

Supplementary Discussion

Possible scenarios for chondrocyte and cartilage origins.

Our analysis of cartilage formation revealed deep conservation between two distantly related protostomes and vertebrates, supporting the hypothesis that a common genetic program for cartilage development exists across the Bilateria (Fig. 4c). These findings indicate that, despite the apparent convergent evolution of cartilage tissue in vertebrates, arthropods, and molluscs, there is a deep homology¹ of the genetic program for cartilage development.

The majority of protostome and deuterostome lineages lack cartilage (or cartilage-like) endoskeletal tissues. The phylogenetic distribution of cartilage among the main three branches of Bilateria shows a lack of historical continuity of this tissue type. This distribution suggests that cartilage evolved more than once within Bilateria, although the parent cell type may have persisted in the absence of differentiated cartilage. Alternatively, while less parsimonious, it is formally possible that cartilage could have evolved in the last common ancestor of Bilateria, but was then lost multiple times (again, loss of cartilaginous tissue does not necessarily imply loss of the parent cell type or the gene regulatory network (GRN)). In light of the data presented here, we see three possible scenarios for the evolution of cartilage (only scenarios 2 and 3 are shown in Fig. 4c):

1) Evolution of cartilage/chondrocyte predates the divergence of bilaterian lineages: In this scenario, the ancient genetic circuit that we refer to as the core chondrogenic GRN is causally correlated with the origin of chondrocytes. Therefore, the chondrocyte is a homologous cell type in cartilaginous endoskeletal tissues of Bilateria. Accordingly,

similarities in the structure of cartilage ECM and in the regulation of chondrogenesis are due to common descent. A significant implication of this model is that bilaterian lineages that lack cartilage must have lost this tissue type during their evolutionary history. This hypothesis predicts that homology of the core chondrogenic GRN in Bilateria underlies homology of cartilage and the chondrocyte, as a tissue and cell type, respectively. We find this hypothesis to be the least likely of the 3 proposed here.

2) *Independent origins of cartilage/chondrocyte from a homologous cell type:* An alternative to the above interpretation is that chondrocytes evolved independently and in parallel, but from a homologous progenitor cell type. Therefore cartilage (and cartilage-like) tissues in different bilaterian lineages could be sister tissue types that evolved by diversification of a common ancestral progenitor. Conservation of a homologous GRN does not necessarily imply homology at higher levels of biological organization, such as the cell type (chondrocyte) or tissue type (cartilage). It is plausible that connective tissue made from other cell types could be the source of cartilage-forming cells. For example, chondrocytes could have evolved independently from homologous (yet non-cartilaginous) fibroblast-like cells in multiple lineages of Bilateria. In this scenario, parallel evolution of chondrocytes from a fibroblast-like parent cell type involved redeployment of core GRN multiple times within Bilateria. This putative fibroblast-like cell type could still be present in bilaterian lineages that lack cartilage-like tissues. This model proposes non-homology of cartilage as a tissue type due to the lack of historical continuity of the chondrocyte as a cell type across Bilaterian. However, a homologous putative fibroblast-like parent cell type would serve as a recurrent source for convergent evolution of chondrocytes across Bilateria. A similar phenomenon could underlie other cases of convergent evolution, such as the independent origin of

specialized placental tissues and viviparity in mammals and in some reptile lineages (i.e., placental tissues may have arisen independently from homologous extra-embryonic tissues, specifically the chorioallantois)².

3) *Independent origins of cartilage/chondrocyte from non-homologous cell types:*

A third scenario for the origin of bilaterian chondrocytes involves the independent recruitment of a homologous GRN by non-homologous cell types. By contrast to scenario 2 (above), in which the GRN is recurrently activated to the same cell type, this third scenario proposes that the tissue/cell type where the GRN is activated could have been potentially different every time cartilage evolved. Accordingly, the independent evolution of cartilage implies non-homology at high levels of organization (structure and cell types) but deep homology of the chondrogenesis GRN within Bilateria. A frequently cited example of this process is the independent evolution of animal appendages, which show no evidence of homology at higher (structural) levels of organization, yet they display deep homology of the GRN that controls appendage development¹. This study provides functional molecular evidence for deep homology of the GRN controlling chondrogenesis.

Supplementary references

- 1 Shubin, N., Tabin, C. & Carroll, S. Deep homology and the origins of evolutionary novelty. *Nature* **457**, 818-823, doi:10.1038/nature07891 (2009).
- 2 Blackburn, D. G. Chorioallantoic placentation in squamate reptiles: Structure, function, development, and evolution. *Journal of Experimental Zoology* **266**, 414-430, doi:10.1002/jez.1402660508 (1993).

Supplementary Tables

Table S1. Sequence identifiers for annotated reference sequences (Ref-seq) and non-annotated expressed sequence tags (EST) in NCBI used for the collagen phylogenetic analysis.

Species name	Genbank
<i>Acyrtosiphon pisum</i>	XM_001942728
<i>Alvinella pompejana</i>	AAC35289
<i>Anopheles gambiae</i>	NT_078267
<i>Apis mellifera</i>	XM_391942, XM_393523
<i>Aplysia californica</i>	EB249446, GD229044
<i>Arenicola marina</i>	AAC47545
<i>Branchiostoma floridae</i>	ABG36939
<i>Ciona intestinalis</i>	NP_001029004, XP_004226827
<i>Cochliomyia hominivorax</i>	FG294109
<i>Daphnia pulex</i>	FE335735
<i>Epiperipatus sp.</i>	AM499446
<i>Euprymna scolopes</i>	DW282906, DW252688
<i>Haematobia irritans</i>	FD463616
<i>Haliotis discus</i>	BAA75668, BAA75669
<i>Helobdella robusta</i>	EY380843, EY365668
<i>Hirudo medicinalis</i>	FP591665
<i>Hirudo medicinalis</i>	EY479974
<i>Homo sapiens</i>	AAB69977, AAH86874, BAD92412, BAD92923
<i>Ixodes scapularis</i>	EL516416
<i>Locusta migratoria</i>	CO856142

<i>Lottia gigantea</i>	FC620367, FC564501
<i>Petrolisthes cinctipes</i>	FE776091
<i>Tribolium castaneum</i>	XM_966372

Table S2. Sequence identifiers for annotated reference sequences (Ref-seq) and non-annotated expressed sequence tags (EST) in NCBI used for the phylogenetic analysis of Sox genes.

Species name	Genbank
<i>Metaseiulus occidentalis</i>	XM_003746176, XM_003741168, XP_003746224
<i>Mus musculus</i>	XM_006529276, NP_035578, NP_035576, NP_033260
<i>Xenopus laevis</i>	NM_001087769
<i>Drosophila melanogaster</i>	NM_166792, NP_651839, NP_523739, NP_476894, NP_001014695
<i>Strongylocentrotus purpuratus</i>	NM_214472, XP_786809, XP_791983, XP_798084
<i>Nasonia vitripennis</i>	XM_001603093, XP_001604932, XP_001603143
<i>Ixodes scapularis</i>	XM_002415871, XP_002404127, XP_002415916
<i>Crassostrea gigas</i>	BQ426397
<i>Apis mellifera</i>	XP_001122996, XP_006562841, XP_006570327
<i>Anopheles gambiae</i>	XP_560125

Table S4. *S. officinalis* embryos used for each drug treatment and for DMSO controls.

A total of 72 embryos were used to test the effects of small molecule inhibitors on chondrogenesis. Treatment groups were analyzed by *ColA* and *Hh* in situ hybridizations, PCNA/ β catenin immunofluorescence and/or Masson's trichrome histological staining.

	Total <i>n</i>	Masson's	<i>ColA</i>	<i>βcatenin</i>	<i>Hh</i>	PCNA/ β catenin	Death
DMSO	21	5	4	3	3	5	1
Alsterpaullone	19	4	4	3	3	4	1
SANT-1	15	4	4	3	-	4	0
Cyclopamine	16	4	4	3	-	4	1
BIO	5	4	-	-	-	-	1
IWR-1	4	4	-	-	-	-	0
PNU	4	4	-	-	-	-	0
Total	84	29	16	12	6	17	3