

Supplementary information S1 (table) | **The role of pattern recognition receptors (PRRs) in regulating intestinal epithelial cells (IECs) and tissue homeostasis.**

PRRs and signalling adaptors in IECs		In vivo effect on IECs
TLRs	TLR1 ^{1,2}	ND
	TLR2 ^{3–7}	<ul style="list-style-type: none"> Protects IECs from injury³ Enhances M cell transport⁴ Promotes gap junction communication⁶ Increased TFF3 production⁷ Promotes tight junction integrity⁵
	TLR3 ^{8,9}	<ul style="list-style-type: none"> Promotes IEC IL-15 production and IEL cytotoxicity^{8,9}
	TLR4 ^{3,10–14}	<ul style="list-style-type: none"> Protects IECs from injury³ Promotes tumour development and growth^{12,14} Increases prostaglandin production¹¹ Promotes necrotizing enterocolitis (in neonates)^{15,16} Increases serum amyloid A production¹⁷ Enhances M cell transport⁴
	TLR5 ^{18–23}	<ul style="list-style-type: none"> Promotes chemokine and cytokine production¹⁸ Protects against apoptosis^{19,21} Limits IL-1β mediated inflammation²² Prevents bacterial overgrowth²⁰
	TLR9 ^{16,24–28}	<ul style="list-style-type: none"> Inhibits NF-κB activation and tolerizes subsequent TLR signalling²⁵ Protects against necrotizing enterocolitis (in neonates)¹⁶ Promotes Paneth cell degranulation²⁴ Promotes AMP production^{24,26} Protects IECs from injury^{16,25,27,28}
	TLR10 ²⁹	<ul style="list-style-type: none"> ND
	TLR11 ^{*30–32}	<ul style="list-style-type: none"> Prevents epithelial entry and dissemination of pathogenic <i>Salmonella</i> species^{31,32}
	TRIF ³³	<ul style="list-style-type: none"> Promotes IEC apoptosis³³
	MYD88 ^{3,34–43}	<ul style="list-style-type: none"> Protects IECs from injury^{3,38} Promotes AMP secretion^{37,41,42} Increases mucin production⁴² Promotes prostaglandin production³⁸ Promotes tumour formation and growth^{36,39} Increases IEC IL-15 to promote IEL survival³⁴ Promotes APRIL and BAFF production to promote B cell IgA production^{35,44} Increases plgR expression⁴² Promotes IEC autophagy and defence against intracellular bacteria⁴³
RLRs	TAK1 ^{45–48}	<ul style="list-style-type: none"> Prevents IEC apoptosis^{45–48} Promotes IEC proliferation^{45,48} Promotes tight junction formation⁴⁶ Limits accumulation of ROS^{46,47}
	TAB1–TAB2 ⁴⁷	<ul style="list-style-type: none"> Limits accumulation of ROS⁴⁷
	RIG-I ^{49,50}	<ul style="list-style-type: none"> ND
	MDA5 ⁵⁰	<ul style="list-style-type: none"> Promotes interferon production⁵⁰
NLRs	MAVS ^{50,51}	<ul style="list-style-type: none"> Protects IECs from injury⁵¹
	NOD1 ^{52–54}	<ul style="list-style-type: none"> Protects IECs from injury⁵³ Inhibits tumour development⁵³ Induces intestinal lymphoid tissue formation⁵² Recruits autophagy proteins⁵⁴
	NOD2 ^{54–59}	<ul style="list-style-type: none"> Recruits autophagy proteins⁵⁴ Promotes AMP production^{57,58} Promotes ROS production⁵⁹
	NLRP3 ^{60–66}	<ul style="list-style-type: none"> Inhibits tumour development^{61,64} Promotes or limits colitis (depending on circumstances)^{61–63,65} Promotes AMP production⁶⁶ Regulates commensal microbial composition⁶⁶
	NLRP6 ^{65,67–69}	<ul style="list-style-type: none"> Protects IECs from injury^{67,68} Promotes colonic wound healing⁶⁸ Inhibits tumour development^{67,68} Regulates commensal microbial composition⁶⁵ Inhibits NF-κB and MAP kinase responses to bacterial infection⁶⁹

NLRs (cont.)	NLRCA ^{70,71}	<ul style="list-style-type: none"> Inhibits tumour development and proliferation⁷⁰ Protects IECs from injury^{70,71} Protects against <i>Salmonella</i> infection⁷¹
	NLRP12 ^{72–74}	<ul style="list-style-type: none"> Limits inflammatory response to injury^{72–74} Limits tissue repair following injury⁷² Inhibits tumour formation and growth^{72–74}
	Caspase 1 ^{61,63,72,75}	<ul style="list-style-type: none"> Protects IECs from injury^{61,63,72} Inhibits tumour development^{61,72}
	Caspase 12 ⁷²	<ul style="list-style-type: none"> Inhibits epithelial repair⁷² Inhibits tumour development⁷²
PRR signalling pathways in IECs		
NF-KB Signalling	IKK1 ⁷⁶	<ul style="list-style-type: none"> Inhibits IEC apoptosis^{76,77} Promotes AMP production^{76,77} Protects IECs from injury^{77,78} Promotes tumour development^{79,80}
	IKK2 ^{76,78–82}	
	NEMO ⁷⁶	
	RelA ⁷⁷	
MAP Kinase Signalling	p38α ^{82,83}	<ul style="list-style-type: none"> Promotes goblet cell differentiation⁸³ Inhibits proliferation⁸³
IEC negative regulators of PRR signalling		
	SIGIRR ^{84,85}	<ul style="list-style-type: none"> Inhibits TLR signalling^{84,85} Limits proliferation⁸⁵ Protects IECs from injury⁸⁵ Inhibits tumour development and progression⁸⁵
	A20 ^{86–92}	<ul style="list-style-type: none"> Inhibits apoptosis^{89,90,92} Protects IECs from injury^{89,90,92} Inhibits NF-κB activation by PRRs and cytokine receptor^{86,87,89,90}
	MKP1 ⁹³	<ul style="list-style-type: none"> ND
	miR-146a ⁹⁴	<ul style="list-style-type: none"> Inhibits TLR signalling during the neonatal period⁹⁵

Bold font designates evidence for IEC-specific role based on targeted deletion or transgenic expression in murine models

* Indicates expression only in mice, and not in humans

AMP, antimicrobial peptide; APRIL, a proliferation-inducing ligand; BAFF, B cell activating factor; IEC, intestinal epithelial cell; IEL, intraepithelial lymphocyte; ND, not determine; pIgR, polymeric immunoglobulin receptor; ROS, reactive oxygen species; TFF3, trefoil factor 3; TLR, toll-like receptor;

1. Melmed, G. et al. Human Intestinal Epithelial Cells Are Broadly Unresponsive to Toll-Like Receptor 2-Dependent Bacterial Ligands: Implications for Host-Microbial Interactions in the Gut. *J. Immunol.* 170, 1406–1415 (2003).
2. Otte, J.-M., Cario, E. & Podolsky, D. K. Mechanisms of cross hyporesponsiveness to toll-like receptor bacterial ligands in intestinal epithelial cells. *Gastroenterology* 126, 1054–1070 (2004).
3. Rakoff-Nahoum, S., Paglino, J., Eslami-Varzaneh, F., Edberg, S. & Medzhitov, R. Recognition of Commensal Microflora by Toll-Like Receptors Is Required for Intestinal Homeostasis. *Cell* 118, 229–241 (2004).
4. Chabot, S., Wagner, J. S., Farrant, S. & Neutra, M. R. TLRs Regulate the Gatekeeping Functions of the Intestinal Follicle-Associated Epithelium. *J. Immunol.* 176, 4275–4283 (2006).
5. Cario, E., Gerken, G. & Podolsky, D. K. Toll-Like Receptor 2 Controls Mucosal Inflammation by Regulating Epithelial Barrier Function. *Gastroenterology* 132, 1359–1374 (2007).
6. Ey, B., Eyking, A., Gerken, G., Podolsky, D. K. & Cario, E. TLR2 Mediates Gap Junctional Intercellular Communication through Connexin-43 in Intestinal Epithelial Barrier Injury. *J. Biol. Chem.* 284, 22332–22343 (2009).

SUPPLEMENTARY INFORMATION

7. Podolsky, D. K., Gerken, G., Eyking, A. & Cario, E. Colitis-associated variant of TLR2 causes impaired mucosal repair because of TFF3 deficiency. *Gastroenterology* 137, 209–220 (2009).
8. Zhou, R., Wei, H., Sun, R. & Tian, Z. Recognition of Double-Stranded RNA by TLR3 Induces Severe Small Intestinal Injury in Mice. *J. Immunol.* 178, 4548–4556 (2007).
9. Zhou, R., Wei, H., Sun, R., Zhang, J. & Tian, Z. NKG2D recognition mediates Toll-like receptor 3 signaling-induced breakdown of epithelial homeostasis in the small intestines of mice. *Proc. Natl. Acad. Sci.* 104, 7512–7515 (2007).
10. Ortega-Cava, C. F. et al. Strategic Compartmentalization of Toll-Like Receptor 4 in the Mouse Gut. *J. Immunol.* 170, 3977–3985 (2003).
11. Fukata, M. et al. Cox-2 Is Regulated by Toll-Like Receptor-4 (TLR4) Signaling: Role in Proliferation and Apoptosis in the Intestine. *Gastroenterology* 131, 862–877 (2006).
12. Fukata, M. et al. Toll-Like Receptor-4 Promotes the Development of Colitis-Associated Colorectal Tumors. *Gastroenterology* 133, 1869–1869.e14 (2007).
13. Shang, L. et al. Toll-Like Receptor Signaling in Small Intestinal Epithelium Promotes B-Cell Recruitment and IgA Production in Lamina Propria. *Gastroenterology* 135, 529–538.e1 (2008).
14. Fukata, M. et al. Constitutive activation of epithelial TLR4 augments inflammatory responses to mucosal injury and drives colitis-associated tumorigenesis. *Inflamm. Bowel Dis.* 17, 1464–1473 (2011).
15. Lotz, M. et al. Postnatal acquisition of endotoxin tolerance in intestinal epithelial cells. *J. Exp. Med.* 203, 973–984 (2006).
16. Gribar, S. C. et al. Reciprocal Expression and Signaling of TLR4 and TLR9 in the Pathogenesis and Treatment of Necrotizing Enterocolitis. *J. Immunol.* 182, 636–646 (2009).
17. Reigstad, C. S., Lundén, G. Ö., Felin, J. & Bäckhed, F. Regulation of Serum Amyloid A3 (SAA3) in Mouse Colonic Epithelium and Adipose Tissue by the Intestinal Microbiota. *PLoS ONE* 4, e5842 (2009).
18. Rhee, S. H. et al. Pathophysiological role of Toll-like receptor 5 engagement by bacterial flagellin in colonic inflammation. *Proc. Natl. Acad. Sci. U. S. A.* 102, 13610–13615 (2005).
19. Burdelya, L. G. et al. An Agonist of Toll-Like Receptor 5 Has Radioprotective Activity in Mouse and Primate Models. *Science* 320, 226–230 (2008).
20. Vijay-Kumar, M. et al. Metabolic Syndrome and Altered Gut Microbiota in Mice Lacking Toll-Like Receptor 5. *Science* 328, 228–231 (2010).
21. Jarchum, I., Liu, M., Lipuma, L. & Pamer, E. G. Toll-Like Receptor 5 Stimulation Protects Mice from Acute Clostridium difficile Colitis. *Infect. Immun.* 79, 1498–1503 (2011).
22. Carvalho, F. A., Aitken, J. D., Gewirtz, A. T. & Vijay-Kumar, M. TLR5 activation induces secretory interleukin-1 receptor antagonist (sIL-1Ra) and reduces inflammasome-associated tissue damage. *Mucosal Immunol.* 4, 102–111 (2011).
23. Rhee, S. H., Im, E. & Pothoulakis, C. Toll-Like Receptor 5 Engagement Modulates Tumor Development and Growth in a Mouse Xenograft Model of Human Colon Cancer. *Gastroenterology* 135, 518–528.e3 (2008).
24. Rumio, C. et al. Degranulation of paneth cells via toll-like receptor 9. *Am. J. Pathol.* 165, 373–381 (2004).
25. Lee, J. et al. Maintenance of colonic homeostasis by distinctive apical TLR9 signalling in intestinal epithelial cells. *Nat. Cell Biol.* 8, 1327–1336 (2006).
26. Foureau, D. M. et al. TLR9-Dependent Induction of Intestinal α -Defensins by Toxoplasma gondii. *J. Immunol.* 184, 7022–7029 (2010).
27. Ciorba, M. A. et al. Induction of IDO-1 by immunostimulatory DNA limits severity of experimental colitis. *J. Immunol.* 184, 3907–3916 (2010).

SUPPLEMENTARY INFORMATION

28. Saha, S. et al. TLR9 Agonist Protects Mice from Radiation-Induced Gastrointestinal Syndrome. *PLoS ONE* 7, e29357 (2012).
29. Regan, T. et al. Identification of TLR10 as a Key Mediator of the Inflammatory Response to *Listeria monocytogenes* in Intestinal Epithelial Cells and Macrophages. *J. Immunol.* 191, 6084–6092 (2013).
30. Gopal, R., Birdsell, D. & Monroy, F. P. Regulation of toll-like receptors in intestinal epithelial cells by stress and *Toxoplasma gondii* infection. *Parasite Immunol.* 30, 563–576 (2008).
31. Shi, Z. et al. Toll-like Receptor 11 (TLR11) Prevents *Salmonella* Penetration into the Murine Peyer Patches. *J. Biol. Chem.* 287, 43417–43423 (2012).
32. Mathur, R. et al. A Mouse Model of *Salmonella Typhi* Infection. *Cell* 151, 590–602 (2012).
33. McAllister, C. S. et al. TLR3, TRIF, and Caspase 8 Determine Double-Stranded RNA-Induced Epithelial Cell Death and Survival In Vivo. *J. Immunol.* 190, 418–427 (2013).
34. Yu, Q. et al. MyD88-dependent signaling for IL-15 production plays an important role in maintenance of CD8 alpha alpha TCR alpha beta and TCR gamma delta intestinal intraepithelial lymphocytes. *J. Immunol. Baltim. Md* 1950 176, 6180–6185 (2006).
35. He, B. et al. Intestinal Bacteria Trigger T Cell-Independent Immunoglobulin A2 Class Switching by Inducing Epithelial-Cell Secretion of the Cytokine APRIL. *Immunity* 26, 812–826 (2007).
36. Rakoff-Nahoum, S. & Medzhitov, R. Regulation of Spontaneous Intestinal Tumorigenesis Through the Adaptor Protein MyD88. *Science* 317, 124–127 (2007).
37. Vaishnava, S., Behrendt, C. L., Ismail, A. S., Eckmann, L. & Hooper, L. V. Paneth cells directly sense gut commensals and maintain homeostasis at the intestinal host-microbial interface. *Proc. Natl. Acad. Sci.* 105, 20858–20863 (2008).
38. Brandl, K. et al. MyD88 signaling in nonhematopoietic cells protects mice against induced colitis by regulating specific EGF receptor ligands. *Proc. Natl. Acad. Sci.* 107, 19967–19972 (2010).
39. Lee, S. H. et al. ERK activation drives intestinal tumorigenesis in Apcmin/+ mice. *Nat. Med.* 16, 665–670 (2010).
40. Salcedo, R. et al. MyD88-mediated signaling prevents development of adenocarcinomas of the colon: role of interleukin 18. *J. Exp. Med.* 207, 1625–1636 (2010).
41. Vaishnava, S. et al. The Antibacterial Lectin RegIIy Promotes the Spatial Segregation of Microbiota and Host in the Intestine. *Science* 334, 255–258 (2011).
42. Frantz, A. L. et al. Targeted deletion of MyD88 in intestinal epithelial cells results in compromised antibacterial immunity associated with downregulation of polymeric immunoglobulin receptor, mucin-2, and antibacterial peptides. *Mucosal Immunol.* 5, 501–512 (2012).
43. Benjamin, J. L., Sumpter Jr., R., Levine, B. & Hooper, L. V. Intestinal Epithelial Autophagy Is Essential for Host Defense against Invasive Bacteria. *Cell Host Microbe* 13, 723–734 (2013).
44. Xu, W. et al. Epithelial cells trigger frontline immunoglobulin class switching through a pathway regulated by the inhibitor SLPI. *Nat. Immunol.* 8, 294–303 (2007).
45. Kajino-Sakamoto, R. et al. Enterocyte-Derived TAK1 Signaling Prevents Epithelium Apoptosis and the Development of Ileitis and Colitis. *J. Immunol.* 181, 1143–1152 (2008).
46. Kajino-Sakamoto, R. et al. TGF-β-Activated Kinase 1 Signaling Maintains Intestinal Integrity by Preventing Accumulation of Reactive Oxygen Species in the Intestinal Epithelium. *J. Immunol.* 185, 4729–4737 (2010).
47. Omori, E., Inagaki, M., Mishina, Y., Matsumoto, K. & Ninomiya-Tsuji, J. Epithelial transforming growth factor β-activated kinase 1 (TAK1) is activated through two independent mechanisms and regulates reactive oxygen species. *Proc. Natl. Acad. Sci.* 109, 3365–3370 (2012).

SUPPLEMENTARY INFORMATION

48. Kim, J.-Y., Kajino-Sakamoto, R., Omori, E., Jobin, C. & Ninomiya-Tsuji, J. Intestinal Epithelial-Derived TAK1 Signaling Is Essential for Cytoprotection against Chemical-Induced Colitis. *PLoS ONE* 4, e4561 (2009).
49. Mukherjee, A. et al. Retinoic Acid-induced Gene-1 (RIG-I) Associates with the Actin Cytoskeleton via Caspase Activation and Recruitment Domain-dependent Interactions. *J. Biol. Chem.* 284, 6486–6494 (2009).
50. Broquet, A. H., Hirata, Y., McAllister, C. S. & Kagnoff, M. F. RIG-I/MDA5/MAVS Are Required To Signal a Protective IFN Response in Rotavirus-Infected Intestinal Epithelium. *J. Immunol.* 186, 1618–1626 (2011).
51. Li, X.-D. et al. Mitochondrial antiviral signaling protein (MAVS) monitors commensal bacteria and induces an immune response that prevents experimental colitis. *Proc. Natl. Acad. Sci.* 108, 17390–17395 (2011).
52. Bouskra, D. et al. Lymphoid tissue genesis induced by commensals through NOD1 regulates intestinal homeostasis. *Nature* 456, 507–510 (2008).
53. Chen, G. Y., Shaw, M. H., Redondo, G. & Núñez, G. The Innate Immune Receptor Nod1 Protects the Intestine from Inflammation-Induced Tumorigenesis. *Cancer Res.* 68, 10060–10067 (2008).
54. Travassos, L. H. et al. Nod1 and Nod2 direct autophagy by recruiting ATG16L1 to the plasma membrane at the site of bacterial entry. *Nat. Immunol.* 11, 55–62 (2009).
55. Hugot, J.-P. et al. Association of NOD2 leucine-rich repeat variants with susceptibility to Crohn's disease. *Nature* 411, 599–603 (2001).
56. Ogura, Y. et al. A frameshift mutation in NOD2 associated with susceptibility to Crohn's disease. *Nature* 411, 603–606 (2001).
57. Kobayashi, K. S. et al. Nod2-Dependent Regulation of Innate and Adaptive Immunity in the Intestinal Tract. *Science* 307, 731–734 (2005).
58. Petnicki-Ocwieja, T. et al. Nod2 is required for the regulation of commensal microbiota in the intestine. *Proc. Natl. Acad. Sci.* 106, 15813–15818 (2009).
59. Lipinski, S. et al. DUOX2-derived reactive oxygen species are effectors of NOD2-mediated anti-bacterial responses. *J. Cell Sci.* 122, 3522–3530 (2009).
60. Villani, A.-C. et al. Common variants in the NLRP3 region contribute to Crohn's disease susceptibility. *Nat. Genet.* 41, 71–76 (2009).
61. Allen, I. C. et al. The NLRP3 inflammasome functions as a negative regulator of tumorigenesis during colitis-associated cancer. *J. Exp. Med.* 207, 1045–1056 (2010).
62. Bauer, C. et al. Colitis induced in mice with dextran sulfate sodium (DSS) is mediated by the NLRP3 inflammasome. *Gut* 59, 1192–1199 (2010).
63. Zaki, M. H. et al. The NLRP3 Inflammasome Protects against Loss of Epithelial Integrity and Mortality during Experimental Colitis. *Immunity* 32, 379–391 (2010).
64. Zaki, M. H., Vogel, P., Body-Malapel, M., Lamkanfi, M. & Kanneganti, T.-D. IL-18 Production Downstream of the Nlrp3 Inflammasome Confers Protection against Colorectal Tumor Formation. *J. Immunol.* 185, 4912–4920 (2010).
65. Elinav, E. et al. NLRP6 Inflammasome Regulates Colonic Microbial Ecology and Risk for Colitis. *Cell* 145, 745–757 (2011).
66. Hirota, S. A. et al. NLRP3 inflammasome plays a key role in the regulation of intestinal homeostasis. *Inflamm. Bowel Dis.* 17, 1359–1372 (2011).
67. Chen, G. Y., Liu, M., Wang, F., Bertin, J. & Núñez, G. A Functional Role for Nlrp6 in Intestinal Inflammation and Tumorigenesis. *J. Immunol.* 186, 7187–7194 (2011).

SUPPLEMENTARY INFORMATION

68. Normand, S. et al. Nod-like receptor pyrin domain-containing protein 6 (NLRP6) controls epithelial self-renewal and colorectal carcinogenesis upon injury. *Proc. Natl. Acad. Sci.* 108, 9601–9606 (2011).
69. Anand, P. K. et al. NLRP6 negatively regulates innate immunity and host defence against bacterial pathogens. *Nature* 488, 389–393 (2012).
70. Hu, B. et al. Inflammation-induced tumorigenesis in the colon is regulated by caspase-1 and NLRC4. *Proc. Natl. Acad. Sci.* 107, 21635–21640 (2010).
71. Carvalho, F. A. et al. Cytosolic flagellin receptor NLRC4 protects mice against mucosal and systemic challenges. *Mucosal Immunol.* 5, 288–298 (2012).
72. Dupaul-Chicoine, J. et al. Control of Intestinal Homeostasis, Colitis, and Colitis-Associated Colorectal Cancer by the Inflammatory Caspases. *Immunity* 32, 367–378 (2010).
73. Allen, I. C. et al. NLRP12 Suppresses Colon Inflammation and Tumorigenesis through the Negative Regulation of Noncanonical NF- κ B Signaling. *Immunity* 36, 742–754 (2012).
74. Zaki, M. H. et al. The NOD-Like Receptor NLRP12 Attenuates Colon Inflammation and Tumorigenesis. *Cancer Cell* 20, 649–660 (2011).
75. Siegmund, B., Lehr, H.-A., Fantuzzi, G. & Dinarello, C. A. IL-1 β -converting enzyme (caspase-1) in intestinal inflammation. *Proc. Natl. Acad. Sci.* 98, 13249–13254 (2001).
76. Nenci, A. et al. Epithelial NEMO links innate immunity to chronic intestinal inflammation. *Nature* 446, 557–561 (2007).
77. Steinbrecher, K. A., Harmel-Laws, E., Sitcheran, R. & Baldwin, A. S. Loss of Epithelial RelA Results in Deregulated Intestinal Proliferative/Apoptotic Homeostasis and Susceptibility to Inflammation. *J. Immunol.* 180, 2588–2599 (2008).
78. Chen, L.-W. et al. The two faces of IKK and NF- κ B inhibition: prevention of systemic inflammation but increased local injury following intestinal ischemia-reperfusion. *Nat. Med.* 9, 575–581 (2003).
79. Greten, F. R. et al. IKK β Links Inflammation and Tumorigenesis in a Mouse Model of Colitis-Associated Cancer. *Cell* 118, 285–296 (2004).
80. Vlantis, K. et al. Constitutive IKK2 activation in intestinal epithelial cells induces intestinal tumors in mice. *J. Clin. Invest.* 121, 2781–2793 (2011).
81. Zaph, C. et al. Epithelial-cell-intrinsic IKK- β expression regulates intestinal immune homeostasis. *Nature* 446, 552–556 (2007).
82. Guma, M. et al. Constitutive intestinal NF- κ B does not trigger destructive inflammation unless accompanied by MAPK activation. *J. Exp. Med.* 208, 1889–1900 (2011).
83. Otsuka, M. et al. Distinct Effects of p38 α Deletion in Myeloid Lineage and Gut Epithelia in Mouse Models of Inflammatory Bowel Disease. *Gastroenterology* 138, 1255–1265.e9 (2010).
84. Wald, D. et al. SIGIRR, a negative regulator of Toll-like receptor-interleukin 1 receptor signaling. *Nat. Immunol.* 4, 920–927 (2003).
85. Xiao, H. et al. The Toll-Interleukin-1 Receptor Member SIGIRR Regulates Colonic Epithelial Homeostasis, Inflammation, and Tumorigenesis. *Immunity* 26, 461–475 (2007).
86. Boone, D. L. et al. The ubiquitin-modifying enzyme A20 is required for termination of Toll-like receptor responses. *Nat. Immunol.* 5, 1052–1060 (2004).
87. Hitotsumatsu, O. et al. The Ubiquitin-Editing Enzyme A20 Restricts Nucleotide-Binding Oligomerization Domain Containing 2-Triggered Signals. *Immunity* 28, 381–390 (2008).
88. Turer, E. E. et al. Homeostatic MyD88-dependent signals cause lethal inflammation in the absence of A20. *J. Exp. Med.* 205, 451–464 (2008).
89. Wang, J., Ouyang, Y., Guner, Y., Ford, H. R. & Grishin, A. V. Ubiquitin-Editing Enzyme A20 Promotes Tolerance to Lipopolysaccharide in Enterocytes. *J. Immunol.* 183, 1384–1392 (2009).

SUPPLEMENTARY INFORMATION

90. Vereecke, L. et al. Enterocyte-specific A20 deficiency sensitizes to tumor necrosis factor-induced toxicity and experimental colitis. *J. Exp. Med.* 207, 1513–1523 (2010).
91. Lu, T. T. et al. Dimerization and ubiquitin mediated recruitment of A20, a complex deubiquitinating enzyme. *Immunity* 38, 896–905 (2013).
92. Rhee, L. et al. Expression of TNFAIP3 in intestinal epithelial cells protects from DSS- but not TNBS-induced colitis. *Am. J. Physiol. - Gastrointest. Liver Physiol.* 303, G220–G227 (2012).
93. Wang, J., Ford, H. R. & Grishin, A. V. NF-kappaB-mediated expression of MAPK phosphatase-1 is an early step in desensitization to TLR ligands in enterocytes. *Mucosal Immunol.* 3, 523–534 (2010).
94. Chassin, C. et al. miR-146a Mediates Protective Innate Immune Tolerance in the Neonate Intestine. *Cell Host Microbe* 8, 358–368 (2010).
95. Cherrier, M., Ohnmacht, C., Cording, S. & Eberl, G. Development and function of intestinal innate lymphoid cells. *Curr. Opin. Immunol.* 24, 277–283 (2012).