considered [95].

# 6. MSCs: A Promising Treatment

Researchers are now turning focus to the development of pathogen-specific drugs to address host immune and inflammatory abnormalities with cell therapy, to curb high mortality rates for SARS-CoV-2 and COVID-19. Host-directed therapy (HDT) advancements have provided options to both modulate the immune response and inhibit excessive inflammation. Stem cell therapy is a promising treatment that has been used as a treatment for a variety of refractory diseases involving diabetes, bone disease, and neurological diseases, and it has been used to repair and regenerate damaged or lost tissues [96–98].

MSCs are important members of the stem cell family and are derived from the mesoderm and ectoderm in early development. MSCs are pluripotent stem cells, first discovered in bone marrow in 1968 [24, 99]. They are one of the most widely used stem cells and have the advantage of an extensive source. MSCs are mainly found in the connective tissue and interstitium of organs of the body [100, 101], and they come from a variety of sources, including the umbilical cord, bone marrow, adipose tissue, and endothelial progenitor cells [99, 102]. MSCs have high amplification ability, and the gene stability remains viable after multiple passages in vitro. Moreover, MSCs have multidifferentiation potential and can repair various tissues and organs. Additionally, they can be administered to the body through various ways with low immunogenicity. The expression of the class II major histocompatibility complex (MHC-II) and costimulatory molecules is low on the MSC surface [103]. MSCs display remarkably high resistance to infection by viruses [104]. Specifically, IFN is produced only when a virus invades, which activates virus-resistant genes and recruits immune cells to fight viral infections. Pluripotent cells can continuously activate many antiviral genes without relying on IFN [104, 105]. Due to the properties of immune regulation, paracrine, directional chemotaxis of tissue damage, and the advantages of no ethical issues [106–108], MSCs can serve as a promising tool for treating COVID-19 patients.

6.1. Therapeutic Effects of MSCs on COVID-19. In many acute lung injury (ALI) and ARDS mouse models, MSC treatment can reduce lung inflammatory injury and edema, improve oxygenation, prevent the development of ARDS, and significantly prolong the survival of mice. [109] Studies have found that MSCs significantly reduced ALI associated with the influenza H5N1 virus and prolonged survival [109]. Notably, the spectrum of inflammatory cytokines induced by H5N1 and COVID-19 is similar [110]; extremely high concentrations of IL-6, GCSF, IP10, MCP-1, MIP1A, and TNF- $\alpha$  and may lead to a large degree of severe organ damage and subsequent death [34, 55]. Recent studies have shown that MSCs that secreted exosomes effectively reduced the level of detectable fibrosis through reduced collagen deposition and restored alveolar epithelial structure in bleomycininduced pulmonary fibrosis in mice and restored normal lung structure [111].

Bone marrow MSCs are widely used in cell therapy, including in a large number of preclinical studies and basic trials [112, 113]. Many clinical trials have demonstrated the safety and efficacy of MSCs. Recent studies reported on multiple clinical trials that used stem cell therapy for COVID-19. A 70-year-old critically severe female COVID-19 patient from Kunming, China, was reported to have a significant improvement in lung function and reduction in clinical symptoms after bone marrow MSC infusion [114]. More than sixty clinical trials have commenced to investigate the use of MSCs for COVID-19 treatment. Seven clinical trials, including from the United States, China, Pakistan, Turkey, Japan, and Indonesia, have recently concluded and are marked "completed" on clinicaltrials.gov. [115] (Table 1). The registered trials have different designs, especially with dose administration and primary schedules, which suggest that there is a lack of global consensus for MSC therapy. Wang and his team successfully completed phase II clinical trials, which included one hundred patients [116]. Notably, they showed no serious infusion-associated adverse events that were observable during UC-MSC intravenous infusion [117]. Outcomes of this clinical trial showed that stem cells inhibited lung inflammation, attenuated abnormal immune activation, and reduced lung injury. Stem cell therapy accelerated the shortening of the course of the disease, and severe patients did not progress to the critical stage. A study by Leng et al. demonstrated that MSC transplantation was safe and effective in patients with COVID-19 with pneumonia (two mild, four severe, and one critical), especially for critically ill patients [49].

6.2. Potential Mechanisms of MSCs in Treatment of COVID-19. The underlying mechanism of improvement after infusion of MSCs in COVID-19 patients also showed the strong anti-inflammatory activity of MSCs. It is clear that the results showed effectiveness, including increased peripheral blood lymphocyte count, reduction of C-reactive protein, and immune cell (CXCR3 + NK cells, CXCR3 + CD4 + T cells, and CXCR3 + CD8 + T cells) secretion of overactive cytokines into the blood [49]. Meanwhile, in the MSCs group, IL-10 levels were higher and TNF- $\alpha$  were significantly lower [49]. In addition, the gene expression profile showed that

TABLE 1: Clinical trials investigated the use of MSCs for COVID-19 treatment market	d "completed" on clinicaltrials.go	ov.
---	------------------------------------	-----

Trial ID no.	Status	Responsible country	Study title	Interventions
NCT04288102	Completed	China	Treatment with human umbilical cord-derived mesenchymal stem cells for severe corona virus disease 2019 (COVID-19)	UC-MSCs
NCT04355728	Completed	United States	Use of UC-MSCs for COVID-19 patients	UC-MSCs
NCT04492501	Completed	Pakistan	Investigational treatments for COVID-19 in tertiary care hospital of Pakistan	MSCs
NCT04522986	Completed	Japan	An exploratory study of ADR-001 in patients with severe pneumonia caused by SARS-CoV-2 infection (COVID-19)	AD-MSCs
NCT04535856	Completed	Indonesia	Therapeutic study to evaluate the safety and efficacy of DW-MSC in COVID-19 patients (DW-MSC)	MSCs
NCT04573270	Completed	United States	Mesenchymal stem cells for the treatment of COVID-19	UC-MSCs
NCT04713878	Completed	Turkey	Mesenchymal stem cells the rapy in patients with COVID-19 pneumonia $$	MSCs

UC-MSCs: human umbilical cord mesenchymal stem cells; AD-MSCs: adipose-derived mesenchymal stem cells.

MSCs were ACE2 and TMPRSS2, indicating that MSCs had no risk of COVID-19 infection and further characterizing MSC expression profiles [49]. Notably, studies also showed that after SARS-CoV-2 infection, even in older patients, MSCs given intravenously were capable of regulating the pulmonary microenvironment and had immunomodulatory activity, thereby promoting tissue repair while dampening the immune system [49]. Intravenous injection of bone marrow MSCs usually causes them to accumulate in the lungs and increased secreted paracrine factors within the lung tissue [118]. The secreted paracrine factors contributed significantly to the regeneration of alveolar epithelial cells, the resolution of fibrosis, and the restoration of lung function [65].

6.3. Immunomodulatory Functions of MSCs. Abnormal activation of innate immunity and adaptive immunity was found in human COVID-19, wherein uncontrolled inflammatory responses led to local and systemic tissue damage. In the human body, ACE2 is expressed by both monocytes and macrophages. Therefore, these cells can be infected by SARS-CoV and SARS-CoV-2 [119], leading to the activation and transcription of proinflammatory genes [120]. Patients with COVID-19 exhibited activated phenotypic morphology (FSC-high) and production capacity for IL-6, IL-10, and TNF- $\alpha$ , despite normal monocyte blood counts [121]. Moreover, in bronchoalveolar lavage fluid, proinflammatory macrophages were higher in severe COVID-19 patients, compared to mild patients, which contributed to a cytokine storm [122]. Elevated levels of neutrophils or macrophages may help predict disease severity and clinical outcome [123]. Meanwhile, COVID-19 can also affect the adaptive immune system. Reduction in lymphocytes is a common feature, especially in those patients with severe and critically ill COVID-19. Clinical studies have shown that a decrease in the frequency of circulating lymphocytes, including CD4+ and CD8+ T cells, is strongly associated with the severity of COVID-19 [124]. The negative relationship between proinflammatory cytokines and circulating T cells is likely related to the redistribution of cells in the tissue and/or induction of cell apoptosis [125]. However, activated Th1 cells, together with Th17 cells could directly stimulate monocytes to secret proinflammatory cytokines and accelerate cytokine storm [119, 126]. Therefore, impaired and unbalanced innate and adaptive immune responses are involved in the process of COVID-19.

MSCs have the potential to exert immunomodulatory functions in both innate and adaptive immune responses, including direct and indirect interactions with immune cells [106]. Early studies have demonstrated that MSCs can promote the polarization of monocytes/macrophages toward an anti-inflammatory type 2 subtype, which is characterized by high levels of IL-10 and decreased IL-12 and TNF- $\alpha$  production [127, 128]. Moreover, MSCs affected the subsequent activation of antigen-specific CD4+ T cells by inhibiting the expression of MHC class II and CD86 on macrophages. MSC-derived paracrine factors, for example, prostaglandin E2 (PGE2), may be associated with anti-inflammatory and immunosuppressive functions of MSCs [129, 130]. In addition, transforming growth factor- $\beta$  (TGF- $\beta$ ) is also closely related to the immunosuppressive effects of MSCs on natural killer (NK) cell proliferation and proinflammatory cytokines produced by NK cells [131]. T cells are an important part of the adaptive immune system and play a key role in the body's specific immune response. The abnormal activation of T lymphocytes is an important cause of CSS and ARDS in COVID-19. More than 30 soluble factors, such as TGF- $\beta$ , PGE2, indoleamine 2,3-dioxygenase (IDO), and hepatocyte growth factor (HGF) have been demonstrated to exert the immunomodulatory capacity of MSCs by inhibiting CD4+ Th1 and Th17 cells and CD8+ T cell proliferation and inducing Foxp3+ Treg differentiation [132, 133]. In addition, MSCs suppressed IFN-γ released by Th1 and increased IL-4 production from Th2 [134]. Therefore, MSCs can reduce the secretion of inflammatory factors involved in CSS and increase the secretion of anti-inflammatory factors by regulating the innate and adaptive immunity, which can help treat and reduce the mortality of COVID-19 patients [49].

6.4. Reparative Functions of MSCs. MSC influences over the repair of tissue defects have been extensively reported [135–137]; MSCs are adult stem cells which can keep a proliferating state for long periods. Additionally, they remain undifferentiated and differentiate into a variety of cell types including

cardiomyocytes, hepatocytes, neurons, and astrocytes [138–140]. It has been reported that the immunomodulatory properties of MSCs are one of the main factors in the process of lung repair and regeneration in pathologic conditions such as ALI, chronic obstructive pulmonary disease, bronchopulmonary dysplasia, and idiopathic pulmonary fibrosis [99, 102, 141]. COVID-19 mainly causes lung injury, which is manifested by damage to alveolar epithelial cells and capillary endothelial cells, as well as diffuse pulmonary interstitial and alveolar edema, leading to acute hypoxic respiratory insufficiency and eventually ARDS.

MSCs can affect P13, MAPK, NF-κB, and other signaling pathways through potential differentiation ability, the production of cytokines, and the secretion of a large amount of exosomes and vesicles containing microRNA (miRNAs) to repair lung injury [142]. MSCs partially gathered in the injured lung after intravenous injection can differentiate into alveolar epithelial cells and pulmonary vascular endothelial cells, but the efficiency to do so was reported to be extremely low [143]. Moreover, evidence suggests that MSCs express a variety of growth factors and are involved in the regulation of cell proliferation, apoptosis, and differentiation. MSCs can secrete cell nutrition factors (such as keratinocyte growth factor (KGF), HGF, Ang-1, and granulocyte macrophage colony stimulating factor (GM-CSF), while reducing lung tissue expression of TGF- $\beta$ , TNF- $\alpha$ , type I collagen, and type III collagen [144, 145]. It has been proven that MSC therapy can secrete extracellular vesicles enriched with HGF, which protected or restored alveolar epithelium and lung endothelial cells and reduced the inflammatory response and increased autophagy [146-149]. MSC therapy have the ability of regeneration through activation of the WNT/catenin signaling pathway, which has also been shown to promote the direct differentiation of MSCs into type II alveolar epithelial cells [150-153]. Moreover, MSCs protected alveolar epithelial cells from inflammatory and oxidative stress damage by secreting IL-1, Ang-1, PGE2, and HGF or by scavenging oxidants and free radicals [154-157]. MSCs were also shown to regulate tissue remodeling processes and attenuated lung fibrosis by increasing metalloproteinase- (MMP-) 8 and decreasing the levels of tissue inhibitors of metalloproteinase- (TIMP-) 1, IL-1 $\beta$ , and TGF- $\beta$ 1 in animal models of ARDS [158, 159].

Vascular endothelial growth factor (VEGF), epidermal growth factor (EGF), and HGF are factors with important implications in epithelial maturation, regeneration, and repair of the alveolar epithelial barrier during the recovery process of ARDS lung injury [160–162]. Meanwhile, MSCs accumulated in the lungs after intravenous infusion, which improved the tissue microenvironment to one more conducive for resident lung cells, proangiogenic cells, and proregenerative cells [136, 137, 163, 164]. Bone marrow MSCs accelerate healing through regulatory mechanisms that affect immunoregulation, apoptosis, and angiogenesis, while supporting the recruitment, growth, and differentiation of local stem cells and progenitor cells.

As ARDS worsens, multiple organ failure may occur, leading to an increase in morbidity and mortality. MSC administration can reduce the histopathological impairment

of lung tissues and promote functional recovery in ARDS models [153, 159, 165]. MSCs also improved the repair and functional recovery in other distal organs, including the heart [166], liver [159, 167], kidney [159, 168], and gut [169, 170]. Therefore, MSC administration can not only improve the recovery of lung function but also delay or inhibit the development of ARDS to multiorgan injury.

# 7. Prospects

The anti-inflammatory effects of MSCs were mainly attributed to the release of paracrine factors, despite the poor survivability of donor-derived MSCs in the host lung tissue after intravenous treatment or lung engraftment [163, 171]. Nevertheless, there are many hurdles to overcome before it can be applied to clinical trials, including the heterogeneity of cells, large-scale production, and harsh storage conditions; it also highly possibly causes tumor formation after ectopic engraftment [172]. So its use is restricted by security consideration. Recent studies showed that certain therapeutic effects of MSCs depend on their ability to secrete extracellular vesicles (EVs). Extracellular vesicles are secreted by nearly all mammalian cell types with a diameter of approximately 20-2000 nm [173]. The extracellular vesicles were identified as unneeded compounds of cells initially [174]. To date, we know that extracellular vesicles are responsible for the component exchange and communication between cells instead of just waste carriers and act as signaling vehicles to recipient or as a consequence of pathological developments [175–177]. According to the recent statement from the International Society for Extracellular Vesicles (ISEV) in 2018, EVs are generally classified into exosomes, microvesicles (MVs), and apoptotic bodies (ABs) according to their sizes, surface markers, and cellular generation mechanism [178, 179] (Table 2). EVs can transfer mRNA, microRNA, proteins, lipids, and even organelles such as mitochondria to target cells and tissues, change gene expression, and regulate target cell behavior to reduce the inflammatory response, thus mediating and mimicking therapeutic roles of their parental MSCs (Figure 2) [180–182]. Moreover, there is accumulating evidence indicating that MSC-EVs have shown equal or even better treatment efficacies than MSCs in many diseases. EVs can be considered as potent reservoirs of bioactive substances within the MSC secretome [183, 184].

Extracellular vesicles are released by various cell types, including epithelial cells, tumor cells, macrophages, and MSCs, which are believed to play a unique role in intercellular communication. MSC-EVs are composed of soluble proteins, including a variety of cytokines, chemokines, and growth factors [185]. Once released, EVs and soluble proteins interact with the target cell and regulate the cellular response. MSC-EVs, unlike monoclonal antibodies, can act simultaneously and possibly synergistically on many cytokines. Other than that, the functions of EVs mainly are identified to depend on the cell origin. EVs can activate endogenous stem cells, progenitor cells, and genetic material transfer; inhibit cell apoptosis; regulate inflammatory response; stimulate extracellular matrix remodeling, angiogenesis, and reduce fibrosis; and mediate chemotaxis [186].

TABLE 2: The characteristics and categories of EVs.	
---	--

Characteristic	Exosomes	Microvesicles	Apoptotic bodies
Apoptotic bodies	50-150	100-1000	500-2000
Origin	Multivesicular body exocytosis	Cell membrane budding and fission	Plasma membrane, endoplasmic reticulum
Morphology	Cup/round shaped	Various shapes	Heterogeneous
Sucrose gradient	1.13–1.19 g/mL	$1.04-1.07\mathrm{g/mL}$	1.16-1.28 g/mL
Surface markers	Annexins, tetraspanins, heat-shock proteins	CD40, cholesterol, sphingomyelin, ceramide	Annexin V positivity, TSP, C3b
Contents	Proteins, nucleic acids, lipid	Proteins, nucleic acids, lipid	Nuclear fractions, DNA, cell organelles
Isolation technique	Centrifugation at 100,000 gravity	Ultracentrifugation	Ultracentrifugation

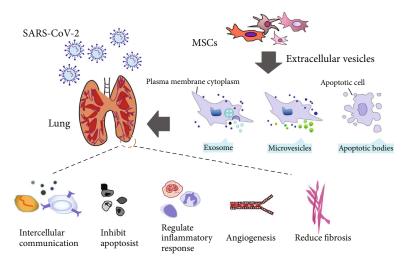


FIGURE 2: SARS-CoV-2 enters into the human lung and the MSCs-EVs are considered a promising treatment due to their effects.

Moreover, EVs are generally considered safer than cell therapy because they lack the capacity to induce endogenous tumor formation, while possessing low immunogenicity. In addition, EVs are easier to manipulate and store than cells and, thus, cost less. More importantly, EVs can take advantage of multitargeted therapy, which is far superior to monotherapy, especially in instances of COVID-19 multiorgan dysfunction [187]. Therefore, MSC-EVs are a promising tool for cell-free therapy of pulmonary diseases and may be more suitable for human use as COVID-19 therapy.

Preclinical trials have proven that MSC-derived EVs can be used as acellular substitutes for ARDS therapy [182]. Indeed, MSC-EV infusion can reduce proinflammatory factors and the subsequent cytokine storm that drives ARDS progression. Meanwhile, there is also a corresponding increase of anti-inflammatory signaling mediators, which reduce the severity of lung injury by increasing the permeability and function of the alveolar epithelium [182]. Furthermore, virus reproduction could be inhibited by MSC-derived EVs directly [188, 189]. A key intrinsic component of EVs is miRNA, which is closely associated with physiological processes such as development and immunoregulation through epigenetic alterations [190]. Packaged miRNAs in EVs influence the differentiation and function of multiple types of

cells, and excessive levels are associated with a variety of diseases, including cancer, lung diseases, obesity, diabetes, and cardiovascular disease [191–196].

Studies demonstrated that miRNA derived from MSC EVs could alter epigenetic activity, leading to changes in cell receptor expression and the subsequent exchange of genetic material, which also helped prevent RNA viruses, including coronaviruses from cell entry [188]. In a swine influenza model, infection and virus shredding were significantly inhibited following the administration of MSC-derived EVs [197]. Studies have utilized different techniques to track the biological distribution of EVs throughout the body in different animal models. EV tracking allowed researchers to show that in rat models of ALI, intravenously injected EVs reached lung tissues [198]. Numerous studies have shown that EVs from various sources could restore lung injury, improve respiratory function, and, in some cases, improve survival. EV-based therapies for COVID-19-related lung injury have great potential because they target multiple pathways implicated in COVID-19 insult while enhancing tissue regeneration.

As evidenced by an increase in the number of EV studies, recently published, EV therapy remains a popular research field and continues to receive a high amount of attention.

TABLE 3: MSC-EV treatment trials in COVID-	19 registered on Clinical Trials. The treatment of COVID-19 disease, including trial numbers,
source of EVs, routes of administration, and p	orimary endpoints.

Trial ID no.	Status	Responsible country	Source of extracellular vesicles	Administration	Primary endpoints
NCT04276987	Completed	China	Exosome derived from allogenic adipose mesenchymal stem cells	Inhalation	Adverse reaction
NCT04491240	Completed has results	Russian Federation	MSC exosomes	Inhalation	Adverse events
NCT04493242	Not recruiting	United States	Bone marrow-derived extracellular vesicles	IV	All-cause mortality
NCT04602442	Enrolling by invitation	Russian Federation	Mesenchymal stem cell exosomes	Inhalation	Adverse events during trial
NCT04657458	Available	United States	Bone marrow-derived extracellular vesicles	IV	Not recorded
NCT04798716	Not recruiting	<b>United States</b>	MSC exosomes	IV	Adverse events

The literature reported that the clinical efficacy and safety of MSC products in preclinical models of pulmonary diseases and the indication that noninvasive methods can be utilized (e.g., inhalation) offer novel perspectives [199]. A previous study by Dinh et al. demonstrated that inhalation of globular cell secreted exosomes in mice treated with different lung injury patterns and promoted pulmonary fibrosis repair [111]. Table 3 lists the clinical trials applied for COVID-19: MSC-derived extracellular vesicle treatment for COVID-19 registered on clinicaltrials.gov [200]. To be specific, clinical trials (NCT04276987 and NCT04491240) registered by China and Russia are using aerosol inhalation of MSCderived exosomes to treat COVID-19 and have been marked as "completed" on clinicaltrials.gov. NCT04491240 has reported some results. The researchers found that the MSC exosome treating reduced the C-reactive protein (CRP) and lactic acid dehydrogenase (LDH) level in serum and no adverse reactions, such as allergy and bronchospasm, were found during the test. A nonrandomized open-label cohort study reported that born marrow MSC-derived exosomes could improve oxygenation, reduce neutrophil count, elevate average CD3+, CD4+, and CD8+ lymphocyte counts, and reduce the levels of acute-phase reactants in severe COVID-19 patients [201]. These findings demonstrated the potential of MSC-derived exosomes for the treatment of COVID-19 patients. However, even though the generic term extracellular vesicles are currently used to refer to these secreted membrane vesicles, they are in fact highly heterogeneous. Before converting EV therapy into human clinical applications, efforts should be made to address many issues such as the optimal dose and route of administration in animal models. Although there are many obstacles and drawbacks that need to be addressed, inhaled drug administration activity in these studies could provide a way to treat lung injury in COVID-19 pneumonia.

MSCs remain the most relevant stem cell technology for a wide range of diseases. On the other hand, delivery via EVs offers an opportunity to avoid the use of cell therapy. Although this requires further research, it may help ensure the safety of MSC treatment. MSCs and their secretome are being used to study a large number of diseases involving skin pathology, cardiovascular disease, neuropathology, meta-

bolic disorders, spinal cord injury, and autoimmune diseases [202]. We are encouraged that other sources of MSCs are gradually expanding including the embryonic stem cells (ESCs) and induced pluripotent stem cell- (iPSC-) derived MSCs, ESC-MSCs, and iPSC-MSCs [202]. These have been shown to overcome age-related problems and limited proliferation rates of adult MSCs. In addition, iPSC-MSCs showed signs of rejuvenation. Although many clinical trials have been conducted and clinical trial details can be found at ClinicalTrials.gov, there are fewer and less consistent clinical trials to complete, so we need to continue to pay attention to this aspect [203].

The current epidemic has entered a severe and complex period. By April 2021, more than a hundred million people worldwide have been diagnosed with COVID-19, with more severe cases being reported daily. For severely ill COVID-19 patients, it is more important to treat CSS, ARDS, and ALI function in addition to routine antiviral therapy, to reduce the mortality rate. We believe that both MSCs and secreted exosomes have great potential in repairing lung injury. Therefore, they have the potential to serve as promising treatment options for severe COVID-19.

#### 8. Conclusion

COVID-19 is at a critical stage wherein control is paramount, but there are still no targeted or highly effective treatment options for patients with severe COVID-19 or those that are clear from infection but must live with lung complications and other consequences of the disease. MSC therapy has played an important role in clinical trials. Although MSC therapy has many difficulties to overcome before reaching its full therapeutic potential (source, heterogeneity, and quality control), its derivative EVs have attracted attention and have been tested in COVID-19 clinical trials with promising results. However, current data is limited and a better understanding of stem cell therapy and EV therapy and their safety development into effective preclinical endpoints is needed to recognize and address these therapeutics and their progress toward truly meeting the clinical needs of COVID-19.

# **Data Availability**

The processed data are available from the corresponding author upon request.

# **Conflicts of Interest**

The authors indicated no potential conflicts of interest.

#### **Authors' Contributions**

Lu Sang, Xiaoqin Guo, and Qi Lv did the conception and design and manuscript writing; Shi Jie, Qi Lv, Haojun Fan, Shike Hou did the manuscript review, writing, and final approval of the manuscript. Lu Sang and Xiaoqin Guo contributed equally to this work. Lu Sang and Xiaoqin Guo are co- first author.

# Acknowledgments

This study was supported by funding from the National Natural Science Foundation of China, no. 81971878; Opening Project of Military Logistics, no. BLB19J006; Tianjin Natural Science Foundation, no. 20JCQNJC01260; and Tianjin University Independent Innovation Fund, no. 2020XRG-0027 and no. 2020XYF-0041.

#### References

- [1] X. Y. Ge, J. L. Li, X. L. Yang et al., "Isolation and characterization of a bat SARS-like coronavirus that uses the ACE2 receptor," *Nature*, vol. 503, no. 7477, pp. 535–538, 2013.
- [2] B. L. Haagmans, S. H. S. al Dhahiry, C. B. E. M. Reusken et al., "Middle East respiratory syndrome coronavirus in dromedary camels: an outbreak investigation," *The Lancet Infectious Diseases*, vol. 14, no. 2, pp. 140–145, 2014.
- [3] B. Hu, L. P. Zeng, X. L. Yang et al., "Discovery of a rich gene pool of bat SARS-related coronaviruses provides new insights into the origin of SARS coronavirus," *PLoS Pathogens*, vol. 13, no. 11, article e1006698, 2017.
- [4] V. D. Menachery, B. L. Yount Jr., K. Debbink et al., "A SARS-like cluster of circulating bat coronaviruses shows potential for human emergence," *Nature Medicine*, vol. 21, no. 12, pp. 1508–1513, 2015.
- [5] V. D. Menachery, B. L. Yount Jr., A. C. Sims et al., "SARS-like WIV1-CoV poised for human emergence," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 113, no. 11, pp. 3048–3053, 2016.
- [6] J. S. M. Sabir, T. T. Y. Lam, M. M. M. Ahmed et al., "Co-circulation of three camel coronavirus species and recombination of MERS-CoVs in Saudi Arabia," *Science*, vol. 351, no. 6268, pp. 81–84, 2016.
- [7] S. R. Weiss and S. Navas-Martin, "Coronavirus pathogenesis and the emerging pathogen severe acute respiratory syndrome coronavirus," *Microbiology and Molecular Biology Reviews*, vol. 69, no. 4, pp. 635–664, 2005.
- [8] Anon, "Emerging understandings of 2019-nCoV," *The Lancet*, vol. 395, no. 10221, p. 311, 2020.
- [9] World Health Organization, "Coronavirus Disease (COVID-2019) Situation Reports," 2021, https://covid19.who.int/ table/.

[10] Y. R. Guo, Q. D. Cao, Z. S. Hong et al., "The origin, transmission and clinical therapies on coronavirus disease 2019 (COVID-19) outbreak-an update on the status," *Military Medical Research*, vol. 7, no. 1, p. 11, 2020.

- [11] R. Lu, X. Zhao, J. Li et al., "Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding," *The Lancet*, vol. 395, no. 10224, pp. 565–574, 2020.
- [12] S. R. Weiss and J. L. Leibowitz, "Coronavirus pathogenesis," *Advances in Virus Research*, vol. 81, pp. 85–164, 2011.
- [13] P. S. Masters and S. Perlman, "Coronaviridae," in *Fields Virology*, D. M. Knipe and P. M. Howley, Eds., pp. 825–858, Lippincott Williams & Wilkins, 2013.
- [14] Z. W. Ye, S. F. Yuan, K. S. Yuen, S. Y. Fung, C. P. Chan, and D. Y. Jin, "Zoonotic origins of human coronaviruses," *International Journal of Biological Sciences*, vol. 16, no. 10, pp. 1686–1697, 2020.
- [15] H. Lvd, "Human coronaviruses, what do they cause?," *Antiviral Therapy*, vol. 12, no. 4 Part B, pp. 651–658, 2007.
- [16] K. McIntosh, J. H. Dees, W. B. Becker, A. Z. Kapikian, and R. M. Chanock, "Recovery in tracheal organ cultures of novel viruses from patients with respiratory disease," *Proceedings of* the National Academy of Sciences of the United States of America, vol. 57, no. 4, pp. 933–940, 1967.
- [17] H. Dorothy and J. P. John, "A new virus isolated from the human respiratory tract," *Proceedings of the Society for Experimental Biology and Medicine*, vol. 121, no. 1, pp. 190–193, 1966
- [18] S. Su, G. Wong, W. Shi et al., "Epidemiology, genetic recombination, and pathogenesis of coronaviruses," *Trends in Microbiology*, vol. 24, no. 6, pp. 490–502, 2016.
- [19] D. Forni, R. Cagliani, M. Clerici, and M. Sironi, "Molecular evolution of human coronavirus genomes," *Trends in Microbiology*, vol. 25, no. 1, pp. 35–48, 2017.
- [20] K. McIntosh, A. Z. Kapikian, K. A. Hardison, J. W. Hartley, and R. M. Chanock, "Antigenic relationships among the coronaviruses of man and between human and animal coronaviruses," *Journal of Immunology*, vol. 102, no. 5, pp. 1109– 1118, 1969.
- [21] M. M. C. Lai and D. Cavanagh, "The molecular biology of coronaviruses," *Advances in Virus Research*, vol. 48, pp. 1– 100, 1997.
- [22] J. Cui, F. Li, and Z. L. Shi, "Origin and evolution of pathogenic coronaviruses," *Nature Reviews. Microbiology*, vol. 17, no. 3, pp. 181–192, 2019.
- [23] P. C. Woo, S. K. Lau, C. S. Lam et al., "Discovery of seven novel mammalian and avian coronaviruses in the genus delta coronavirus supports bat coronaviruses as the gene source of alphacoronavirus and beta coronavirus and avian coronaviruses as the gene source of gamma coronavirus and delta coronavirus," *Journal of Virology*, vol. 86, no. 7, pp. 3995–4008, 2012.
- [24] C. M. Coleman and M. B. Frieman, "Coronaviruses: important emerging human pathogens," *Journal of Virology*, vol. 88, no. 10, pp. 5209–5212, 2014.
- [25] N. Zhu, D. Zhang, W. Wang et al., "A novel coronavirus from patients with pneumonia in China, 2019," *The New England Journal of Medicine*, vol. 382, no. 8, pp. 727–733, 2020.
- [26] W. B. Yu, G. D. Tang, L. Zhang, and R. T. Corlett, "Decoding the evolution and transmissions of the novel pneumonia coronavirus (SARS-CoV-2) using whole genomic data," *Zoological Research*, vol. 41, no. 3, pp. 247–257, 2020.

[27] Y. Guan, B. J. Zheng, Y. Q. He et al., "Isolation and characterization of viruses related to the SARS coronavirus from animals in southern China," *Science*, vol. 302, no. 5643, pp. 276–278, 2003.

- [28] A. N. Alagaili, T. Briese, N. Mishra et al., "Middle East respiratory syndrome coronavirus infection in dromedary camels in Saudi Arabia," *mBio*, vol. 5, no. 2, p. e00884, 2014.
- [29] M. G. Hemida, R. A. Perera, P. Wang et al., "Middle East Respiratory Syndrome (MERS) coronavirus seroprevalence in domestic livestock in Saudi Arabia 2010 to 2013," *Eurosur-veillance*, vol. 18, no. 50, article 20659, 2013.
- [30] P. Zhou, X.-L. Yang, X.-G. Wang et al., "A pneumonia outbreak associated with a new coronavirus of probable bat origin," *Nature*, vol. 579, no. 7798, pp. 270–273, 2020.
- [31] Z. Hu, C. Song, C. Xu et al., "Clinical characteristics of 24 asymptomatic infections with COVID-19 screened among close contacts in Nanjing, China," *Science China. Life Sciences*, vol. 63, no. 5, pp. 706–711, 2020.
- [32] S. A. Lauer, K. H. Grantz, Q. Bi et al., "The incubation period of coronavirus disease 2019 (COVID-19) from publicly reported confirmed cases: estimation and application," *Annals of Internal Medicine*, vol. 172, no. 9, pp. 577–582, 2020
- [33] L. Zhou, K. Liu, and H. G. Liu, "Cause analysis and treatment strategies of "recurrence" with novel coronavirus pneumonia (COVID-19) patients after discharge from hospital," *Zhon-ghua Jie He Hu Xi Za Zhi*, vol. 43, no. 4, pp. 281–284, 2020.
- [34] D. Wang, B. Hu, C. Hu et al., "Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China," *JAMA*, vol. 323, no. 11, pp. 1061–1069, 2020.
- [35] Q. Li, X. Guan, P. Wu et al., "Early transmission dynamics in Wuhan, China, of novel coronavirus-infected pneumonia," *The New England Journal of Medicine*, vol. 382, no. 13, pp. 1199–1207, 2020.
- [36] L. E. Gralinski and V. D. Menachery, "Return of the coronavirus: 2019-nCoV," Viruses, vol. 12, no. 2, p. 135, 2020.
- [37] A. Banerjee, K. Kulcsar, V. Misra, M. Frieman, and K. Mossman, "Bats and coronaviruses," *Viruses*, vol. 11, no. 1, p. 41, 2019.
- [38] C. C. Lai, T. P. Shih, W. C. Ko, H. J. Tang, and P. R. Hsueh, "Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and coronavirus disease-2019 (COVID-19): the epidemic and the challenges," *International journal of antimicrobial agents*, vol. 55, no. 3, article 105924, 2020
- [39] L. Zhang and Y. Liu, "Potential interventions for novel coronavirus in China: a systematic review," *Journal of Medical Virology*, vol. 92, no. 5, pp. 449–479, 2020.
- [40] World Health Organization, "Coronavirus disease (COVID-2019) situation reports," 2020, https://www.who.int/emergencies/diseases/novel-coronavirus/.
- [41] E. L. Anderson, P. Turnham, R. G. John, and C. C. Clarke, "Consideration of the aerosol transmission for COVID-19 and public health," *Risk Analysis*, vol. 40, no. 5, pp. 902– 907, 2020.
- [42] Y. Kunz, W. Horninger, and G. M. Pinggera, "Are urologists in trouble with SARS-CoV-2? Reflections and recommendations for specific interventions," *BJU International*, vol. 126, no. 6, pp. 670–678, 2020.

[43] C. W. Lu, X. F. Liu, and Z. F. Jia, "2019-nCoV transmission through the ocular surface must not be ignored," *The Lancet*, vol. 395, no. 10224, article e39, 2020.

- [44] X. Ma, J. Lin, and S. Fang, "Precautions in ophthalmic practice in a hospital with the risk of COVID-19: experience from China," *Acta ophthalmologica*, vol. 98, no. 4, pp. e520–e521, 2020.
- [45] J. Hu, C. L. He, and Q. Z. Gao, "The D614G mutation of SARS-CoV-2 spike protein enhances viral infectivityand decreases neutralization sensitivity to individual convalescent sera," *bioRxiv*, no. article 161323, 2020.
- [46] J. F. Ludvigsson, "Systematic review of COVID-19 in children shows milder cases and a better prognosis than adults," *Acta Paediatrica*, vol. 109, no. 6, pp. 1088–1095, 2020.
- [47] Y. Yang, Q. Lu, M. Liu et al., "Epidemiological and clinical features of the 2019 novel coronavirus outbreak in China," medRxiv, no. article 20021675, 2020.
- [48] Y. H. Jin, L. Cai, Z. S. Cheng et al., "A rapid advice guideline for the diagnosis and treatment of 2019 novel coronavirus (2019-nCoV) infected pneumonia (standard version)," *Military Medical Research*, vol. 7, no. 1, p. 4, 2020.
- [49] Z. Leng, R. Zhu, W. Hou et al., "Transplantation of ACE2(-) mesenchymal stem cells improves the outcome of patients with COVID-19 pneumonia," *Aging and Disease*, vol. 11, no. 2, pp. 216–228, 2020.
- [50] V. J. Munster, M. Koopmans, and N. V. Doremalen, "A novel coronavirus emerging in China-key questions for impact assessment," *The New England Journal of Medicine*, vol. 382, no. 8, pp. 692–694, 2020.
- [51] C. Sohrabi, Z. Alsafi, N. O'Neill et al., "World Health Organization declares global emergency: A review of the 2019 novel coronavirus (COVID-19)," *International Journal of Surgery*, vol. 76, pp. 71–76, 2020.
- [52] C. Mattiuzzi and G. Lippi, "Which lessons shall we learn from the 2019 novel coronavirus outbreak?," Annals of translational medicine, vol. 8, no. 3, 2020.
- [53] I. Hamming, W. Timens, M. L. Bulthuis, A. T. Lely, G. V. Navis, and H. van Goor, "Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis," *The Journal of Pathology*, vol. 203, no. 2, pp. 631–637, 2004.
- [54] W. J. Guan, Z. Y. Ni, Y. Hu et al., "Clinical characteristics of coronavirus disease 2019 in China," New England Journal of Medicine, vol. 382, no. 18, pp. 1708–1720, 2020.
- [55] C. Huang, Y. Wang, X. Li et al., "Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China," *The Lancet*, vol. 395, no. 10223, pp. 497–506, 2020.
- [56] F. Chen and D. Lou, "Rising concern on damaged testis of COVID-19 patients," *Urology*, vol. 142, p. 42, 2020.
- [57] C. Fan, K. Li, Y. Ding, W. L. Lu, and J. Wang, "ACE2 expression in kidney and testis may cause kidney and testis damage after 2019-nCoV infection 2020," 2020, https://www.medrxiv.org/content/10.1101/2020.02.12.20022418v1%20.abstract/.
- [58] A. Akhmerov and E. Marbán, "COVID-19 and the heart," *Circulation Research*, vol. 126, no. 10, pp. 1443–1455, 2020.
- [59] J. R. Lechien, C. M. Chiesa-Estomba, D. R. De Siati et al., "Olfactory and gustatory dysfunctions as a clinical presentation of mild-to-moderate forms of the coronavirus disease (COVID-19): a multicenter European study," *European Archives of Oto-Rhino-Laryngology*, vol. 277, no. 8, pp. 2251–2261, 2020.

[60] W. Wang, J. Tang, and F. Wei, "Updated understanding of the outbreak of 2019 novel coronavirus (2019-nCoV) in Wuhan, China," *Journal of Medical Virology*, vol. 92, no. 4, pp. 441–447, 2020.

- [61] W. Liu, Z.-W. Tao, L. Wang et al., "Analysis of factors associated with disease outcomes in hospitalized patients with 2019 novel coronavirus disease," *Chinese Medical Journal*, vol. 133, no. 9, pp. 1032–1038, 2020.
- [62] A. K. Shetty, R. Upadhya, L. N. Madhu, and M. Kodali, "Novel insights on systemic and brain aging, stroke, amyotrophic lateral sclerosis, and Alzheimer's disease," *Aging and Disease*, vol. 10, no. 2, pp. 470–482, 2019.
- [63] X. Yang, Y. Yu, J. Xu et al., "Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: a single-centered, retrospective, observational study," *The Lancet Respiratory Medicine*, vol. 8, no. 5, pp. 475–481, 2020.
- [64] R. Thomas, W. Wang, and D.-M. Su, "Contributions of agerelated thymic involution to immunosenescence and inflammaging," *Immunity & Ageing*, vol. 17, p. 1, 2020.
- [65] S. J. Oh, J. K. Lee, and O. S. Shin, "Aging and the immune system: the impact of immunosenescence on viral infection, immunity and vaccine immunogenicity," *Immune Network*, vol. 19, no. 6, article e37, 2019.
- [66] A. K. Shetty, M. Kodali, R. Upadhya, and L. N. Madhu, "Emerging anti-aging strategies-scientific basis and efficacy," *Aging and Disease*, vol. 9, no. 6, pp. 1165–1184, 2018.
- [67] J. H. Kuhn, W. Li, H. Choe, and M. Farzan, "Angiotensin-converting enzyme 2: a functional receptor for SARS coronavirus," *Cellular and Molecular Life Sciences*, vol. 61, no. 21, pp. 2738–2743, 2004.
- [68] Z. Qian, E. A. Travanty, L. Oko et al., "Innate immune response of human alveolar type II cells infected with severe acute respiratory syndrome-coronavirus," *American Journal* of Respiratory Cell and Molecular Biology, vol. 48, no. 6, pp. 742–748, 2013.
- [69] E. Mick, J. Kamm, A. O. Pisco et al., "Upper airway gene expression differentiates COVID-19 from other acute respiratory illnesses and reveals suppression of innate immune responses by SARS-CoV-2," 2020, medRxiv.
- [70] D. Wrapp, N. Wang, K. S. Corbett et al., "Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation," *Science*, vol. 367, no. 648, pp. 1260–1263, 2020.
- [71] H. Chu, J. F. Chan, Y. Wang et al., "Comparative replication and immune activation profiles of SARS-CoV-2 and SARS-CoV in human lungs: an ex vivo study with implications for the pathogenesis of COVID-19," *Clinical Infectious Diseases*, vol. 71, no. 6, pp. 1400–1409, 2020.
- [72] M. Hoffmann, H. Kleine-Weber, S. Schroeder et al., "SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor," *Cell*, vol. 181, no. 2, pp. 271–280.e278, 2020.
- [73] F. Coperchini, L. Chiovato, L. Croce, F. Magri, and M. Rotondi, "The cytokine storm in COVID-19: an overview of the involvement of the chemokine/chemokine-receptor system," Cytokine & Growth Factor Reviews, vol. 53, pp. 25–32, 2020.
- [74] P. Mehta, M. A. DF, M. Brown, E. Sanchez, R. S. Tattersall, and J. J. Manson, "COVID-19: consider cytokine storm syndromes and immunosuppression," *The Lancet*, vol. 395, no. 10229, pp. 1033-1034, 2020.

- [75] Y. Yang, C. Shen, J. Li et al., "Plasma IP-10 and MCP-3 levels are highly associated with disease severity and predict the progression of COVID-19," *Journal of Allergy* and Clinical Immunology, vol. 146, no. 1, pp. 119– 127.e114, 2020.
- [76] K. Liu, Y. Y. Fang, Y. Deng et al., "Clinical characteristics of novel coronavirus cases in tertiary hospitals in Hubei Province," *Chinese Medical Journal*, vol. 133, no. 9, pp. 1025– 1031, 2020.
- [77] Z. Xu, L. Shi, Y. Wang et al., "Pathological findings of COVID-19 associated with acute respiratory distress syndrome," *The Lancet Respiratory Medicine*, vol. 8, no. 4, pp. 420–422, 2020.
- [78] P. Spagnolo, E. Balestro, S. Aliberti et al., "Pulmonary fibrosis secondary to COVID-19: a call to arms?," *The Lancet Respira*tory Medicine, vol. 8, no. 8, pp. 750–752, 2020.
- [79] X. Lu, Y. Xing, and G. W. Wong, "COVID-19: lessons to date from China," *Archives of Disease in Childhood*, vol. 105, no. 12, pp. 1146–1150, 2020.
- [80] K. K. Sahu, A. K. Mishra, and A. Lal, "Trajectory of the COVID-19 pandemic: chasing a moving target," *Annals* of *Translational Medicine*, vol. 8, no. 11, pp. 694–694, 2020.
- [81] Q. L. Zeng, Z. J. Yu, J. J. Gou et al., "Effect of convalescent plasma therapy on viral shedding and survival in patients with coronavirus disease 2019," *The Journal of Infectious Diseases*, vol. 222, no. 1, pp. 38–43, 2020.
- [82] W. Han, M. Zhu, J. Chen et al., "Lung transplantation for elderly patients with end-stage COVID-19 pneumonia," *Annals of Surgery*, vol. 272, no. 1, pp. e33–e34, 2020.
- [83] R. E. Ferner and J. K. Aronson, "Remdesivir in covid-19," BMJ, vol. 369, p. m1610, 2020.
- [84] M. Wang, R. Cao, L. Zhang et al., "Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro," *Cell Research*, vol. 30, no. 3, pp. 269–271, 2020.
- [85] T. U. Singh, S. Parida, M. C. Lingaraju, M. Kesavan, D. Kumar, and R. K. Singh, "Drug repurposing approach to fight COVID-19," *Pharmacological Reports*, vol. 72, no. 6, pp. 1479–1508, 2020.
- [86] J. P. Zhao, Y. Hu, C. Z. S. Du RH et al., "Expert consensus on the use of corticosteroid in patients with 2019-nCoV pneumonia," *Zhonghua Jie He He Hu Xi Za Zhi*, vol. 43, no. 3, pp. 183-184, 2020.
- [87] A. Torres, O. Sibila, M. Ferrer et al., "Effect of corticosteroids on treatment failure among hospitalized patients with severe community-acquired pneumonia and high inflammatory response," *JAMA*, vol. 313, no. 7, pp. 677–686, 2015.
- [88] Z. Hu, M. Yang, and C. Xie, "Efficacy and safety of Lian-Hua Qing-Wen granule for COVID-2019," *Medicine*, vol. 99, no. 23, article e20203, 2020.
- [89] D. Gurwitz, "Angiotensin receptor blockers as tentative SARS-CoV-2 therapeutics," *Drug Development Research*, vol. 81, no. 5, pp. 537–540, 2020.
- [90] U.S. National Institutes of Health, "ACE Inhibitors, Angiotensin II Type-I Receptor Blockers and Severity of COVID-19 (CODIV-ACE)," 2020, https://clinicaltrials.gov/ct2/show/NCT04318418?term=NCT04318418&draw=2&rank=1.
- [91] Y. Y. Zheng, Y. T. Ma, J. Y. Zhang, and X. Xie, "COVID-19 and the cardiovascular system," *Nature Reviews. Cardiology*, vol. 17, no. 5, pp. 259-260, 2020.

[92] M. Esler and D. Esler, "Can angiotensin receptor-blocking drugs perhaps be harmful in the COVID-19 pandemic?," *Journal of Hypertension*, vol. 38, no. 5, pp. 781-782, 2020.

- [93] H. Wang, Y. Zhang, B. Huang et al., "Development of an inactivated vaccine candidate, BBIBP-CorV, with potent protection against SARS-CoV-2," *Cell*, vol. 182, no. 3, pp. 713– 721, 2020.
- [94] Anon, "China currently has 15 COVID-19 vaccines in clinical trials," 2020, https://baijiahao.baidu.com/s?id= 1686692193551096109&wfr=spider&for=pc.[Accessed,% 202020/.
- [95] B. M. Henry, "COVID-19, ECMO, and lymphopenia: a word of caution," *The Lancet Respiratory Medicine*, vol. 8, no. 4, 2020
- [96] J. B. Sneddon, Q. Tang, P. Stock et al., "Stem cell therapies for treating diabetes: progress and remaining challenges," *Cell Stem Cell*, vol. 22, no. 6, pp. 810–823, 2018.
- [97] L. Arranz, A. Sánchez-Aguilera, D. Martín-Pérez et al., "Neuropathy of haematopoietic stem cell niche is essential for myeloproliferative neoplasms," *Nature*, vol. 512, no. 7512, pp. 78–81, 2014.
- [98] J. Burman, A. Tolf, H. Hagglund, and H. Askmark, "Autologous haematopoietic stem cell transplantation for neurological diseases," *Journal of Neurology, Neurosurgery, and Psychiatry*, vol. 89, no. 2, pp. 147–155, 2018.
- [99] J. Jia and Z. Jia, "Cell-based therapy in lung regenerative medicine," Regenerative Medicine Research, vol. 2, no. 1, p. 7, 2014.
- [100] C. M. Kolf, E. Cho, and R. S. Tuan, "Mesenchymal stromal cells. Biology of adult mesenchymal stem cells: regulation of niche, self-renewal and differentiation," *Arthritis Research & Therapy*, vol. 9, no. 1, p. 204, 2007.
- [101] J. D. Glenn, "Mesenchymal stem cells: emerging mechanisms of immunomodulation and therapy," World Journal of Stem Cells, vol. 6, no. 5, pp. 526–539, 2014.
- [102] J. Behnke, S. Kremer, T. Shahzad et al., "MSC based therapies-new perspectives for the injured lung," *Journal of Clinical Medicine*, vol. 9, no. 3, p. 682, 2020.
- [103] P. De Miguel, S. Fuentes-Julian, A. Blazquez-Martinez et al., "Immunosuppressive properties of mesenchymal stem cells: advances and applications," *Current Molecular Medicine*, vol. 12, no. 5, pp. 574–591, 2012.
- [104] X. Wu, V. L. D. Thi, Y. Huang et al., "Intrinsic immunity shapes viral resistance of stem cells," *Cell*, vol. 172, no. 3, pp. 423–438, 2018.
- [105] D. C. Burke, C. F. Graham, and J. M. Lehman, "Appearance of interferon inducibility and sensitivity during differentiation of murine teratocarcinoma cells in vitro," *Cell*, vol. 13, no. 2, pp. 243–249, 1978.
- [106] A. Uccelli, L. Moretta, and V. Pistoia, "Mesenchymal stem cells in health and disease," *Nature Reviews Immunology*, vol. 8, no. 9, pp. 726–736, 2008.
- [107] X. Fu, G. Liu, A. Halim, Y. Ju, Q. Luo, and G. Song, "Mesenchymal stem cell migration and tissue repair," *Cells*, vol. 8, no. 8, 2019.
- [108] C. Sobacchi, E. Palagano, A. Villa, and C. Menale, "Soluble factors on stage to direct mesenchymal stem cells fate," Frontiers in Bioengineering and Biotechnology, vol. 5, p. 32, 2017.
- [109] H. Loy, D. I. Kuok, K. P. Hui et al., "Therapeutic implications of human umbilical cord mesenchymal stromal cells in attenuating influenza A (H5N1) virus-associated acute lung

- injury," The Journal of Infectious Diseases, vol. 219, no. 2, pp. 186–196, 2019.
- [110] I. Darwish, M. Samira, and W. Conrad Liles, "Immunomodulatory therapy for severe influenza," *Expert Review of Anti-Infective Therapy*, vol. 9, no. 7, pp. 807–822, 2011.
- [111] P. U. Dinh, D. Paudel, H. Brochu et al., "Inhalation of lung spheroid cell secretome and exosomes promotes lung repair in pulmonary fibrosis," *Nature Communications*, vol. 11, no. 1, p. 1064, 2020.
- [112] P. Connick, M. Kolappan, C. Crawley et al., "Autologous mesenchymal stem cells for the treatment of secondary progressive multiple sclerosis: an open-label phase 2a proof-ofconcept study," *The Lancet Neurology*, vol. 11, no. 2, pp. 150–156, 2012.
- [113] J. G. Wilson, K. D. Liu, H. Zhuo et al., "Mesenchymal stem (stromal) cells for treatment of ARDS: a phase 1 clinical trial," *The Lancet Respiratory Medicine*, vol. 3, no. 1, pp. 24–32, 2015.
- [114] L. Gu, T. Li, Y. Zeng et al., "Human umbilical cord mesenchymal stem cells combined with antiviral therapy to treat COVID-19," *Journal of Kunming Medical University*, vol. 41, no. 3, pp. 96–100, 2020.
- [115] U.S. National Institutes of Health, "Mesenchymal stem cell|-Completed Studies|COVID-19," 2021, https://clinicaltrials .gov/ct2/results?term=mesenchymal.+stem+cell&%20recrs= e&cond=COVID-19&draw=2&rank=1#rowId0/.
- [116] F. Wang, "Stem cell therapy is safe and effective. Patients with COVID-19 can receive stem cell therapy at the onset and later stages," 2020, https://www.sohu.com/a/421016082\_ 120054627.%202020/.
- [117] A. Zumla, F.-S. Wang, G. Ippolito et al., "Reducing mortality and morbidity in patients with severe COVID-19 disease by advancing ongoing trials of mesenchymal stromal (stem) cell (MSC) therapy-achieving global consensus and visibility for cellular host-directed therapies," *International Journal of Infectious Diseases*, vol. 96, pp. 431–439, 2020.
- [118] R. H. Lee, A. A. Pulin, M. J. Seo et al., "Intravenous hMSCs improve myocardial infarction in mice because cells embolized in lung are activated to secrete the anti-inflammatory protein TSG-6," Cell Stem Cell, vol. 5, no. 1, pp. 54–63, 2009.
- [119] Y. Zhou, B. Fu, X. Zheng et al., "Pathogenic T-cells and inflammatory monocytes incite inflammatory storms in severe COVID-19 patients," *National Science Review*, vol. 7, no. 6, pp. 998–1002, 2020.
- [120] X. Yang, T. Dai, X. Zhou et al., "Analysis of adaptive immune cell populations and phenotypes in the patients infected by SARS-CoV-2," 2020, https://www.medrxiv.org/content/10 .1101/2020.03.23.20040675v2/.
- [121] D. Zhang, R. Guo, L. Lei et al., "COVID-19 infection induces readily detectable morphological and inflammation-related phenotypic changes in peripheral blood monocytes, the severity of which correlate with patient outcome," 2020, https://www.medrxiv.org/content/10.1101/2020.03.24 .20042655v1/.
- [122] M. Liao, Y. Liu, J. Yuan et al., "Single-cell landscape of bronchoalveolar immune cells in patients with COVID-19," *Nature Medicine*, vol. 26, no. 6, article 901, pp. 842–844, 2020
- [123] J. Chen, H. Fan, L. Zhang et al., "Retrospective analysis of clinical features in 101 death cases with COVID-19," 2020, https://www.medrxiv.org/content/10.1101/2020.03.09 .20033068v2/.

- [124] S. Wan, Q. Yi, S. Fan et al., "Characteristics of lymphocyte subsets and cytokines in peripheral blood of 123 hospitalized patients with 2019 novel coronavirus pneumonia (NCP)," 2020, https://www.medrxiv.org/content/10.1101/2020.02.10 20021832v1/
- [125] S. Gupta, R. Bi, C. Kim, S. Chiplunkar, L. Yel, and S. Gollapudi, "Role of NF- κB signaling pathway in increased tumor necrosis factor- \_α\_ -induced apoptosis of lymphocytes in aged humans," *Cell Death and Differentiation*, vol. 12, no. 2, pp. 177–183, 2005.
- [126] D. Wu and X. O. Yang, "TH17 responses in cytokine storm of COVID-19: an emerging target of JAK2 inhibitor Fedratinib," *Journal of Microbiology, Immunology, and Infection*, vol. 53, no. 3, pp. 368–370, 2020.
- [127] Y. Deng, Y. Zhang, L. Ye et al., "Umbilical cord-derived mesenchymal stem cells instruct monocytes towards an IL10-producing phenotype by secreting IL6 and HGF," *Scientific Reports*, vol. 6, no. 1, article 37566, 2016.
- [128] D. I. Cho, M. R. Kim, H. Y. Jeong et al., "Mesenchymal stem cells reciprocally regulate the M1/M2 balance in mouse bone marrow-derived macrophages," *Experimental & Molecular Medicine*, vol. 46, no. 1, p. e70, 2014.
- [129] J. Kim and P. Hematti, "Mesenchymal stem cell-educated macrophages: a novel type of alternatively activated macrophages," *Experimental Hematology*, vol. 37, no. 12, pp. 1445–1453, 2009.
- [130] J. Maggini, G. Mirkin, I. Bognanni et al., "Mouse bone marrow-derived mesenchymal stromal cells turn activated macrophages into a regulatory-like profile," *PLoS One*, vol. 5, no. 2, article e9252, 2010.
- [131] G. M. Spaggiari, A. Capobianco, S. Becchetti, M. C. Mingari, and L. Moretta, "Mesenchymal stem cell-natural killer cell interactions: evidence that activated NK cells are capable of killing MSCs, whereas MSCs can inhibit IL-2-induced NK-cell proliferation," *Blood*, vol. 107, no. 4, pp. 1484–1490, 2006
- [132] Y. Y. Zhou, Y. Yamamoto, Z. D. Xiao, and T. Ochiya, "The Immunomodulatory functions of mesenchymal stromal/stem cells mediated via paracrine activity," *Journal of Clinical Medicine*, vol. 8, no. 7, p. 1025, 2019.
- [133] Q. Zhao, H. Ren, and Z. Han, "Mesenchymal stem cells: immunomodulatory capability and clinical potential in immune diseases," *Journal of Cellular Immunotherapy*, vol. 2, no. 1, pp. 3–20, 2016.
- [134] A. R. R. Weiss and M. H. Dahlke, "Immunomodulation by mesenchymal stem cells (MSCs): mechanisms of action of living, apoptotic, and dead MSCs," *Frontiers in Immunology*, vol. 10, p. 1191, 2019.
- [135] H. Fayyad-Kazan, W. H. Faour, B. Badran, L. Lagneaux, and M. Najar, "The immunomodulatory properties of human bone marrow-derived mesenchymal stromal cells are defined according to multiple immunobiological criteria," *Inflammation Research*, vol. 65, no. 6, pp. 501–510, 2016.
- [136] G. F. Curley, M. Hayes, B. Ansari et al., "Mesenchymal stem cells enhance recovery and repair following ventilator-induced lung injury in the rat," *Thorax*, vol. 67, no. 6, pp. 496–501, 2012.
- [137] M. A. Antunes, S. C. Abreu, F. F. Cruz et al., "Effects of different mesenchymal stromal cell sources and delivery routes in experimental emphysema," *Respiratory Research*, vol. 15, no. 1, p. 118, 2014.

[138] K. D. Lee, T. K. C. Kuo, J. Whang-Peng et al., "In vitro hepatic differentiation of human mesenchymal stem cells," *Hepatol*ogy, vol. 40, no. 6, pp. 1275–1284, 2004.

- [139] P. Tropel, N. Platet, J. C. Platel et al., "Functional neuronal differentiation of bone marrow-derived mesenchymal stem cells," *Stem Cells*, vol. 24, no. 12, pp. 2868–2876, 2006.
- [140] M. Z. Ratajczak, K. Marycz, A. Poniewierska-Baran, K. Fiedorowicz, M. Zbucka-Kretowska, and M. Moniuszko, "Very small embryonic-like stem cells as a novel developmental concept and the hierarchy of the stem cell compartment," *Advances in Medical Sciences*, vol. 59, no. 2, pp. 273–280, 2014.
- [141] D. J. Weiss, "Mesenchymal Stem Cells for Lung Repair and Regeneration," in *Stem Cells in the Respiratory System*, pp. 25–42, Humana Press, Totowa, New Jersey, 2010.
- [142] C. R. Harrell, N. Jovicic, V. Djonov, N. Arsenijevic, and V. Volarevic, "Mesenchymal stem cell-derived exosomes and other extracellular vesicles as new remedies in the therapy of inflammatory diseases," *Cells*, vol. 8, no. 12, p. 1605, 2019.
- [143] M. Rojas, J. Xu, C. R. Woods et al., "Bone marrow-derived mesenchymal stem cells in repair of the injured lung," American Journal of Respiratory Cell and Molecular Biology, vol. 33, no. 2, pp. 145–152, 2005.
- [144] J. W. Lee, X. Fang, N. Gupta, V. Serikov, and M. A. Matthay, "Allogeneic human mesenchymal stem cells for treatment of E. coli endotoxin-induced acute lung injury in the ex vivo perfused human lung," *PNAS*, vol. 106, no. 38, pp. 16357–16362, 2009.
- [145] Y. Yang, S. Hu, X. Xu et al., "The vascular endothelial growth factors-expressing character of mesenchymal stem cells plays a positive role in treatment of acute lung injury in vivo," *Mediators of Inflammation*, vol. 2016, Article ID 2347938, 12 pages, 2016.
- [146] Y. Yang, Q. H. Chen, A. R. Liu, X. P. Xu, J. B. Han, and H. B. Qiu, "Synergism of MSC-secreted HGF and VEGF in stabilising endothelial barrier function upon lipopolysaccharide stimulation via the Rac1 pathway," *Stem Cell Research & Therapy*, vol. 6, no. 1, p. 250, 2015.
- [147] Z. Zhou and Z. You, "Mesenchymal stem cells alleviate LPS-induced acute lung injury in mice by MiR-142a-5p-controlled pulmonary endothelial cell autophagy," *Cellular Physiology and Biochemistry*, vol. 38, no. 1, pp. 258–266, 2016.
- [148] S. Hu, J. Park, A. Liu et al., "Mesenchymal stem cell microvesicles restore protein permeability across primary cultures of injured human lung microvascular endothelial cells," *Stem Cells Translational Medicine*, vol. 7, no. 8, pp. 615–624, 2018.
- [149] S. S. Meng, F. M. Guo, X. W. Zhang et al., "MTOR/STAT-3 pathway mediates mesenchymal stem cell-secreted hepatocyte growth factor protective effects against lipopolysaccharide-induced vascular endothelial barrier dysfunction and apoptosis," *Journal of Cellular Biochemistry*, vol. 120, no. 3, pp. 3637–3650, 2019.
- [150] A.-r. Liu, L. Liu, S. Chen et al., "Activation of canonical wnt pathway promotes differentiation of mouse bone marrowderived MSCs into type II alveolar epithelial cells, confers resistance to oxidative stress, and promotes their migration to injured lung tissue in vitro," *Journal of Cellular Physiology*, vol. 228, no. 6, pp. 1270–1283, 2013.
- [151] S. Cai, A. Liu, S. Chen et al., "Activation of Wnt/β-catenin signalling promotes mesenchymal stem cells to repair injured

alveolar epithelium induced by lipopolysaccharide in mice," *Stem Cell Research & Therapy*, vol. 6, no. 1, p. 65, 2015.

- [152] Y. Li, X. Shi, L. Yang et al., "Hypoxia promotes the skewed differentiation of umbilical cord mesenchymal stem cells toward type II alveolar epithelial cells by regulating micro-RNA-145," *Gene*, vol. 630, pp. 68–75, 2017.
- [153] L. Zhang, Q. Li, W. Liu, Z. Liu, H. Shen, and M. Zhao, "Mesenchymal stem cells alleviate acute lung injury and inflammatory responses induced by paraquat poisoning," *Medical Science Monitor*, vol. 25, article 915804, pp. 2623–2632, 2019.
- [154] X. Fang, A. P. Neyrinck, M. A. Matthay, and J. W. Lee, "Allogeneic Human Mesenchymal Stem Cells Restore Epithelial Protein Permeability in Cultured Human Alveolar Type II Cells by Secretion of Angiopoietin-1," *Journal of Biological Chemistry*, vol. 285, no. 34, pp. 26211–26222, 2010.
- [155] A. Goolaerts, N. Pellan-Randrianarison, J. Larghero et al., "Conditioned media from mesenchymal stromal cells restore sodium transport and preserve epithelial permeability in an in vitro model of acute alveolar injury," American Journal of Physiology. Lung Cellular and Molecular Physiology, vol. 306, no. 11, pp. L975–L985, 2014.
- [156] O. Bernard, F. Jeny, Y. Uzunhan et al., "Mesenchymal stem cells reduce hypoxia-induced apoptosis in alveolar epithelial cells by modulating HIF and ROS hypoxic signaling," *American Journal of Physiology. Lung Cellular and Molecular Physiology*, vol. 314, no. 3, pp. L360–L371, 2018.
- [157] X. Yan, X. Fu, Y. Jia et al., "Nrf2/Keap1/ARE signaling mediated an antioxidative protection of human placental mesenchymal stem cells of fetal origin in alveolar epithelial cells," Oxidative Medicine and Cellular Longevity, Article ID 2654910, 2019.
- [158] T. Maron-Gutierrez, J. D. Silva, K. D. Asensi et al., "Effects of mesenchymal stem cell therapy on the time course of pulmonary remodeling depend on the etiology of lung injury in mice," *Critical Care Medicine*, vol. 41, no. 11, pp. e319– e333, 2013.
- [159] J. D. Silva, M. Lopes-Pacheco, A. H. R. Paz et al., "Mesenchymal stem cells from bone marrow, adipose tissue, and lung tissue differentially mitigate lung and distal organ damage in experimental acute respiratory distress syndrome," *Critical Care Medicine*, vol. 46, no. 2, pp. e132–e140, 2018.
- [160] A. R. Medford and A. B. Millar, "Vascular endothelial growth factor (VEGF) in acute lung injury (ALI) and acute respiratory distress syndrome (ARDS): paradox or paradigm?," *Thorax*, vol. 61, no. 7, pp. 621–626, 2006.
- [161] A. K. Kaza, V. E. Laubach, J. A. Kern et al., "Epidermal growth factor augments postpneumonectomy lung growth," *The Journal of Thoracic and Cardiovascular Surgery*, vol. 120, no. 5, pp. 916–922, 2000.
- [162] R. A. Panganiban and R. M. Day, "Hepatocyte growth factor in lung repair and pulmonary fibrosis," *Acta Pharmacologica Sinica*, vol. 32, no. 1, pp. 12–20, 2011.
- [163] Z. H. Qin, J. F. Xu, J. M. Qu et al., "Intrapleural delivery of MSCs attenuates acute lung injury by paracrine/endocrine mechanism," *Journal of Cellular and Molecular Medicine*, vol. 16, no. 11, pp. 2745–2753, 2012.
- [164] B. Antebi, A. Mohammadipoor, A. I. Batchinsky, and L. C. Cancio, "The promise of mesenchymal stem cell therapy for acute respiratory distress syndrome," *Journal of Trauma and Acute Care Surgery*, vol. 84, no. 1, pp. 183–191, 2018.

- [165] M. R. Mokhber Dezfouli, M. Jabbari Fakhr, S. Sadeghian Chaleshtori, M. M. Dehghan, A. Vajhi, and R. Mokhtari, "Intrapulmonary autologous transplant of bone marrow-derived mesenchymal stromal cells improves lipopolysaccharide-induced acute respiratory distress syndrome in rabbit," *Critical Care*, vol. 22, no. 1, article 2272, p. 353, 2018.
- [166] S. Golpanian, A. Wolf, K. E. Hatzistergos, and J. M. Hare, "Rebuilding the damaged heart: mesenchymal stem cells, cell-based therapy, and engineered heart tissue," *Physiological Reviews*, vol. 96, no. 3, pp. 1127–1168, 2016.
- [167] C. W. Lee, Y. F. Chen, H. H. Wu, and O. K. Lee, "Historical perspectives and advances in mesenchymal stem cell research for the treatment of liver diseases," *Gastroenterology*, vol. 154, no. 1, pp. 46–56, 2018.
- [168] L. Perico, M. Morigi, C. Rota et al., "Human mesenchymal stromal cells transplanted into mice stimulate renal tubular cells and enhance mitochondrial function," *Nature Communications*, vol. 8, no. 1, article 937, p. 983, 2017.
- [169] D. Garcia-Olmo and D. A. Schwartz, "Cumulative Evidence That Mesenchymal Stem Cells Promote Healing of Perianal Fistulas of Patients With Crohn's Disease-Going From Bench to Bedside," *Gastroenterology*, vol. 149, no. 4, pp. 853–857, 2015.
- [170] I. Molendijk, B. A. Bonsing, H. Roelofs et al., "Allogeneic Bone Marrow-Derived Mesenchymal Stromal Cells Promote Healing of Refractory Perianal Fistulas in Patients With Crohn's Disease," *Gastroenterology*, vol. 149, no. 4, pp. 918–927.e6, 2015.
- [171] J. E. Millar, V. von Bahr, M. V. Malfertheiner et al., "Administration of mesenchymal stem cells during ECMO results in a rapid decline in oxygenator performance," *Thorax*, vol. 74, no. 2, pp. 194–196, 2019.
- [172] D. Rubio, S. Garcia, T. de la Cueva et al., "Human mesenchymal stem cell transformation is associated with a mesenchymal-epithelial transition," *Experimental cell research*, vol. 314, no. 4, pp. 691–698, 2008.
- [173] M. Breitbach, T. Bostani, W. Roell et al., "Potential risks of bone marrow cell transplantation into infarcted hearts," *Blood*, vol. 110, no. 4, pp. 1362–1369, 2007.
- [174] R. M. Johnstone, M. Adam, J. R. Hammond, L. Orr, and C. Turbide, "Vesicle formation during reticulocyte maturation. Association of plasma membrane activities with released vesicles (exosomes).," *Journal of Biological Chemistry*, vol. 262, no. 19, pp. 9412–9420, 1987.
- [175] M. Colombo, G. Raposo, and C. Thery, "Biogenesis, secretion, and Intercellular Interactions of exosomes and other extracellular vesicles," *Annual review of cell and developmental biology*, vol. 30, no. 1, pp. 255–289, 2014.
- [176] A. Lo Cicero, P. D. Stahl, and G. Raposo, "Extracellular vesicles shuffling intercellular messages: for good or for bad," *Current opinion in cell biology*, vol. 35, pp. 69–77, 2015.
- [177] M. Yáñez-Mó, P. R. M. Siljander, Z. Andreu et al., "Biological properties of extracellular vesicles and their physiological functions," *Journal of extracellular vesicles*, vol. 4, no. 1, article 27066, 2015.
- [178] R. Crescitelli, C. Lässer, T. G. Szabó et al., "Distinct RNA profiles in subpopulations of extracellular vesicles: apoptotic bodies, microvesicles and exosomes," *Journal of extracellular vesicles*, vol. 2, no. 1, article 20677, 2013.
- [179] G. Turturici, R. Tinnirello, G. Sconzo, and F. Geraci, "Extracellular membrane vesicles as a mechanism of cell-to-cell

communication: advantages and disadvantages," *American Journal of Physiology. Cell Physiology*, vol. 306, no. 7, pp. C621–C633, 2014.

- [180] Q. Hao, V. Gudapati, A. Monsel et al., "Mesenchymal stem cell-derived extracellular vesicles decrease lung injury in mice," *Journal of Immunology*, vol. 203, no. 7, pp. 1961– 1972, 2019.
- [181] J. H. Lee, J. Park, and J. W. Lee, "Therapeutic use of mesenchymal stem cell-derived extracellular vesicles in acute lung injury," *Transfusion*, vol. 59, no. S1, pp. 876–883, 2019.
- [182] A. Abraham and A. Krasnodembskaya, "Mesenchymal stem cell-derived extracellular vesicles for the treatment of acute respiratory distress syndrome," *Stem Cells Translational Medicine*, vol. 9, no. 1, pp. 28–38, 2020.
- [183] A. I. Caplan, "Mesenchymal stem cells: time to change the name!," *Stem Cells Translational Medicine*, vol. 6, no. 6, pp. 1445–1451, 2017.
- [184] A. I. Caplan, "What's in a name?," Tissue Engineering. Part A, vol. 16, no. 8, pp. 2415–2417, 2010.
- [185] B. Crivelli, T. Chlapanidas, S. Perteghella et al., "Mesenchymal stem/stromal cell extracellular vesicles: from active principle to next generation drug delivery system," *Journal of Controlled Release*, vol. 262, pp. 104–117, 2017.
- [186] G. Di Rocco, S. Baldari, and G. Toietta, "Towards therapeutic delivery of extracellular vesicles: strategies for in vivo tracking and biodistribution analysis," *Stem cells international*, vol. 2016, Article ID 5029619, 12 pages, 2016.
- [187] B. J. Gaborit, J.-F. Bergmann, C. Mussini et al., "Plea for multitargeted interventions for severe COVID-19," *The Lancet Infectious Diseases*, vol. 20, no. 10, pp. 1122-1123, 2020.
- [188] M. Wang, Q. Yuan, and L. Xie, "Mesenchymal stem cell-based immunomodulation: properties and clinical application," Stem Cells International, vol. 2018, Article ID 3057624, 12 pages, 2018.
- [189] X. Qian, C. Xu, S. Fang et al., "Exosomal microRNAs derived from umbilical mesenchymal stem cells inhibit hepatitis C virus infection," *Stem cells translational medicine*, vol. 5, no. 9, pp. 1190–1203, 2016.
- [190] L. Margolis and Y. Sadovsky, "The biology of extracellular vesicles: the known unknowns," *PLoS Biology*, vol. 17, no. 7, article e3000363, 2019.
- [191] F. Fanini and M. Fabbri, "Cancer-derived exosomic micro-RNAs shape the immune system within the tumor microen-vironment: State of the art," Seminars in cell & developmental biology, vol. 67, no. 23-28, pp. 23-28, 2017.
- [192] U. Agarwal, A. George, S. Bhutani et al., "Experimental, systems, and computational approaches to understanding the microRNA-mediated reparative potential of cardiac progenitor cell-derived exosomes from pediatric patients," *Circulation Research*, vol. 120, no. 4, pp. 701–712, 2017.
- [193] F. Prattichizzo, A. Giuliani, V. de Nigris et al., "Extracellular microRNAs and endothelial hyperglycaemic memory: a therapeutic opportunity?," *Diabetes, Obesity & Metabolism*, vol. 18, no. 9, pp. 855–867, 2016.
- [194] I. Huang-Doran, C. Y. Zhang, and A. Vidal-Puig, "Extracellular vesicles: novel mediators of cell communication in metabolic disease," *Trends in Endocrinology and Metabolism*, vol. 28, no. 1, pp. 3–18, 2017.
- [195] A. Khalyfa, L. Kheirandish-Gozal, A. A. Khalyfa et al., "Circulating plasma extracellular microvesicle microRNA cargo and endothelial dysfunction in children with obstructive sleep

- apnea," American Journal of Respiratory and Critical Care Medicine, vol. 194, no. 9, pp. 1116-1126, 2016.
- [196] T. Thomou, M. A. Mori, J. M. Dreyfuss et al., "Adipose-derived circulating miRNAs regulate gene expression in other tissues," *Nature*, vol. 542, no. 7642, pp. 450–455, 2017.
- [197] M. Khatri, L. A. Richardson, and T. Meulia, "Mesenchymal stem cell-derived extracellular vesicles attenuate influenza virus-induced acute lung injury in a pig model," *Stem Cell Research & Therapy*, vol. 9, no. 1, article 774, p. 17, 2018.
- [198] K. Jiang, J. Yang, S. Guo, G. Zhao, H. Wu, and G. Deng, "Peripheral circulating exosome-mediated delivery of miR-155 as a novel mechanism for acute lung inflammation," *Molecular Therapy*, vol. 27, no. 10, pp. 1758–1771, 2019.
- [199] E. Bari, I. Ferrarotti, M. L. Torre, A. G. Corsico, and S. Perteghella, "Mesenchymal stem/stromal cell secretome for lung regeneration: The long way through "pharmaceuticalization" for the best formulation," *Journal of Controlled Release*, vol. 309, pp. 11–24, 2019.
- [200] U.S. National Institutes of Health, "Studies found for: exosome|COVID19," 2020, https://clinicaltrials.gov/ct2/results?cond=COVID19&term=exosome +&cntry=&state=&city=&dist=. 2020.
- [201] V. Sengupta, S. Sengupta, A. Lazo, P. Woods, A. Nolan, and N. Bremer, "Exosomes derived from bone marrow Mesenchymal stem cells as treatment for severe COVID-19," Stem Cells and Development, vol. 29, no. 12, pp. 747–754, 2020.
- [202] K. J. Juárez-Navarro, E. Padilla-Camberos, N. F. Díaz, A. Miranda-Altamirano, and N. E. Díaz-Martínez, "Human Mesenchymal Stem Cells: The Present Alternative for High-Incidence Diseases, Even SARS-Cov-2," Stem cells international, vol. 2020, Article ID 8892189, 13 pages, 2020.
- [203] F. A. Alzahrani, I. M. Saadeldin, A. Ahmad et al., "The potential use of mesenchymal stem cells and their derived exosomes as immunomodulatory agents for COVID-19 Patients," Stem Cells International, vol. 2020, Article ID 8835986, 11 pages, 2020.