

Document downloaded from:

<http://hdl.handle.net/10251/63472>

This paper must be cited as:

Figás Moreno, MDR.; Prohens Tomás, J.; Raigón Jiménez, MD.; Fita Fernández, AM.; García Martínez, MD.; Casanova Calancha, C.; Borrás, D.... (2015). Characterization of composition traits related to organoleptic and functional quality for the differentiation, selection and enhancement of local varieties of tomato from different cultivar groups. *Food Chemistry*. 187:517-524. doi:10.1016/j.foodchem.2015.04.083.



The final publication is available at

<http://dx.doi.org/10.1016/j.foodchem.2015.04.083>

Copyright Elsevier

Additional Information

1 **Characterization of composition traits related to organoleptic and functional**
2 **quality for the differentiation, selection and enhancement of local varieties of**
3 **tomato from different cultivar groups**

4

5 Running title: Composition of local tomato varieties from different cultivar groups

6

7 Maria R. Figàs ^a, Jaime Prohens ^{a,*}, María D. Raigón ^b, Ana Fita ^a, María D. García-
8 Martínez ^b, Cristina Casanova ^a, Dionís Borràs ^a, Mariola Plazas ^a, Isabel Andújar ^a,
9 Salvador Soler ^a

10 ^a Institut de Conservació i Millora de l'Agrodiversitat Valenciana, Universitat
11 Politècnica de València, Camí de Vera 14, 46022 València, Spain

12 ^b Departament de Química, Universitat Politècnica de València, Camí de Vera 14,
13 46022 València, Spain

14

15 *Corresponding author.

16 Tel.: +34 963879424; fax: +34 963879422

17 E-mail address: jprohens@btc.upv.es (J. Prohens)

18 **Abstract**

19 Tomato (*Solanum lycopersicum*) local varieties are having an increasing demand. We
20 characterized 69 local tomato accessions from eight cultivar groups for proximate
21 composition traits, major sugars, acids and antioxidants. A large diversity was found,
22 with differences among accessions of almost ten-fold for lycopene. Significant
23 differences were found among cultivar group means for most traits. The Cherry and
24 Penjar groups generally presented higher dry matter, soluble solids content, titratable
25 acidity, taste index, β -carotene, ascorbic acid, total phenolics, and antioxidant activity
26 that the other groups. Wide ranges of variation were found within each cultivar group.
27 Positive correlations were found between proximate traits related to taste and
28 antioxidants. The multivariate principal components analysis confirms the distinct
29 profile of the Cherry and Penjar groups and the large variation within groups. The
30 results will be useful for the differentiation, enhancement and selection of local tomato
31 varieties with improved organoleptic properties and functional quality.

32

33 **Keywords:** *Solanum lycopersicum*, chemical composition, local varieties, organoleptic
34 quality, functional quality, cultivar groups, selection

35

36

37 **1. Introduction**

38

39 Commercial production of tomato (*Solanum lycopersicum* L.) in developed
40 regions of the world is mostly based on modern varieties, frequently genetically uniform
41 F1 hybrids, which have a high yield, multiple resistance to diseases, and long shelf life
42 (Díez and Nuez, 2008). Despite the clear productive advantages of modern tomato

43 varieties, consumers often complain of their reduced organoleptic quality (Causse et al.,
44 2010). Low quality of many modern tomato varieties has been attributed to a tradeoff
45 between yield and concentration of compounds involved in taste, in particular when
46 cultivated under suboptimal conditions of temperature and illumination, as well as to the
47 pleiotropic effect on organoleptic quality of long-shelf life mutations, like *rin*, present in
48 many commercial hybrids of tomato (Díez and Nuez, 2008). This has increased the
49 demand for local and “heirloom” varieties of tomato, which are associated to a better
50 flavour (“flavour of the past”), to local production (Brugarolas et al., 2009), and in some
51 cases, to an increased content in bioactive compounds compared to standard long shelf
52 life varieties (Vrebalov et al., 2002). In consequence, local tomato varieties often reach
53 market prices much higher than those of standard modern varieties (Cebolla-Cornejo et
54 al., 2007; Brugarolas et al., 2009). This increased demand opens the opportunity for the
55 recovery of local tomato varieties for an expanding market.

56 Tomato is grown in all tropical, subtropical and temperate regions of the world,
57 and differences in local preferences and agroclimatic conditions, in conjunction with
58 microevolutionary forces have resulted in the accumulation of a large phenotypic
59 diversity in local varieties of tomato, with a wide diversity and array of combinations
60 for fruit size, shape, and colour (Rodríguez-Burruezo et al., 2005; Díez and Nuez,
61 2008). One of the regions with a greatest diversity in local varieties of tomato is the
62 Mediterranean region, which is considered a secondary center of diversity for this crop
63 (Terzopoulos and Bebeli, 2008; Mazzucato et al., 2008; García-Martínez et al., 2013;
64 Figàs et al., 2015), with many local tomato varieties being locally appreciated. In
65 Europe, some of these traditional varieties, like Marmande, Oxheart or San Marzano
66 among others, have also obtained widespread recognition as high quality varieties and

67 are found in markets throughout the continent (Di Gioia et al., 2010; Casals et al., 2011;
68 Ercolano et al., 2014).

69 Enhancement of local varieties of vegetables for a more economically
70 productive horticulture can be achieved using characterization, selection and breeding
71 approaches (Hurtado et al., 2014). In this respect, the characterization of composition
72 traits relevant for organoleptic and bioactive properties is of interest, as it is increasingly
73 valued by consumers. This information may allow determining composition
74 characteristics distinctive of specific local cultivar groups, the diversity for the chemical
75 composition within each group, as well as to identify accessions within each cultivar
76 group with enhanced values for the target composition traits.

77 In tomato, taste is mostly related to the content in sugars, acids and their ratio
78 (Navez et al., 1999; Causse et al., 2010; Siddiqui et al., 2015). The main sugars in
79 tomato are the monosaccharides glucose and fructose, which are usually present at
80 equimolar ratios (Beckles, 2012). Due to high activity of acid invertase, the disaccharide
81 sucrose is not detectable or present at low levels in the cultivated tomato fruit (Beauvoit
82 et al., 2014). Regarding acids, citric acid is the main organic acid acid of the tomato
83 fruit (Fernández-Ruiz et al., 2004; Siddiqui et al., 2015). Malic acid and oxalic acid,
84 which usually rank as the second and third most abundant organic acids of tomato, are
85 present at much lower levels than citric acid (Fernández-Ruiz et al., 2004).

86 The carotenoid lycopene is the most characteristic antioxidant compound of
87 tomato and is responsible of the red color of the ripe fruit (Siddiqui et al., 2015).
88 Another carotenoid of bioactive relevance present in the tomato fruit is β -carotene,
89 although its levels are normally much lower than those of lycopene (Cortés-Olmos et
90 al., 2014). Lycopene and β -carotene intake has been correlated to a reduced risk of
91 certain types of cancer and cardiovascular diseases (Riccioni, 2009; Keikel et al., 2011).

92 Ascorbic acid, which has antioxidant as well multiple biological effects beneficial for
93 human health, like antiscorbutic and anticarcinogen properties (Du et al., 2012), is also
94 present at significant levels in the tomato fruit (Cortés-Olmos et al., 2014). Phenolics, in
95 particular chlorogenic acid and quercetin, are also present in the tomato fruit in
96 significant concentrations (Siddiqui et al., 2015). Apart from the antioxidant activity
97 displayed by tomato phenolics, they are increasingly recognized as having important
98 biological properties, including anti-inflammatory, anti-microbial, neuroprotective and
99 cardioprotective effects (Del Río et al., 2013).

100 We have characterized a collection of local varieties of tomato from the
101 Mediterranean region of València (Spain) for chemical composition traits involved in
102 taste and functional quality. The collection contains accessions from different locally
103 recognized cultivar groups (Figàs et al., 2015). Chemical characterization data will
104 provide relevant information on the diversity for these traits among local tomato
105 varieties, as well as on differences among and within cultivar groups. This information
106 will be of interest for the enhancement and selection of local varieties of tomato.

107

108 **2. Material and Methods**

109

110 *2.1. Plant material*

111

112 A total of 69 accessions of local varieties of tomato from the region of València,
113 situated in the Mediterranean coast of Spain, were used for the analyses of composition.
114 The local varieties used correspond to eight cultivar groups commonly recognized in the
115 region and present different fruit characteristics (Table 1). These accessions have been
116 previously morphologically characterized using conventional descriptors and the

117 phenomics tool Tomato Analyzer (Figàs et al., 2015). Passport data on each of the
118 accessions used is included as Electronic Supplementary Material - Table S1).

119 Five plants per accession were grown during the spring-summer season of 2013
120 in an open field plot in Vila-Real (Region of València, Spain) following the standard
121 horticultural practices used in the area for local varieties of tomato. Plants were
122 distributed following a completely randomized design. Further details of plant
123 cultivation conditions can be consulted elsewhere (Figàs et al., 2015).

124

125 *2.2. Preparation of samples*

126

127 Five samples of healthy vine-ripened mature red-ripe stage fruit were used for
128 each accession. Fruits were harvested between June 17 and August 16. Each sample
129 consisted of at least three tomatoes from the second to fourth trusses, with a minimum
130 total weight for sample of 250 g. Fruits were brought to the laboratory and were washed
131 and squeezed using a domestic juice extractor. Two aliquots were obtained; one was
132 used for the immediate determination of ascorbic acid and for proximate traits
133 measurement (dry matter, soluble solids, pH, and titratable acidity), and the other was
134 frozen in liquid N₂ and stored at -80°C until analyzed for the rest of traits.

135

136 *2.3. Analytical methods*

137

138 Dry matter was calculated from a 10 ml juice sample as $100 \times dw/fw$, where fw
139 and dw are, respectively, fresh weight and dry weight after drying at 105°C to constant
140 weight. Soluble solids (SS) were measured refractometrically using a drop of juice
141 using a hand-held refractometer. pH was determined in juice using an automatic

142 pHmeter. Titratable acidity (TA) was determined potentiometrically by titrating a 100
143 ml diluted (1:5) sample of juice with 0.5 N NaOH to pH 8.1, and expressed as
144 percentage of citric acid. Taste index (TI) was determined according to Navez et al.
145 (1999) from the SS and TA values using the formula $TI=TA+(SS/(20\times TA))$. Glucose
146 and fructose were determined using the D-Fructose/D-Glucose Assay Kit (Megazyme
147 International Ltd., Wicklow, Ireland) according to the manufacturer instructions
148 (Megazyme, 2013). In order to calculate concentrations of glucose and fructose,
149 absorbances were measured at 340 nm (Megazyme, 2013) in a Jenway 6305 (Jenway,
150 Essex, UK) spectrophotometer (Megazyme, 2013). Citric acid was determined using the
151 CI9920 enzymatic kit (BEN S.r.l., Milano, Italy) according to the manufacturer
152 instructions. For lycopene and β -carotene determinations, frozen homogenate was
153 extracted overnight with ethanol:hexane (4:3 v/v) in darkness. Subsequently, the hexane
154 phase was separated and lycopene and β -carotene concentrations were determined from
155 UV/V spectrophotometry absorbance values at 503 nm (lycopene) and 450 nm (β -
156 carotene). Ascorbic acid was determined by potentiometric titration with a Titrino 702
157 (Metrohm, Herisau, Switzerland) using a Metrohm 6.0420.100 combined Pt selective
158 electrode and a 0.005 M chloramine T as standard. Total phenolics were determined
159 according to the Folin-Ciocalteu method using chlorogenic acid as standard, as
160 indicated in Raigón et al. (2008). Antioxidant activity was estimated using the
161 colourimetric DPPH (1,1-diphenyl-2-picrylhydrazyl) assay as described by Sánchez-
162 Moreno et al. (1998) and expressed as Trolox (6-hydroxy-2,5,7,8-tetramethylchroman-
163 2-carboxylic acid) equivalents (TE).

164

165 *2.4. Data analysis*

166

167 Data were subjected to analyses of variance (ANOVA) using a fixed-effects
168 model for variety. The total sums of squares for each trait was partitioned into the sums
169 of squares for accession and residual effects. The average (pooled) standard error (SE)
170 was obtained for each trait from the corresponding ANOVA. The coefficients of
171 phenotypic variation (CV_P) and genotypic variation (CV_G) for each trait were estimated
172 from the mean value and the estimates for the phenotypic variance and genotypic
173 variance obtained from the sums of squares of the ANOVA analyses (Wricke and
174 Weber, 1986). Mean values for each accession were used to perform additional
175 ANOVA analyses to detect differences among cultivar group means. In this case, the
176 total sums of squares for each trait was partitioned into the sums of squares between and
177 within groups. Significant differences among cultivar group means were estimated
178 using the Duncan multiple range test. Pearson linear coefficients of correlation (r) were
179 calculated between pairs of traits and significance ($P < 0.05$) of correlations was
180 evaluated with the Bonferroni test (Hochberg, 1988). Principal components analysis
181 (PCA) was performed for standardized composition data using pairwise Euclidean
182 distances among accessions means. Eigenvalues and percent of variance accounted for
183 each principal component as well as correlation coefficients between composition traits
184 and principal components were calculated. Statistical and PCA analyses were performed
185 with the Statgraphics Centurion XVI version 16.2.04 (StatPoint Technologies Inc,
186 Warrenton, VA, USA) software package.

187

188 **3. Results**

189

190 *3.1. Variation parameters*

191

192 The accession effect was highly significant ($P < 0.001$) for all composition traits
193 studied and represented between 37.31% (for citric acid) and 76.21% (for total
194 phenolics) of the total variance (Table 2). For all traits, except pH, fructose, and citric
195 acid, the accession effect accounted for more than 50% of the total sums of squares. The
196 average dry matter content of the collection was of 6.49%, while for soluble solids
197 content it was of 5.70%. The pH had an average value of 4.24 and titratable acidity of
198 0.46%. As a result of the soluble solids content and titratable acidity values the taste
199 index had an average value of 1.1. Fructose presented slightly higher concentration
200 values than glucose (Table 2). Citric acid concentration values were considerably lower
201 than those of sugars. Regarding antioxidants, the highest average concentration values
202 were obtained for total phenolics ($601.5 \text{ mg}\cdot\text{kg}^{-1}$), followed by ascorbic acid (197.3
203 $\text{mg}\cdot\text{kg}^{-1}$), lycopene ($36.62 \text{ mg}\cdot\text{kg}^{-1}$), and finally by β -carotene ($9.11 \text{ mg}\cdot\text{kg}^{-1}$). The
204 average antioxidant activity was of $2.27 \text{ mmol TE}\cdot\text{kg}^{-1}$. A wide range of variation was
205 found among accession means for composition traits. In this respect, for most traits
206 severalfold differences were observed between the minimum and maximum values
207 (Table 2). The traits with a larger value for the relative range (i.e., maximum/minimum
208 values) were lycopene (9.39-fold) and citric acid (6.27-fold), while those with the
209 lowest relative range were pH (1.14-fold) and taste index (1.51-fold). The lowest values
210 for the coefficients of phenotypic (CV_P) and genotypic (CV_G) variation were obtained
211 for pH, while the highest values for CV_P and CV_G were those of lycopene (Table 2).

212

213 *3.2. Differences among cultivar groups*

214

215 When considering the proximate traits, significant differences ($P < 0.05$) were
216 found among cultivar group means, except for pH (Table 3). For all traits, the within

217 groups sums of squares accounted for more than 50% of the variance. The highest
218 average dry matter contents were found for the Cherry group, followed by the Penjar
219 group; both groups presented a significantly higher dry matter than the rest of groups
220 (Table 3). For soluble solids content, the Cherry group was significantly higher than the
221 rest of groups, except Penjar; the latter in turn was significantly higher than Borseta and
222 Pruna. For titratable acidity, Cherry also presented values significantly higher than the
223 rest of varieties; and, Penjar ranked second, with values significantly higher than those
224 of Borseta and Cor. For the taste index, again the highest values were those of the
225 Cherry group, with average values significantly higher than the rest of accessions,
226 except Penjar, which had significantly higher values than Pruna (Table 3). Despite the
227 existence of significant differences among group means for four of the proximate
228 composition traits, considerable variation was found for accession means within each of
229 the groups (Table 3). In this respect, in most cases the ranges of variation for each of the
230 cultivar groups overlap with the other groups for all traits, the exception being the
231 Cherry group which does not overlap with several other groups for dry matter, soluble
232 solids, titratable acidity, and taste index (Table 3).

233 For the sugars (glucose and fructose) and acid (citric acid) measured, significant
234 differences among cultivar group means were found only for fructose (Table 3). As
235 occurred for proximate traits the within groups sums of squares accounted for more than
236 50% of the variance in all cases. For fructose, the highest values were found for the Cor
237 and Borseta, which were significantly higher than those of Cherry, Pruna, and Penjar.
238 Also, Redona presented values significantly higher than those of Cherry (Table 3). As
239 occurred for proximate traits, considerable variation was found within each of the
240 groups (Table 3). The ranges of variation for each of the cultivar groups overlap with

241 the other groups for all traits, with the exception of the Cherry group, for fructose, does
242 not overlap with the range of variation of Borseta and Cor groups (Table 3).

243 All antioxidants, as well as the antioxidant activity, displayed significant
244 differences among cultivar group means (Table 3). For β -carotene, ascorbic acid and
245 total phenolics the between groups sums of squares accounted for more than 50% of the
246 variance. The Cherry group presented the highest average values for all antioxidant
247 compounds, (Table 3). The group Penjar ranked second for β -carotene, ascorbic acid,
248 total phenolics and antioxidant activity; however it was the cultivar group with lowest
249 content in lycopene (Table 3). For the rest of groups, few significant differences were
250 found among groups. As for the proximate composition traits, sugars and citric acid,
251 large ranges of variation were found within the cultivar groups, and all the groups
252 overlapped in the range of variation, with the exception of the Cherry group, which only
253 overlapped in the range of variation with the Penjar group for all traits but lycopene
254 (Table 3).

255

256 *3.3. Correlations among traits*

257

258 A total of 26 linear correlations were significant according to the Bonferroni test
259 at a significance level of $P \leq 0.05$ (Table 4). All the significant correlations detected were
260 positive. Dry matter, soluble solids and titratable acidity were significantly
261 intercorrelated; dry matter and soluble solids were also significantly correlated with
262 taste index. Dry matter, soluble solids and titratable acidity were also significantly
263 correlated with all antioxidant traits, except lycopene (Table 4). Taste index was
264 significantly correlated with ascorbic acid, total phenolics and antioxidant activity. No
265 significant correlations were found involving pH, sugars, citric acid or lycopene.

266 Antioxidants β -carotene, ascorbic acid and total phenolics, as well as the antioxidant
267 activity, were significantly intercorrelated (Table 4).

268

269 *3.4. Principal components analysis*

270

271 The first and second components of the PCA accounted, respectively for 43.9%
272 and 11.6 % of the total variation among accession means (Table 5). The first component
273 was positively correlated with dry matter, soluble solids, titratable acidity, as well as
274 with all antioxidant traits, except lycopene (Table 5). The second principal component
275 was positively correlated with pH, ascorbic acid and antioxidant activity, and negatively
276 with soluble solids, taste index, glucose, fructose, citric acid and lycopene (Table 5).
277 The centroid values for Borseta, Cor, Plana, Pruna, Redona and Valenciana groups
278 cluster together in the PCA plot, with negative values for the first component and
279 intermediate values for the second component. Groups Penjar and Cherry present
280 positive values for the first component and higher (Penjar) and lower (Cherry) values
281 than the rest of cultivar group centroids for the second component (Fig. 1). The
282 projection of the individual accessions in the PCA plot shows that accessions, with the
283 exception of the Cherry group, accessions of the different groups are intermingled (Fig.
284 1). However, the two most representative groups (Valenciana and Penjar) present a low
285 degree of overlap. The first component separates the accessions of the Cherry group and
286 part of the Penjar group accessions, which plot in the right part of the PCA plot (with
287 first component values above 2), from the rest of accessions (Fig. 1). For the second
288 component, in general for each of the groups there is a broad dispersion. However, for
289 the Penjar and Borseta groups there are no accessions with highly negative values;

290 conversely, for the Cor group there is only one accession having a positive (although
291 low) value (Fig. 1).

292

293 **4. Discussion**

294

295 The large diversity for composition traits relevant for taste and functional quality
296 found in the collection of traditional varieties of tomato evaluated matches the wide
297 morphological variation found in the same set of accessions (Figàs et al., 2015). Our
298 data are in agreement with other studies on the diversity for chemical composition traits
299 of local varieties of tomato (Rodríguez-Burruezo et al., 2005; Labate et al., 2011;
300 Panthee et al., 2013; Cortés-Olmos et al., 2014) and reveal that local varieties are highly
301 variable for composition traits and, therefore, amenable to selection.

302 The average levels for the traits evaluated are generally comparable to other
303 tomato studies (Fernández-Ruiz et al., 2004; Rodríguez-Burruezo et al., 2005; Ilahy et
304 al., 2011; Labate et al., 2011; Panthee et al., 2013; Cortés-Olmos et al., 2014). The
305 somewhat larger values for total phenolics obtained by us compared to other works
306 (Ilahy et al., 2011; Cortés-Olmos et al., 2014) is probably due to the fact that we
307 expressed the results in equivalents of chlorogenic acid, which is the major phenolic
308 compound of tomato (Siddiqui et al., 2015), instead of the commonly used gallic acid
309 (Ilahy et al., 2011) or caffeic acid (Cortés-Olmos et al., 2014). Chlorogenic acid has an
310 antioxidant activity similar to caffeic acid (Rice-Evans et al., 1997), but it has a higher
311 molecular weight, and this may explain the higher values obtained by us. Remarkably,
312 the average taste index, which is related to the relationship between soluble solids
313 content and titratable acidity, had a value above 1, indicating that most of the local
314 varieties evaluated may be considered as tasty (Navez et al., 1999). Probably, the fact

315 that plants used for the present study were cultivated outdoors during the summer
316 season, under conditions that favour the production of tomato fruits with good quality,
317 has contributed to achieving good values for the taste index (Cebolla-Cornejo et al.,
318 2011). With the exception of pH, differences of several-fold have been found for all
319 traits studied. Even in the case of pH, the differences (0.56 units) can be considered as
320 large, as a difference of 1 unit in pH represents 10 times in the concentration of H⁺ ions.
321 The traits with largest phenotypic and genotypic variation were the contents in lycopene
322 and β-carotene, suggesting that important genetic advances can be achieved by selection
323 in the collection studied (Panthee et al., 2013). Apart from its interest for its functional
324 and bioactive properties (Roccioni et al., 2009; Keikel et al., 2011), both carotenoids are
325 determinant for the fruit color (Siddiqui et al., 2015). In particular high levels of
326 lycopene are associated to intense red fruits (Hyman et al., 2004), and selection of local
327 accessions with high content in lycopene would result in an added value (Ilahy et al.,
328 2011).

329 The Cherry and Penjar cultivar groups had an average composition profile
330 different from the rest of cultivar groups, with generally higher levels of dry matter,
331 soluble solids, titratable acidity, taste index, β-carotene, ascorbic acid, total phenolics
332 and antioxidant activity. It is well known that Cherry tomatoes normally are tastier than
333 standard regular size tomatoes (Zanor et al., 2009). The fact that the fruit size and yield
334 per plant in Cherry tomatoes is lower than for regular size tomato varieties is probably a
335 main reason for the higher concentration values for most traits observed in this cultivar
336 group (Panthee et al., 2013). Regarding the Penjar tomatoes, this cultivar group is
337 characterized by the presence of the *alc* mutation (Casals et al., 2012; Bota et al., 2014),
338 which interferes with ripening and confers a long shelf life (Vrebalov et al., 2002; Bota
339 et al., 2014). Penjar fruits, on average, have a smaller fruit size than other groups of

340 local varieties, like Borseta, Cor, Plana, Redona and Valenciana (Figàs et al., 2015).
341 Our results support previous observations indicating that the *alc* mutation, apart from
342 delaying ripening, has pleiotropic effects on the physiology of the plant and also on the
343 fruit composition (Vrebalov et al., 2002). A negative effect of the *alc* mutation in
344 composition is a reduction in the concentration of lycopene (Vrebalov et al., 2002). On
345 the other hand, the high antioxidant activity of Penjar tomatoes may play a role in its
346 extended shelf life.

347 An important diversity has been detected for the traits studied within each of the
348 cultivar groups. Other studies involving cultivar groups of local tomato varieties have
349 found similar results (Cortés-Olmos et al., 2014). In this way, with the exception of the
350 Cherry group, a complete overlap has been found among cultivar groups for all traits.
351 This indicates that the selection of accessions of each cultivar group with an improved
352 content in compounds relevant for taste and functional quality is feasible. This is of
353 great relevance for the enhancement of local varieties associated to high standards of
354 quality (Hurtado et al., 2014), as well as to identify sources of variation for breeding
355 (Rodríguez-Burruezo et al., 2005; Cortés-Olmos et al., 2014). In this respect, genotype
356 × environment (G×E) interactions may be important for the traits studied (Cebolla-
357 Cornejo et al., 2011; Panthee et al., 2013) and further work should be undertaken to
358 evaluate the extent of G×E interactions in this collection.

359 Positive intercorrelations found between dry matter, soluble solids, titratable
360 acidity, and taste index were expected, as these traits are interrelated (Labate et al.,
361 2011; Panthee et al., 2013). However, positive correlations were also found with these
362 most of these traits and β -carotene, ascorbic acid, and total phenolics. This is relevant as
363 it indicates that, in the collection studied, selection of accessions with high values for
364 traits related to improved taste will also have high content in bioactive compounds and

365 antioxidant activity. The different antioxidants, with the exception of lycopene, and the
366 antioxidant activity were also positively intercorrelated. Ascorbic acid and total
367 phenolics presented the highest correlation values with the antioxidant activity. In this
368 respect, among the antioxidants the highest concentration was found for total phenolics,
369 being followed by ascorbic acid (3-fold less on average). Given that the antioxidant
370 activities of the major tomato phenolics chlorogenic acid and quercetin is higher than
371 that of ascorbic acid (Rice-Evans et al., 1997) and the levels of carotenoids are
372 comparatively much lower than those of phenolics and ascorbic acid, our results support
373 the view that the major contributors to antioxidant activity in tomato are phenolics
374 (Toor and Savage, 2005).

375 The multivariate PCA analysis confirmed the results obtained with univariate
376 methods and provided a good separation of the Cherry tomato accessions and most of
377 the Penjar group accessions from the rest of varietal groups, which are intermingled and
378 plot in the same area of the PCA graph. This is in contrast with morphological data,
379 which provide a clear separation of the cultivar groups studied here (Figàs et al., 2015).
380 In this respect, cultivar groups of tomato usually are established by morphological traits
381 rather than for composition traits (Díez and Nuez, 2008). The results also reveal that the
382 Penjar and Valenciana cultivar groups, which are the most characteristic ones in the
383 region of València (Figàs et al., 2015), are clearly separated and have a low degree of
384 overlap in the PCA plot, indicating that both groups have different composition profiles,
385 which is important for marketing strategies (Oltman et al., 2014). In this respect, these
386 two types have different uses, with Valenciana being commonly used in salads, while
387 the fruits of Penjar are mostly used for rubbing onto bread (Casals et al., 2012; Bota et
388 al., 2014). The PCA analysis also reveals that there is a large variation in composition
389 profile among Penjar tomato accessions, which present a wide range of values for the

390 first component. This is probably associated to the fact that the Penjar tomato is a
391 conglomerate of accessions carrying the *alc* mutation introgressed in different genetic
392 backgrounds (Casals et al., 2012), rather than a group of accessions with a common
393 genetic background.

394

395 **5. Conclusions**

396

397 The collection of local varieties evaluated has been very diverse for composition
398 traits involved in taste and functional properties. Two groups (Cherry and Penjar) have
399 shown composition profiles distinct from the rest of groups, which have presented
400 similar average values for the traits studied. The large diversity found within each of the
401 groups, together with the positive correlation between taste and functional quality traits
402 indicates that there are good prospects for the selection of local varieties of each group
403 with improved composition. Our study is of interest for the enhancement of local
404 varieties of tomato as it provides information on differences among and within cultivar
405 groups for traits related to organoleptic and functional quality. These results will allow
406 the selection of local accessions of tomato with better quality and adapted to the
407 demands of consumers.

408

409 **Acknowledgements**

410

411 This work has been funded by Universitat Politècnica de València. Isabel
412 Andújar is grateful to Universitat Politècnica de València for a post-doctoral contract
413 (PAID-10-14).

414

415 **References**

416

- 417 Beauvoit, B. P., Colombié, S., Monier, A., Andrieu, M. H., Blais, B., Bénard, C.,
418 Chéniclet, C., Dieuaide-Noubhani, M., Nazaret, C., Mazat, J.P., & Gibon, Y.
419 (2014). Model-assisted analysis of sugar metabolism throughout tomato fruit
420 development reveals enzyme and carrier properties in relation to vacuole
421 expansion. *The Plant Cell*, *26*, 3222-3223.
- 422 Beckles, D.M. (2012). Factors affecting the postharvest soluble solids and sugar content
423 of tomato (*Solanum lycopersicum* L.) fruit. *Postharvest Biology and Technology*,
424 *63*, 129-140.
- 425 Bota, J., Conesa, M.A., Ochogavia, J.M., Medrano, H., Francis, D.M., & Cifre, J.
426 (2014). Characterization of a landrace collection for Tomàtiga de Ramellet
427 (*Solanum lycopersicum* L.) from the Balearic Islands. *Genetic Resources and*
428 *Crop Evolution*, *61*, 1131-1146.
- 429 Brugarolas, M., Martínez-Carrasco, L., Martínez-Poveda, A., & Ruiz, J.J. (2009). A
430 competitive strategy for vegetable products: traditional varieties of tomato in the
431 local market. *Spanish Journal of Agricultural Research*, *7*, 294-304.
- 432 Casals, J., Pascual, L., Cañizares, J., Cebolla-Cornejo, J., Casañas, F., & Nuez, F.
433 (2011). The risks of success in quality vegetable markets: Possible genetic
434 erosion in Marmande tomatoes (*Solanum lycopersicum* L.) and consumer
435 dissatisfaction. *Scientia Horticulturae*, *130*, 78-84.
- 436 Casals, J., Pascual, L., Cañizares, J., Cebolla-Cornejo, J., Casañas, F., & Nuez, F.
437 (2012). Genetic basis of long shelf life and variability into Penjar tomato.
438 *Genetic Resources and Crop Evolution*, *59*, 219-229.

439 Causse, M., Friguet, C., Coiret, C., Lépiciér, M., Navez, B., Lee, M., Holthuysen, N.,
440 Sinesio, F., Moneta, E., Grandillo, S. (2010). Consumer preferences for fresh
441 tomato at the European scale: A common segmentation on taste and firmness.
442 *Journal of Food Science*, 75, S531-S541.

443 Cebolla-Cornejo, J., Soler, S., & Nuez, F. (2007). Genetic erosion of traditional
444 varieties of vegetable crops in Europe: tomato cultivation in Valencia (Spain) as
445 a case study. *International Journal of Plant Production*, 1, 113-128.

446 Cebolla-Cornejo, J., Roselló, S., Valcárcel, M., Serrano, E., Beltrán, J., & Nuez, F.
447 (2011). Evaluation of genotype and environment effects on taste and aroma
448 flavor components of Spanish fresh tomato varieties. *Journal of Agricultural and*
449 *Food Chemistry*, 59, 2440-2450.

450 Cortés-Olmos, J., Leiva-Brondo, M., Roselló, J., Raigón, M. D., & Cebolla-Cornejo, J.
451 (2014). The role of traditional varieties of tomato as sources of functional
452 compounds. *Journal of the Science of Food and Agriculture*, 94, 2888-2904.

453 Del Río, D., Rodríguez-Mateos, A., Spencer, J. P. E., Tognolini, M., Borges, G., &
454 Crozier, A. (2013). Dietary (poly)phenolics in human health: structures,
455 bioavailability, and evidence of protective effects against chronic diseases.
456 *Antioxidant and Redox Signaling*, 18, 1818-1892.

457 Di Gioia, F., Serio, F., Buttano, D., Ayala, O., & Santamaria, P. (2010). Influence of
458 rootstock on vegetative growth, fruit yield and quality in ‘Cuore di Bue’, an
459 heirloom tomato. *The Journal of Horticultural Science and Biotechnology*, 85,
460 477-482.

461 Díez, M. J., & Nuez, F. (2008). Tomato. In J Prohens, & F Nuez (Eds.), *Handbook of*
462 *plant breeding: Vegetables II* (pp. 249-323). New York: Springer.

463 Du, J., Cullen, J. J., & Buettner, G. R. (2012). Ascorbic acid: Chemistry, biology and
464 the treatment of cancer. *Biochimica et Biophysica Acta*, 1826, 443-457.

465 Ercolano, M. R., Sacco, A., Ferriello, F., D'Alessandro, R., Tononi, P., Traini, A.,
466 Barone, A., Zago, E., Chiusano, M. L., Buson, G., Delledonne, M., &
467 Frusciante, L. (2014). Patchwork sequencing of tomato San Marzano and
468 Vesuviano varieties highlights genome-wide variations. *BMC Genomics*, 15,
469 138.

470 Fernández-Ruiz, V., Sánchez-Mata, M. C., Cámara, M., Torija, M. E., Chaya, C.,
471 Galiana-Balaguer, L., Roselló, S., & Nuez, F. (2004). Internal quality
472 characterization of fresh tomato fruits. *HortScience*, 39, 339-345.

473 Figàs, M. R., Prohens, J., Raigón, M. D., Fernández-de-Córdova, P., Fita, A., & Soler,
474 S. (2015). Characterization of a collection of local varieties of tomato (*Solanum*
475 *lycopersicum* L.) using conventional descriptors and the high-throughput
476 phenomics tool Tomato Analyzer. *Genetic Resources and Crop Evolution*, 62,
477 189-204.

478 García-Martínez, S., Corrado, G., Ruiz, J. J., & Rao, R. (2013). Diversity and structure
479 of a sample of traditional Italian and Spanish tomato accessions. *Genetic*
480 *Resources and Crop Evolution*, 60, 789-798.

481 Hochberg, Y. (1988). A sharper Bonferroni procedure for multiple tests of significance.
482 *Biometrika*, 75, 800-803.

483 Hurtado, M., Vilanova, S., Plazas, M., Gramazio, P., Andújar, I, Herraiz, F. J., Castro,
484 A., & Prohens, J. (2014). Enhancing conservation and use of local vegetable
485 landraces: the *Almagro* eggplant (*Solanum melongena* L.) case study. *Genetic*
486 *Resources and Crop Evolution*, 61, 787-795.

487 Hyman, J. R., Gaus, J., & Foolad, M. R. (2004). A rapid and accurate method for
488 estimating tomato lycopene content by measuring chromaticity values of fruit
489 puree. *Journal of the American Society for Horticultural Science*, *129*, 717-723.

490 Ilahy, R., Hdider, C., Lenucci, M. S., Tlili, I., & Dalessandro, G. (2011). Phytochemical
491 composition and antioxidant activity of high-lycopene tomato (*Solanum*
492 *lycopersicum* L.) cultivars grown in Southern Italy. *Scientia Horticulturae*, *127*,
493 255-261.

494 Keikel, M., Schumacher, M., Dicato, M., & Diederich, M. (2011). Antioxidant and anti-
495 proliferative properties of lycopene. *Free Radical Research*, *45*, 925-940.

496 Labate, J. A., Sheffer, S. M., Balch, T., & Robertson, L. D. (2011). Diversity and
497 population structure in a geographic sample of tomato accessions. *Crop Science*,
498 *51*, 1068-1079.

499 Mazzucato, A., Papa, R., Bitocchi, E., Mosconi, P., Nanni, L., Negri, V., Picarella, M.
500 E., Siligato, F., Soressi, G. P., Tiranti, B., & Veronesi, F. (2008). Genetic
501 diversity, structure and marker-trait associations in a collection of Italian tomato
502 (*Solanum lycopersicum* L.) landraces. *Theoretical and Applied Genetics*, *116*,
503 657-669.

504 Megazyme. (2008). *D-fructose and D-glucose assay procedure K-FRUCL 09/13*.
505 Wicklow, Ireland: Megazyme International Ireland.

506 Navez, B., Letard, M., Graselly, D., & Jost, J. (1999). Les critères de qualité de la
507 tomate. *Infos-Ctifl*, *155*, 41-47.

508 Oltman, A. E., Jervis, S. M., & Drake, M. A. (2014). Consumer attitudes and
509 preferences for fresh market tomatoes. *Journal of Food Science*, *79*, S2091-
510 S2097.

511 Panthee, D. R., Labate, J. A., McGrath, M. T., Breksa III, A. P., & Robertson, L. D.
512 (2013). Genotype and environmental interaction for fruit quality traits in vintage
513 tomato varieties. *Euphytica*, *193*, 169-182.

514 Riccioni, G. (2009). Carotenoids and cardiovascular disease. *Current Atherosclerosis*
515 *Reports*, *11*, 434-439.

516 Raigón, M. D., Prohens, J., Muñoz-Falcón, J. E., & Nuez, F. (2008). Comparison of
517 eggplant landraces and comercial varieties for fruit content of phenolics,
518 minerals, dry matter and protein. *Journal of Food Composition and Analysis*, *21*,
519 370-376.

520 Rice-Evans, C. A., Miller, N. J., & Paganga, G. (1997). Antioxidant properties of
521 phenolic compounds. *Trends in Plant Science*, *2*, 152-159.

522 Rodríguez-Burruezo, A., Prohens, J., Roselló, S., & Nuez, F. (2005). “Heirloom”
523 varieties as sources of variation for the improvement of fruit quality in
524 greenhouse-grown tomatoes. *The Journal of Horticultural Science and*
525 *Biotechnology*, *80*, 453-460.

526 Sánchez-Moreno, C., Larrauri, J. A., & Saura-Calixto, F. (1998). A procedure to
527 measure the antiradical efficiency of polyphenols. *Journal of the Science of*
528 *Food and Agriculture*, *76*, 270-276.

529 Siddiqui, M. W., Ayala-Zavala, J. F., & Dhua, R. S. (2015). Genotypic variation in
530 tomatoes affecting processing and antioxidant properties. *Critical Reviews in*
531 *Food Science and Nutrition*, doi 10.1080/10408398.2012.710278.

532 Terzopoulos, P. J., & Bebeli, P. J. (2010). Phenotypic diversity in Greek tomato
533 (*Solanum lycopersicum* L.) landraces. *Scientia Horticulturae*, *126*, 138-144.

534 Toor, R. K., & Savage, G. P. (2005). Antioxidant activities in different fractions of
535 tomato. *Food Research International*, *38*, 487-494.

536 Vrebalov, J., Ruezinski, D., Padmanabba, V., White, R., Medrano, D., Drake, R.,
537 Schuch, W., & Giovannoni, J. J. (2002). A MADS-box gene necessary for fruit
538 ripening at the tomato ripening inhibitor (*rin*) locus. *Science*, 296, 343-346.

539 Wricke, G., & Weber, W. E. (1986). *Quantitative genetics and selection in plant*
540 *breeding*. Berlin: Walter de Gruyter.

541 Zanor, M. I., Rambla, J. L., Chaib, J., Steppa, A., Medina, A., Granell, A., Fernie, A. R.,
542 & Causse, M. (2009). Metabolic characterization of loci affecting sensory
543 attributes in tomato allows an assessment of the influence of the levels of
544 primary metabolites and volatile organic contents. *Journal of Experimental*
545 *Botany*, 60, 2139-2154.

546 **Figure caption**

547

548 **Fig. 1.** Similarities based on fruit composition among 69 accessions of local varieties of
549 tomato represented on the two first principal components of PCA. First and second
550 components account for 43.9% and 11.6% of the total variation, respectively. The
551 different cultivar groups are represented by different symbols: Borseta (BOR; filled
552 square), Cherry (CHE; filled circle), Cor (COR; filled triangle), Penjar (PEN; filled
553 rhombus), Plana (PLA; open square), Pruna (PRU; open circle), Redona (RED; open
554 triangle), and Valenciana (VAL; open rhombus). First and second component centroids
555 for each of the cultivar groups are indicated using the group code. The continuous and
556 dashed lines encompass, respectively, accessions of the most characteristic local
557 landraces Penjar and Valenciana.

558