

HIV gp120- and methamphetamine-mediated oxidative stress induces astrocyte apoptosis via cytochrome P450 2E1

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HIV-1 glycoprotein 120 (gp120) is known to cause neurotoxicity via several mechanisms including production of proinflammatory cytokines/chemokines and oxidative stress. Likewise, drug abuse is thought to have a direct impact on the pathology of HIV-associated neuroinflammation through the induction of proinflammatory cytokines/chemokines and oxidative stress. In the present study, we demonstrate that gp120 and methamphetamine (MA) causes apoptotic cell death by inducing oxidative stress through the cytochrome P450 (CYP) and NADPH oxidase (NOX) pathways. The results showed that both MA and gp120 induced reactive oxygen species (ROS) production in concentration- and time-dependent manners. The combination of gp120 and MA also induced CYP2E1 expression at both mRNA (1.7 ± 0.2 - and 2.8 ± 0.3 -fold in SVGA and primary astrocytes, respectively) and protein (1.3 ± 0.1 -fold in SVGA and 1.4 ± 0.03 -fold in primary astrocytes) levels, suggesting the involvement of CYP2E1 in ROS production. This was further confirmed by using a selective inhibitor of CYP2E1, diallylsulfide (DAS), and CYP2E1 knockdown using siRNA, which significantly reduced ROS production (30–60%). As the CYP pathway is known to be coupled with the NOX pathway, including Fenton–Weiss–Haber (FWH) reaction, we examined whether the NOX pathway is also involved in ROS production induced by either gp120 or MA. Our results showed that selective inhibitors of NOX, diphenyleneiodonium (DPI), and FWH reaction, deferoxamine (DFO), also significantly reduced ROS production. These findings were further confirmed using specific siRNAs against NOX2 and NOX4 (NADPH oxidase family). We then showed that gp120 and MA both induced apoptosis (caspase-3 activity and DNA lesion using TUNEL (terminal deoxynucleotidyltransferase-mediated dUTP nick-end labeling) assay) and cell death. Furthermore, we showed that DAS, DPI, and DFO completely abolished apoptosis and cell death, suggesting the involvement of CYP and NOX pathways in ROS-mediated apoptotic cell death. In conclusion, this is the first report on the involvement of CYP and NOX pathways in gp120/MA-induced oxidative stress and apoptotic cell death in astrocytes, which has clinical implications in neurodegenerative diseases, including neuroAIDS.

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The HIV-1 envelope protein, glycoprotein 120 (gp120), mediates entry of the virus into host cells, including T cells and monocytes. Infected monocytes can traffic across the blood–brain barrier (BBB) into the central nervous system (CNS) where the virus can replicate in microglia and astrocytes.^{1,2} The neurotoxic effects of HIV-1 proteins, including gp120, can be either through direct or indirect mechanisms.³ In terms of indirect mechanisms, HIV-1 gp120 has been shown to induce the expression of proinflammatory chemokines and cytokines,^{4–6} and to induce oxidative stress in astrocytes and microglia.^{7,8} Both *in vitro* and *in vivo* models have been used to demonstrate that gp120 can directly induce apoptosis in neurons.⁹

A variety of illicit substances such as morphine, cocaine, and methamphetamine (MA) have been shown to exacerbate HIV-associated neurotoxicity.^{9–11} In the CNS, MA has been shown to increase excitotoxicity,¹² BBB damage,^{12,13} the expression of proinflammatory cytokines,¹⁴ and oxidative stress.^{13,15} Owing to its structural similarity to dopamine, MA has been shown to increase dopaminergic activity leading to increased intracellular oxidative stress and neurotoxicity.¹⁶ These findings are consistent with the protective effect of various antioxidants against MA-induced neurotoxicity.^{17–19} Thus, oxidative stress is suggested to have an important role in MA-mediated neurotoxicity. Furthermore, it is likely that concurrent HIV infection and substance abuse may cause

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Abbreviations: BBB, blood–brain barrier; CNS, central nervous system; CYP, cytochrome P450; DAS, diallylsulfide; DFO, deferoxamine; DPI, diphenyleneiodonium; FWH, Fenton–Weiss–Haber reaction; gp120, glycoprotein 120; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; H₂DC-FDA, 5-(and-6)-chloromethyl-2',7'-dichlorodihydrofluorescein diacetate; HFA, human fetal astrocytes; MA, methamphetamine; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide; NAC, N-acetyl cysteine; NOX, NADPH oxidase; OP, orphenadrine; ROS, reactive oxygen species; TUNEL, terminal deoxynucleotidyltransferase-mediated dUTP nick-end labeling; vit. C, vitamin C; vit. E, vitamin E

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enhanced neurotoxicity. We have recently shown that gp120 and MA cooperate synergistically to induce the proinflammatory cytokine IL-6 in astrocytes.¹⁴ However, it is not known whether such cooperation exists at the oxidative stress level.

Recent advances suggest the involvement of various cytochrome P450 (CYP) enzymes in neurotoxicity, perhaps as a result of production of reactive oxygen species (ROS) and/or reactive metabolites. The involvement of CYP46 in Alzheimer's disease²⁰ and CYP2D6 in Parkinson's disease²¹ illustrates the contributions of these enzymes to neurodegenerative disorders. Although the role of CYPs in drug abuse and HIV-1 neuropathogenesis is unexplored, a recent study suggests a potential role for CYP2E1 in alcohol-mediated neurotoxicity.²² Furthermore, NADPH oxidase (NOX) has been shown to induce ROS both independently and in association with CYP-mediated drug metabolism. Therefore, we hypothesize that CYP and NOX pathways are involved in HIV- and/or MA-induced neurotoxicity.

The present study was undertaken to determine whether viral protein gp120 and MA cooperates to cause enhanced oxidative stress leading to increased cell death and whether CYP is involved in gp120/MA-mediated oxidative stress and astrocyte apoptosis.

Results

HIV-1 gp120 and MA independently increase oxidative stress in astrocytes. Several studies have reported gp120-mediated induction of oxidative stress in astrocytes.^{7,8,23} In this study, we used SVGA astrocytic cells and transfected them with a plasmid containing a gp120 expression vector. Astrocytes were transfected for different lengths of time and ROS was measured using 5-(and-6)-chloromethyl-2',7'-dichlorodihydrofluorescein diacetate (H₂DC-FDA) on flow cytometer. As shown in Figure 1a, gp120 induced ROS production with a peak increase ($75.2 \pm 10.6\%$) at 24 h compared with mock-transfected cells. To confirm the effect of exogenous gp120, we used recombinant gp120IIIB protein and obtained similar results that showed an increase in ROS production as early as 6 h. The peak ROS production ($21.7 \pm 5.1\%$) was observed at 12 h of exposure (Figure 1b). The increase in ROS production was also found to be concentration-dependent as 2 nM gp120IIIB showed $22.1 \pm 3.2\%$ increase compared with the control (Figure 1c).

MA is known to induce oxidative stress in various cell types in the brain mainly via dopaminergic mechanism.^{24,25} To determine the effect of MA on ROS production, we treated the astrocytes with varying concentrations of MA. The results showed a concentration-dependent increase in ROS, with 500 μ M showing maximal ROS ($37.3 \pm 2.4\%$) at 24 h (Figure 1d). Furthermore, to test the effect of single dose of MA on ROS production, astrocytes were treated with 500 μ M MA for various lengths of time. This dose of MA was based on the blood concentrations and tissue/serum compartmentalization as reported in literatures.^{26–28} Furthermore, the binge administration of MA in the range of 250 mg–1 g has been found to produce brain concentrations of MA between 164 and 776 μ M.²⁷ As expected, MA increased ROS production in a

time-dependent manner and peak ROS production ($29.4 \pm 3.0\%$) was observed at 24 h (Figure 1e).

HIV-1 gp120 and MA additively increase oxidative stress in astrocytes. Upon demonstrating that both MA and gp120 independently induced ROS in SVGA astrocytes; we examined whether MA and gp120 interact additively or synergistically to increase oxidative stress. We treated SVGA astrocytes with 500 μ M MA and transfected with 2 μ g of gp120-plasmid followed by measurement of ROS at 24 h after treatment. Clearly, astrocytes treated with both MA and gp120 showed ROS levels ($107.1 \pm 12.9\%$) higher than either MA ($41.8 \pm 4.4\%$) or gp120 alone ($67.7 \pm 11.0\%$) (Figure 2a). Furthermore, two-factor ANOVA model suggested an additive effect rather than synergistic ($P=0.91$). To confirm the finding in primary astrocyte culture, we used 2 nM recombinant gp120IIIB protein in combination with 500 μ M MA. We observed similar results in terms of ROS production in primary human fetal astrocytes using flow cytometry ($5.9 \pm 2.5\%$ for MA, $24.3 \pm 4.0\%$ for gp120, and $54.8 \pm 13.1\%$ for MA + gp120) (Figure 2b). Similarly, the fluorescence observed using microscopy was significantly higher when cells were treated with both gp120 and MA as compared with cells treated with either agent alone in SVGA astrocytes (Figure 2c) and primary astrocytes (Figure 2d).

As oxidative stress is known to be associated with both cell death²⁹ and cell proliferation,³⁰ depending on the extent of the ROS production, we analyzed the functional consequences of MA and gp120 treatments in SVGA astrocytes. Clearly, the treatment with MA and gp120 significantly increased the level of terminal deoxynucleotidyltransferase-mediated dUTP nick-end labeling (TUNEL)-positive astrocytes when compared with MA or gp120 alone (Figure 2e). In parallel experiments, we quantified the level of cell death induced by MA and/or gp120 using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay and found that cells treated with gp120 and MA showed an additive effect and significantly higher ($P \leq 0.01$) cell death was observed ($14.1 \pm 2.9\%$) when compared with either MA ($6.8 \pm 1.5\%$) or gp120 ($8.3 \pm 1.8\%$) alone (Figure 2f). These observations indicated that the oxidative stress induced by gp120 and/or MA was lethal to astrocytes and MA and gp120 additively increase the cellular toxicity.

As the combination of MA and gp120 showed increased oxidative stress, we sought to address the molecular mechanism responsible for these effects. First, we pretreated SVGA cells with 100 μ M vitamin C (vit. C), a common antioxidant, followed by treatments with gp120 and MA. As expected, vit. C alone reduced basal levels of ROS production. However, it also abolished the ROS generated by either MA or gp120 alone, as well as MA + gp120 (Figure 3a). In addition, vit. C also abrogated cell death caused by MA and gp120 alone and also in combination (Figure 3b). Similarly, we also used vit. E, another canonical ROS quencher to confirm our results with vit. C. Surprisingly, TROLOX, a water-soluble form of vit. E, did not reduce ROS production (Supplementary Figure 1A). However, when we used vit. E in its native form (α -tocopherol), it reduced ROS generated by MA or gp120 alone, as well as MA + gp120 (Figure 3c). It also rescued the astrocytes from cell death (Figure 3d). Although *N*-acetyl

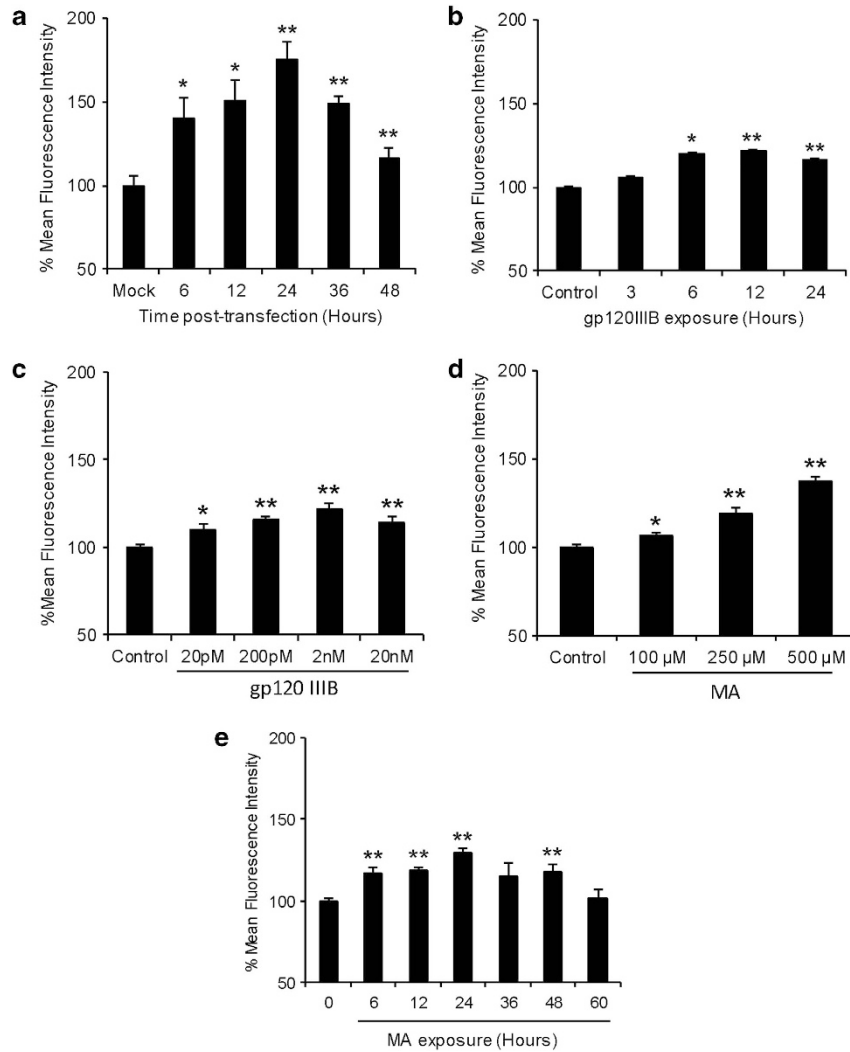


Figure 1 HIV-1 gp120 and MA increase oxidative stress in time-dependent manner. SVGA astrocytes were seeded at 2.75×10^5 per well in 12-well plates and treated with appropriate agents. The cells were then washed two times with PBS and incubated with $5 \mu\text{M}$ solution of $\text{H}_2\text{DC-FDA}$ for 30 min at 37°C . This was followed by two washes with PBS and fluorescence intensity was measured using flow cytometer. (a) Cells were transfected with $2 \mu\text{g}$ of plasmid expressing gp120 for 5 h using Lipofectamine 2000 followed by replacement of the transfection mix with complete medium. ROS was measured after 6, 12, 24, 36, and 48 h after transfection. (b) Cells were treated with $200 \mu\text{M}$ of recombinant gp120 IIIB and ROS was measured after 3, 6, 12, and 24 h after the treatment. (c) Cells were treated at varying concentrations of gp120IIIB and ROS was measured after 24 h of exposure. (d) Cells were treated with different concentrations of MA and ROS was measured after 24 h of exposure. (e) Cells were treated with $500 \mu\text{M}$ MA and ROS was measured. The bars represent mean \pm S.E. of at least two independent experiments with each treatment in triplicates. The P -value ≤ 0.05 (*) and ≤ 0.01 (**). (***) were considered statistically significant using two-tailed Student's t -test

cysteine (NAC) has been shown to abrogate gp120-mediated ROS production in astrocytes,²³ in our studies NAC did not show any reduction in ROS production (Supplementary Figure 1B). This observation is similar to our previous report, where we showed that NAC did not reduce alcohol-mediated ROS in astrocytes.²²

MA and gp120 induced the expression of CYP. As CYP has been shown to be involved in oxidative stress in many organs/tissues including the brain, we measured the mRNA expression levels of various CYPs in astrocytes treated with MA and/or gp120. Both MA and gp120 induced different isozymes of CYP at variable levels (Figure 4a). Among the isozymes induced, CYP2E1 (1.7 ± 0.2 -fold), CYP2D6 (2.3 ± 0.3 -fold), and CYP2B6 (3.2 ± 0.3 -fold) clearly showed

additive increases (two-way ANOVA showed P -value = 0.34, 0.18, and 0.84, respectively, suggesting the absence of synergy) in the levels of mRNA expression with gp120 + MA when compared with either MA (nonsignificant change for CYP2E1 and CYP2D6 and 2.8 ± 0.3 -fold for CYP2B6) or gp120 (1.3 ± 0.1 -fold for CYP2E1 and 1.8 ± 0.3 -fold for CYP2D6 and nonsignificant for CYP2B6) alone (Figure 4a). These results were further confirmed at the protein level for CYP2E1 (1.3 ± 0.1 -fold for gp120 + MA), CYP2D6 (1.7 ± 0.2 -fold for gp120 + MA), and CYP2B6 (1.4 ± 0.1 -fold for gp120 + MA) (Figure 4b). Furthermore, to confirm this phenomenon in primary astrocytes, human fetal astrocytes (HFA) were treated with gp120 IIIB and/or MA and the protein and mRNA expression levels of the CYPs were measured. Similar to SVGA astrocytes, gp120 and MA also

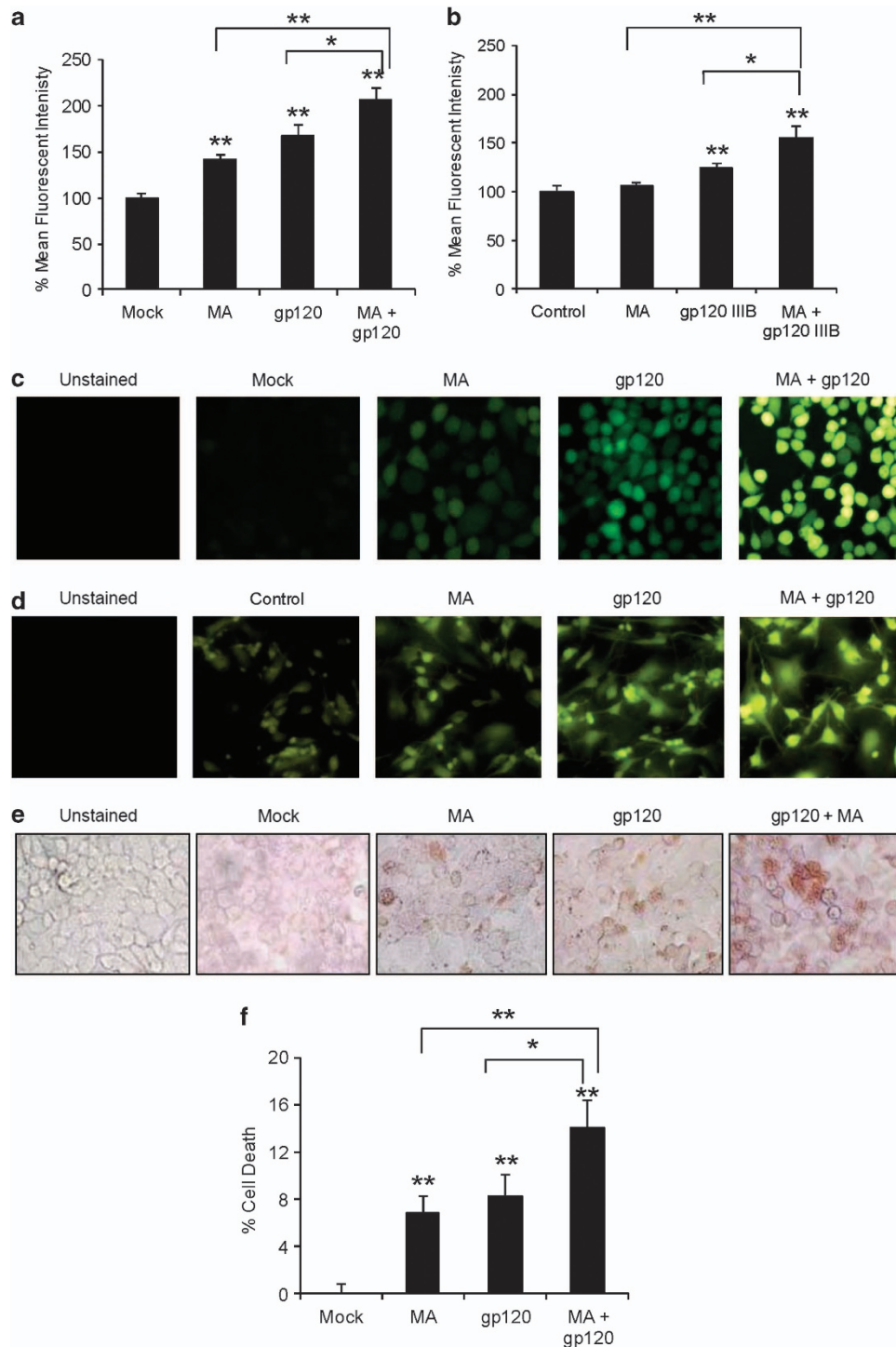


Figure 2 HIV-1 gp120 and MA additively increase oxidative stress in astrocytes. (a and c) SVGA astrocytes were seeded at 2.75×10^5 cells per well in 12-well plates and treated with or without $500 \mu\text{M}$ MA concomitantly with the transfection mix containing $2 \mu\text{g}$ of gp120-plasmid for 24 h. Mean fluorescence intensity was measured to quantify ROS production (a) and fluorescence was observed using the fluorescent microscope (c). (b and d) Primary human fetal astrocytes were seeded at 1×10^6 per well in 12-well plates and treated with either 2 nM gp120IIIIB or $500 \mu\text{M}$ MA as well as both MA and gp120IIIIB for 24 h. The ROS production was quantified using the flow cytometer (b) and fluorescence was observed using the fluorescent microscope (d). SVGA astrocytes treated with MA and/or gp120 were stained for TUNEL (e) or treated with MTT (f) to measure cell death. For MTT assay, cell viability in mock-transfected cells was considered as the control and cell death was expressed as % death in control. The bars represent mean \pm S.E. of at least three independent experiments with each treatment in triplicates. The P -value ≤ 0.05 (*) and ≤ 0.01 (**) were considered statistically significant using Student's t -test and two-way analysis of variance (ANOVA)

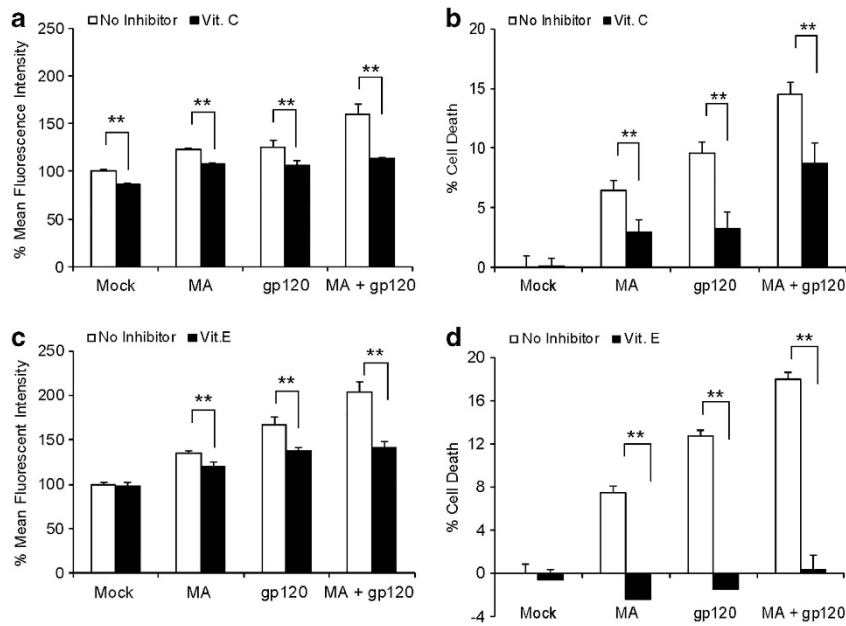


Figure 3 ROS quenchers reduced MA ± gp120-induced oxidative stress and cell death. SVGA astrocytes were treated with or without vit. C and vit. E 1 h before the treatment with MA and/or gp120. ROS (a and c) was measured using the flow cytometer 24 h after the treatment and the effect on cell death (b and d) was measured 48 h after the treatment. The bars represent mean ± S.E. of at least three independent experiments with each treatment in triplicates. The P -value ≤ 0.01 (**) were considered significant using two-tailed Student's t -test and two-way analysis of variance (ANOVA)

showed additive increase in the levels of CYP2E1, CYP2B6, and CYP2D6 mRNA (Figure 4c) and protein (Figure 4d) in HFA primary cells. The analysis run for synergy using two-way ANOVA showed nonsignificant P -values, suggesting no synergistic interaction.

Role of CYP2E1 in gp120- and/or MA-mediated oxidative stress. To examine the role of CYP2E1 in oxidative stress, we pretreated astrocytes with different concentrations of diallylsulfide (DAS), a selective inhibitor of CYP2E1. Clearly, both 10 and 25 μ M DAS reduced the ROS generated by either MA and/or gp120 (Figure 4e). It is of note that DAS alone showed some decrease in the basal ROS production, but its efficacy against MA- and/or gp120-mediated oxidative stress was significant and dose-dependent (P for trend = 0.01). Although DAS did not significantly rescue the cell from death caused by MA alone, it significantly rescued the cell death ($P \leq 0.01$) caused by gp120 alone and MA + gp120 (Figure 4f). In parallel experiments, DAS was also found to reduce the TUNEL staining in the cells treated with MA and/or gp120 (Figure 4g). Furthermore, we also specifically knocked down CYP2E1 using siRNA in order to confirm our findings with chemical inhibitor. As expected, the knockdown of CYP2E1 resulted into reduction of ROS (Figure 4h) and it also showed reduced TUNEL staining in the cells treated with MA and/or gp120 (Figure 4i). The specific knockdown of CYP2E1 was confirmed using western blotting, which showed 75–80% knockdown of CYP2E1 (Figure 4h). These results clearly suggest the involvement of CYP2E1 in oxidative stress and cell death induced by gp120, MA, or both. We also tested fluoxetine and paroxetine, specific inhibitors for CYP2D6, and orphenadrine (OP),

specific inhibitor for CYP2B6, on MA + gp120-mediated ROS generation. However, these antagonists failed to reduce the ROS formation, rather paroxetine and OP further increased oxidative stress (Supplementary Figure 2). In addition, we also specifically knocked down CYP2B6 and CYP2D6 using siRNA. However, as observed with chemical inhibitors, these siRNAs did not show any effect on ROS production (Supplementary Figure 3). Although, oxidative stress is one of the functional consequences of CYP enzyme activity, the possible role of CYP2D6 and CYP2B6 in other physiological functions cannot be disregarded.

NOX and metal chelation are involved in MA/gp120-mediated oxidative stress and cell death. The NOX family of enzymes are responsible for the transfer of electrons across biological membranes and generation of ROS.³¹ In addition, various NOXs also serve as essential coenzymes coupled with CYP2E1-mediated electron transfer in the generation of ROS.³² To examine the involvement of NOX in gp120 ± MA-mediated oxidative stress, we treated the SVGA astrocytes with various concentrations of diphenyleneiodonium (DPI), an inhibitor for NOX. DPI significantly reduced the ROS production observed with either MA or gp120 alone and MA + gp120 (Figure 5a) in a dose-dependent manner (P for trend = 0.01). Furthermore, 25 nM DPI rescued the cell death induced by MA and/or gp120 (Figure 5b), thereby confirming the role of NOX in MA/gp120-mediated cell toxicity. Among the NOX family of enzymes, NOX2 and NOX4 isozymes are predominantly responsible for NOX-derived ROS in astrocytes.^{33,34} Therefore, we knocked down NOX2 and NOX4 using siRNA, which also abrogated the ROS produced by MA and gp120, either alone

or in combination (MA ± gp120) (Figures 5c and d). Furthermore, control siRNA-transfected cells did not show any significant change in the ROS production, when compared with no-siRNA control for the respective treatment groups ($P > 0.3$ when measured using two-way ANOVA).

Superoxides ($O_2^{\cdot -}$) generated via NOX are converted into H_2O_2 , which is further converted into tertiary effector species such as hydroxyl radical ($\cdot OH$) via the Fenton–Weiss–Haber (FWH) reaction.^{35,36} We therefore hypothesized that FWH reaction is a downstream mechanism of NOX-mediated ROS

production. As increased expression of ferritin is an indicator of oxidative stress,³⁷ we measured the expression of ferritin heavy chain in the cells treated with MA and/or gp120. We observed higher ferritin expressions in astrocytes treated with MA and/or gp120 than untreated controls (Figure 6a), which suggested the involvement of $Fe^{+2} \leftrightarrow Fe^{+3}$ cycle. Therefore, we treated astrocytes with various concentrations of deferoxamine (DFO), an inhibitor of FWH reaction, 1 h before the treatment with either MA or gp120 alone and MA + gp120. Among various doses used, 50 nM DFO was found to reduce

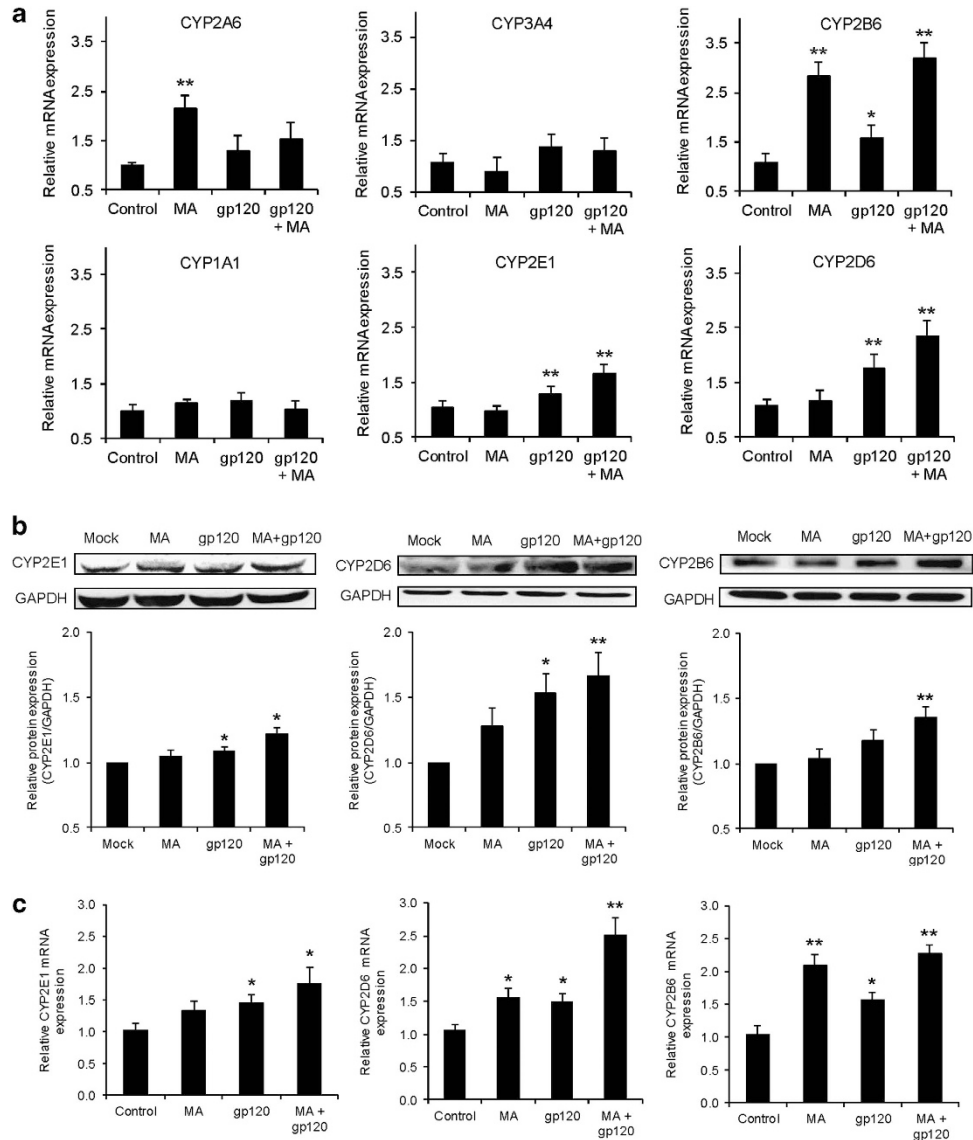


Figure 4 MA and gp120 induced the expression of various isozymes of CYP, and CYP2E1 played an important role in oxidative stress. (a and b) SVGA astrocytes were treated with or without 500 μM MA and transfected with gp120 for 6 h after transfection. Upon termination of the treatments, mRNA was isolated for its expressions for various isozymes of CYP and was measured (a) as described in Materials and Methods. Similarly, cells harvested 12 h post-treatment protein was isolated and the levels of CYPs were measured using western blotting (b), with GAPDH used as the housekeeping gene. (c and d) Primary human fetal astrocytes were treated with MA and/or gp120 for 6 or 12 h and expressions of CYPs were measured at the levels of mRNA (c) and protein (d), respectively. (e–g) The effect of DAS, a selective CYP2E1 inhibitor, on ROS production (e), cell death (f), and TUNEL labeling (g) in astrocytes treated with MA and/or gp120. (h–i) The effect of small interfering RNA (siRNA)-mediated knockdown of CYP2E1 on ROS (h) and TUNEL labeling (i) in MA- and/or gp120-treated SVGA astrocytes. The ROS production and cell death were compared with untreated control that was normalized at 100%. Bar graphs show mean of at least three independent experiments. The blot is representative of three independent experiments. GAPDH was used to normalize the expressions of CYPs. The bars represent mean \pm S.E. of at least three independent experiments with each treatment in triplicates. The P -value ≤ 0.05 (*) and ≤ 0.01 (**) were considered statistically significant using two-tailed Student's t -test and two-way analysis of variance (ANOVA)

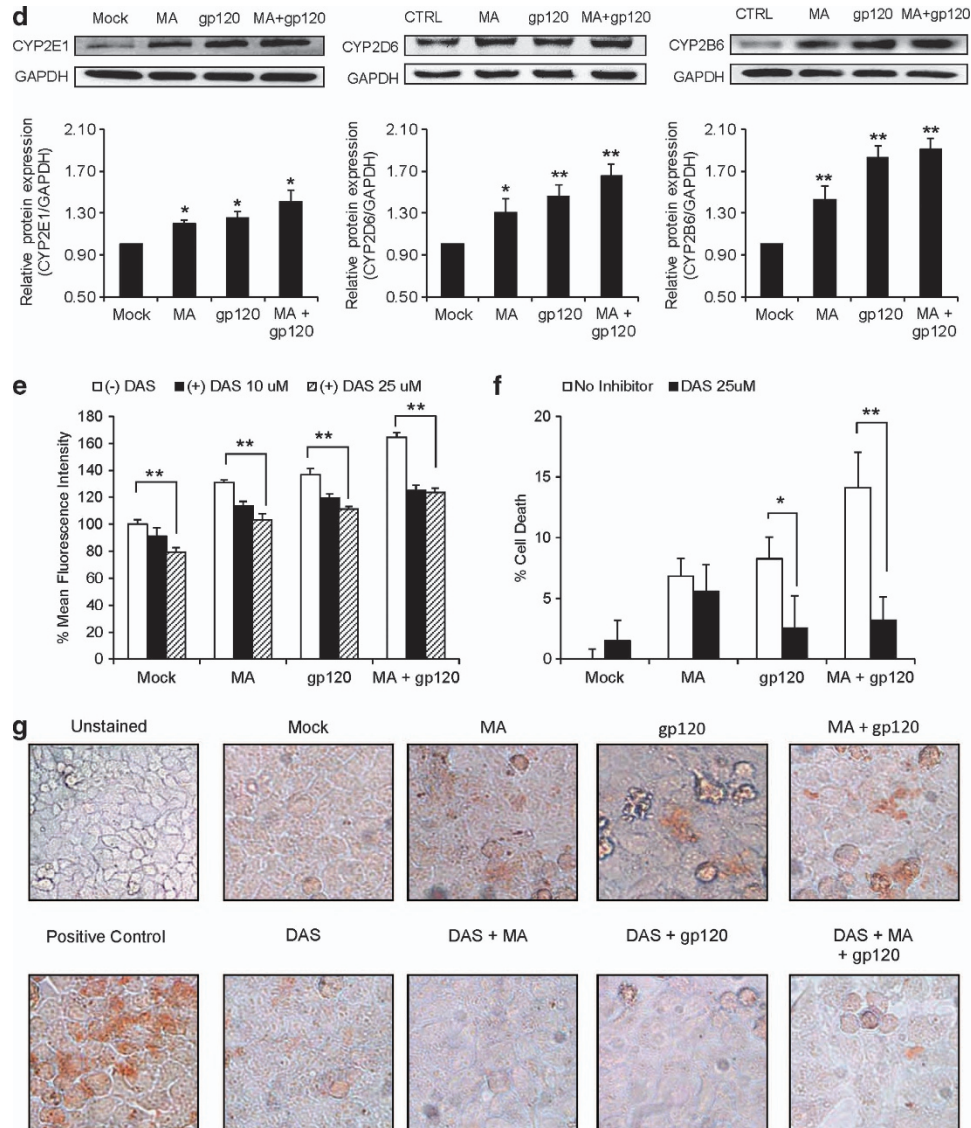


Figure 4 (Continued)

the ROS generated by gp120 alone as well as gp120 + MA (Figure 6b). Furthermore, 50 nM DFO also rescued the astrocytes from cell death induced by oxidative stress (Figure 6c). The involvement of iron cycle can also lead to increased protein carbonylation, which can further lead to apoptotic cell death.^{38,39} Therefore, we measured the levels of carbonylated protein in the astrocytes treated with MA and/or gp120. As expected, we observed increased levels of protein carbonylation (Figure 6d) upon treatments. Overall, these data suggested that NOX2 and NOX4 produced superoxides, which further underwent FWH chemistry to produce peroxides, leading to increased protein carbonylation. Taken together, these resulted in increased oxidative stress and consequently cell death.

Caspase-3 is involved in MA- and/or gp120-mediated apoptosis in astrocytes. Involvement of caspase-3 is a

classical mechanism of apoptosis in a variety of cells. We therefore examined the effect of MA and/or gp120 on caspase-3 protein expression and activity. Our results show increased protein expressions of caspase-3 when astrocytes were treated with MA and/or gp120. The immunoblots showed significantly increased expression of cleaved caspase-3 (17 kDa) in the cells treated with MA, gp120, and MA + gp120. However, procaspase-3 isoform (35 kDa) was slightly reduced in the cells treated with gp120 + MA (Figure 7a). Furthermore, astrocytes treated with either MA or gp120 alone showed increased caspase-3 cleavage activity as compared with untreated cells. In addition, cells treated with gp120 and MA showed significantly higher ($P \leq 0.01$) caspase-3 cleavage activity as compared with either of the agents alone and the effect was found to be additive (Figure 7b). Furthermore, the inhibitors for ROS (Figure 7c), CYP2E1 (Figure 7d), NOX (Figure 7e), and FWH

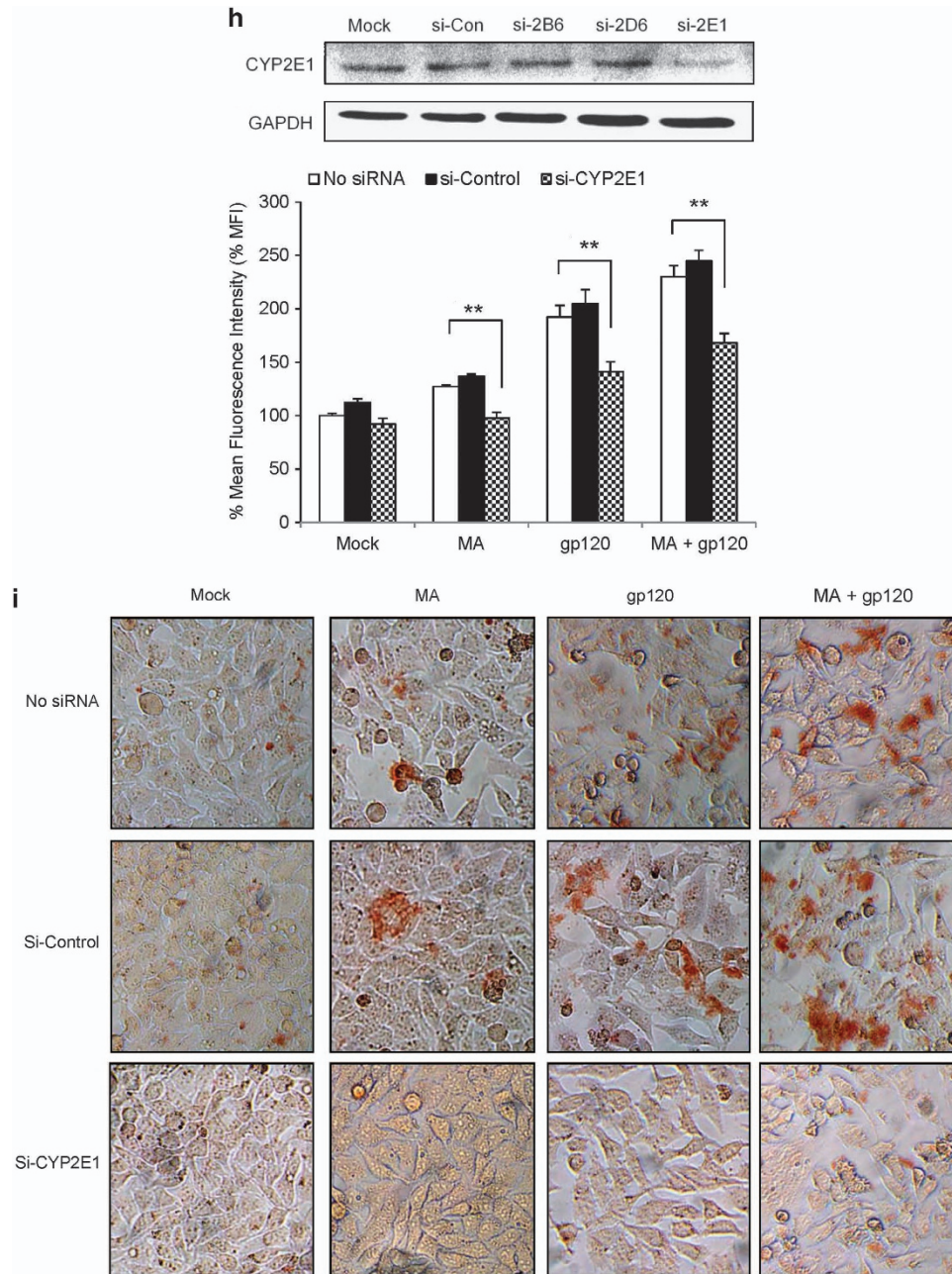


Figure 4 (Continued)

reaction (Figure 7f) significantly reduced MA/gp120-mediated caspase-3 activity. These findings clearly suggested that pathways involving CYP2E1, NOX family of enzymes, and FWH reaction are responsible for MA- and/or gp120-mediated apoptosis in SVGA astrocytes.

Discussion

Substance abuse such as MA is thought to be an additional risk factor in worsening the HIV-associated neurodegeneration.⁹ Viral proteins, such as gp120, have been shown to interplay cooperatively with MA and damage BBB integrity in mouse brain.¹⁷ In addition, MA alone is also known to induce

behavioral impairment in gp120 transgenic mice.⁴⁰ However, there is no direct evidence that elucidates the mechanism responsible for the combined effect of MA- and gp120-induced toxicities in the brain. We have recently shown that MA and gp120 can induce proinflammatory cytokine IL-6 via common signaling mechanism involving PI3K/Akt and NF- κ B pathways.^{6,41} On the other hand, an increasing body of literatures clearly suggests oxidative stress as one of the mediators of neurotoxicity induced by HIV-1 gp120 as well as MA independently. However, no attempt has been made to determine the fate of oxidative stress in a possible situation where HIV-infected individuals were MA user as well. This study was undertaken with the idea of determining whether

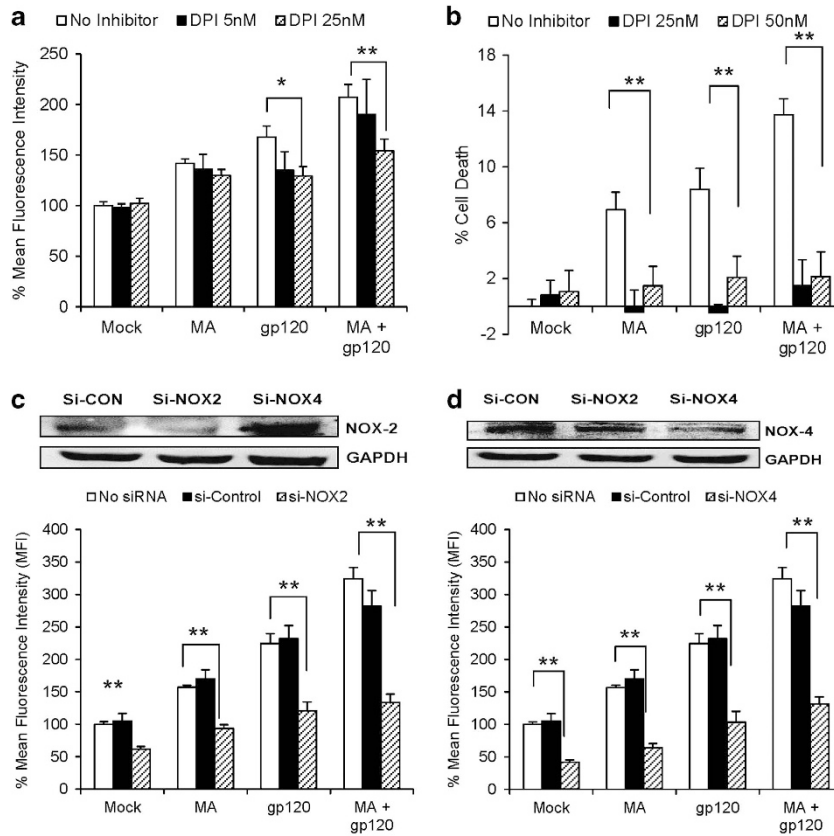


Figure 5 Role of NOX in MA- and/or gp120-induced oxidative stress and cell death. (a and b) SVGA cells were treated with DPI, a selective inhibitor for NOX, at various concentrations 1 h before the treatment with MA and/or gp120 and the effect was observed on ROS production (a) and cell death (b). (c and d) Astrocytes were transfected with 10 nM small interfering RNA (siRNA) targeting either NOX2 or NOX4 subunit of NOX family for 48 h. These cells were further treated with MA and/or gp120. The effect of gene knockdown on ROS production was measured for NOX2 (c) and NOX4 (d). The gene knockdown was confirmed using western blotting for the efficiency of each siRNA. The cells transfected with 10 nM of control siRNA was used as control, which showed no significant change in ROS production. The bars represent mean \pm S.E. of at least three independent experiments with each treatment in triplicates (except DPI 5 nM in (a) and DPI 25 nM in (b)). The P -value ≤ 0.05 (*) and ≤ 0.01 (**) were considered statistically significant using two-tailed Student's t -test and multiple analysis of variance (ANOVA)

gp120 and MA-mediated oxidative stress would cause increased apoptosis in astrocytes and whether CYP450 has any role in oxidative stress.

Oxidative stress is associated with cell toxicity in various neurodegenerative disorders such as Alzheimer's disease and Parkinson's disease.⁴² In addition, several reports have highlighted the role of oxidative stress in the pathology of HIV-associated neurocognitive disorders (HAND).⁴³ This was further confirmed by our studies with morphine, wherein we showed morphine-mediated increased virus replication,⁴⁴ accelerated disease progression,⁴⁵ and increased oxidative stress^{46,47} in macaque model of HIV/AIDS. The oxidative stress produced by either MA or gp120 has also been shown to have neurotoxic potential via various intermediates. MA, in particular, is found to increase oxidative stress predominantly via dopaminergic and glutamatergic mechanisms.¹⁰ The dopaminergic toxicity observed with MA is attributed to its structural similarity with dopamine. Furthermore, gp120 is shown to increase oxidative stress via glutathione and lipid peroxidation.^{8,48} Concurrent with the existing literatures, we also observed increased oxidative stress in astrocytes treated

with gp120 and MA individually, in time- and concentration-dependent manners.

Recently, dopamine has been shown to regulate the expression of various CYP isozymes in the liver.⁴⁹ Furthermore, Bromek *et al.*⁵⁰ has shown that CYP2D increases dopamine formation in the brain. In addition to the metabolism of xenobiotics including substances of abuse, the role of CYPs in oxidative stress is unequivocally accepted in various tissues/organs including the brain.⁵¹ Specifically, the roles of CYP2E1 for alcohol metabolism-mediated toxicity and CYP2A6²² for tobacco/nicotine metabolism-mediated toxicity⁵² have been implicated through the oxidative stress pathway in many tissues/organs. Our group along with others has previously reported that astrocytes also express many CYPs at variable levels, and they have demonstrated the role of CYPs in alcohol- and nicotine-mediated oxidative stress and toxicities.^{52,53} Similarly, MA has been shown to induce mRNA expression of CYP2E1 and activity of CYP2C6 in rats.^{54,55} Our observations with regard to increased expressions of CYP2A6, 2B6, and 2D6 by MA and CYP2E1, 2B6, and 2D6 by gp120 are important to elucidate the roles of these

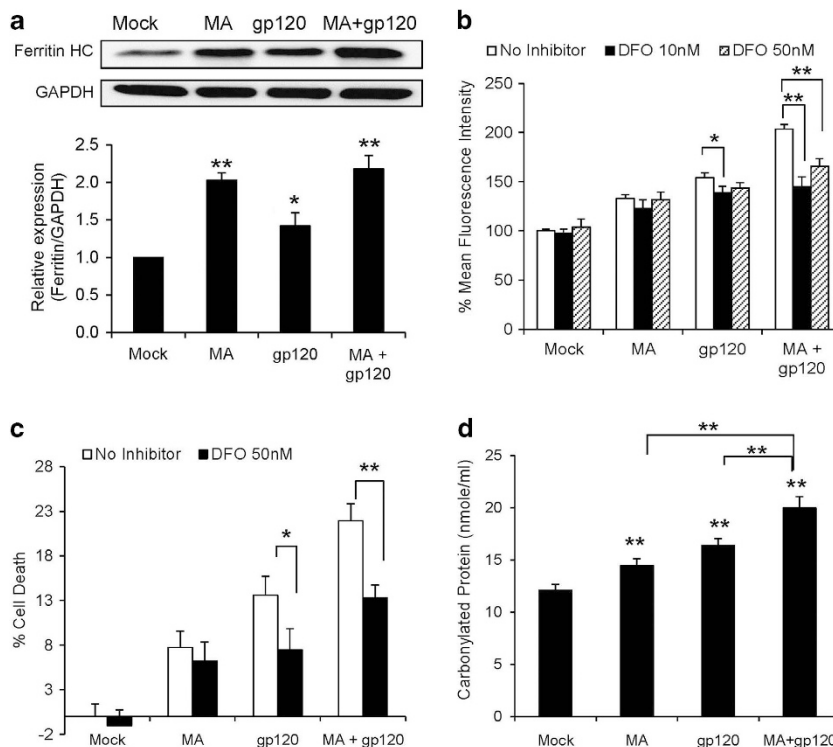


Figure 6 The expression of ferritin and the role of FWH chemistry in MA- and/or gp120-mediated oxidative stress and cell death. SVGA astrocytes were treated with MA and/or gp120 and the expressions of ferritin HC were measured using western blotting (a) and representative image is shown. Cells were treated with varying concentrations of DFO, an iron chelator used as FWH reaction antagonist, for 1 h before the treatment with MA and/or gp120 and the effect was observed on ROS (b) and cell death (c). (d) The amount of carbonylated proteins after 24 h as a result of treatment with MA and/or gp120. The ROS production and cell death were compared with untreated control that was normalized at 100%. The bars represent mean \pm S.E. of at least three independent experiments with each treatment in triplicates. The P -value ≤ 0.05 (*) and ≤ 0.01 (**) were considered statistically significant using two-tailed Student's t -test and multiple analysis of variance (ANOVA)

CYPs in oxidative stress, metabolism of certain xenobiotics, and homeostasis of other resident molecules such as dopamine in the brain. More importantly, the additive increase in the expressions of CYP2D6 and CYP2E1 by gp120 + MA is intriguing, which suggests a possibility that CYPs might have been involved in oxidative stress. This is the first evidence that shows an additive effect between MA or gp120 and CYP expression in the CNS.

Amphetamine analogs including MA are known to interact with CYP2D6 and alters its activity,⁵⁶ which is further supported by the finding that selective inhibitors of CYP2D6 reduces the metabolism of ecstasy (MA analog) *in vitro*.⁵⁷ However, the role of CYP2E1 in the metabolism of MA is largely unknown, except a few recent reports that have shown an involvement of CYP2E1 and CYP2B in the metabolism of either MA or pyrolysis product of MA.^{58,59} CYP2E1 is mainly known for the metabolic activity in the presence of ethanol; however, its involvement in MA-mediated production of ROS leading to increased cell death is not known. We are able to confirm involvement of CYP2E1 in oxidative stress because DAS, a selective inhibitor for CYP2E1, significantly abrogated not only gp120/MA-mediated production of ROS but also cell death caused by these two stimulants individually or in combination. Unlike CYP2E1 inhibitor, the CYP2D6 and CYP2B6 inhibitors increased MA/gp120-induced ROS, suggesting lack of its role in MA/gp120-mediated oxidative stress. The increased oxidative stress by CYP2D6 inhibitors

has been observed before and has been attributed to the direct effect of these inhibitors.⁶⁰

The interaction of various CYPs with NADPH system is tightly regulated by NOX enzymes.⁶¹ CYP2E1 is specifically involved in the metabolism of R-OH compounds into aldehydes, which is coupled with the conversion of NADPH to NADP⁺ and the released electron is consumed in the process. This NADP⁺ is converted via non-CYP mechanism into NADPH, which is recycled back to NADPH by NOX family of enzymes (Figure 8). The NOX family of enzymes, particularly NOX2 and NOX4, is potentially known to increase ROS in astrocytes independently as well as in association with FWH chemistry.^{31,36} Furthermore, the superoxides and peroxides generated via NOX enzymes can result in increased expressions of ferritin and protein carbonylation³⁸ as shown in this study as well. The role of NOX in substance abuse such as alcohol⁶² and MA¹⁸ is known in neuronal toxicity. Furthermore, the NOX enzymes have been implicated in therapeutic intervention of CNS disorders such as strokes and Alzheimer's disease.^{63,64} Our observations with NOX2 and NOX4 knockdown to rescue cells from oxidative damage support the possibility of using NOX as a target for developing potential therapeutics for the treatment of neuroAIDS, especially among MA users.

Various free radicals, generated during oxidative stress, may lead to multifactorial physiological effects in the cells via DNA damage, lipid peroxidation, or activation of apoptotic

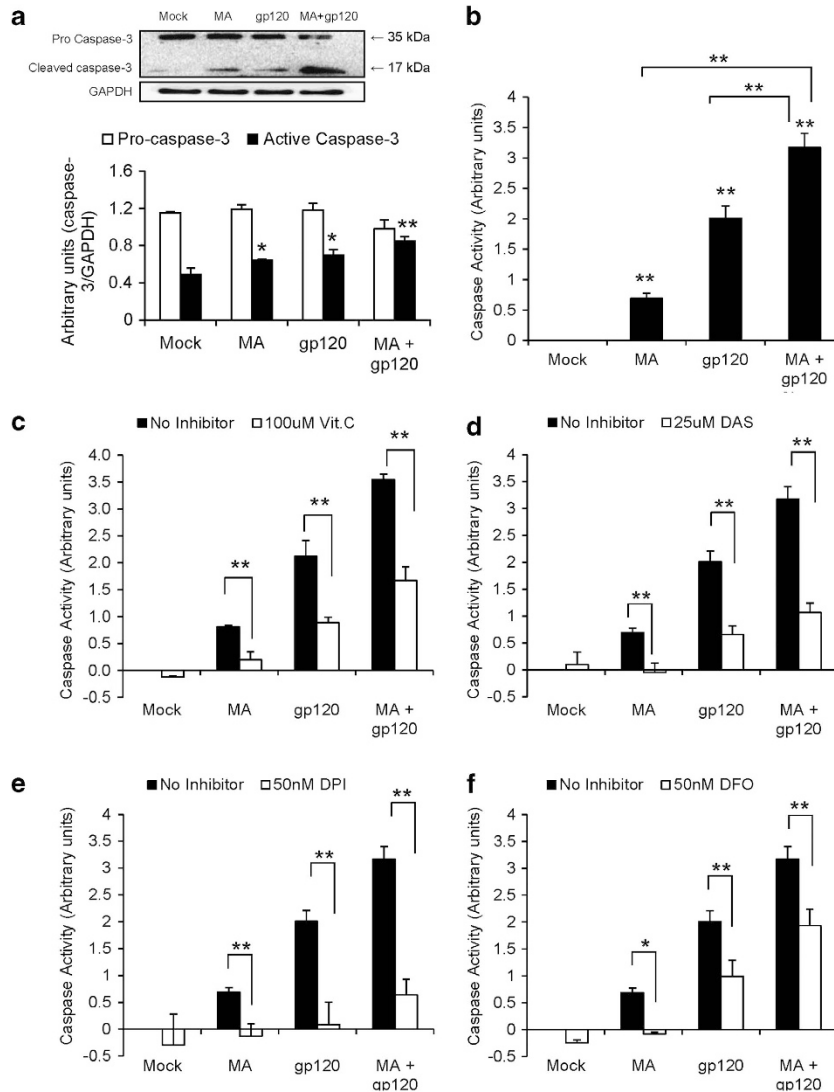


Figure 7 Caspase-3 activation is involved in the MA and/or gp120-mediated apoptosis in astrocytes. (a and b) SVGA cells were treated with MA and/or gp120 for 24 h and the cytoplasmic proteins were obtained, which were used to measure procaspase-3 (35 kDa) and cleaved caspase-3 (17 kDa) protein expressions (a) and caspase-3 cleavage activity (b). The cells were also treated with either 100 μ M vit. C (c), 25 μ M DAS (d), 50 nM DPI (e), or 50 nM DFO (f) 1 h before the treatment with MA and/or gp120 and the effect on caspase-3 cleavage activity was measured. The activity was normalized with basal levels in untreated control and reported as change in the activity. The bars represent mean \pm S.E. of three independent experiments with each treatment in triplicates. The *P*-value ≤ 0.05 (*) and ≤ 0.01 (**) were considered statistically significant using two-tailed Student's *t*-test and multiple analysis of variance (ANOVA). The significant values shown above the bars were calculated using Student's *t*-test for comparison between the treatment and mock-transfected control

mechanisms. The ROS-mediated BBB damage has been shown to cause lipid peroxidation and loss of tight junction protein.^{13,17} MA and gp120 in combination are specifically shown to alter the levels of tight junction proteins to create a 'leaky' BBB, which may enhance the invasion of infected monocytes into the brain.¹⁷ Similarly, cocaine in association with gp120 has also been shown to increase ROS, which involves caspase-3 and NF- κ B activation to induce apoptosis in astrocytes.¹¹ In our prior studies, we have shown an overlapping mechanism between MA and gp120 involving NF- κ B activation.⁴¹ We have further shown the induction of proinflammatory cytokines as a result of NF- κ B activation. In the present study, we observed an increase in the expression of cleaved caspase-3 protein in MA and/or gp120, which were

further confirmed with the increase in the caspase-3 cleavage activity. Our results are consistent with the recent finding that apoptosis is involved in gp120-mediated toxicity.^{23,65} Furthermore, caspase-3 cleavage activity was inhibited by selective inhibitors for CYP2E1, NOX, and FWH reaction. Taken together, these findings suggest that the increase in caspase-3 expression and activity via ROS and cytokines via NF- κ B activation may lead to toxicity in astrocytes. Thus, there is a possibility of a cross-talk between these two mechanisms and therefore further studies are needed to investigate the effect of CYP-mediated ROS on cytokine production or *vice versa*. Our TUNEL staining was also consistent with this notion as both MA and gp120 independently as well as in combination led to increased DNA damage. Our observations with MTT assay

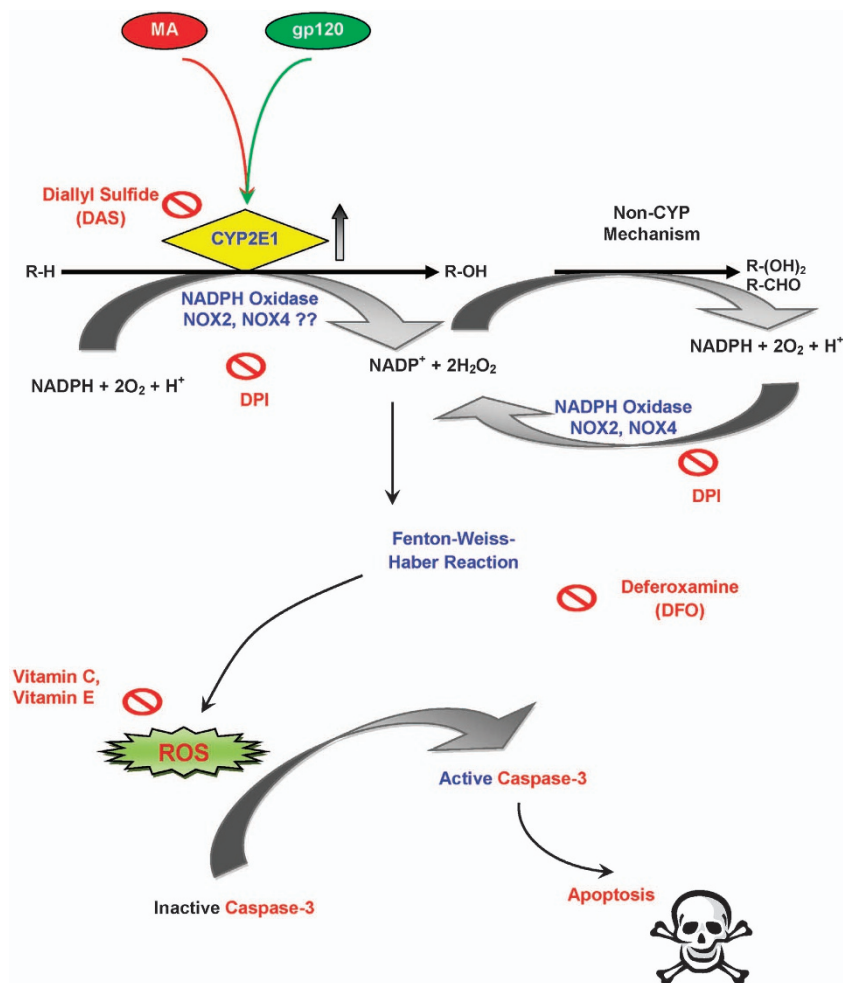


Figure 8 Schematic illustration of MA- and/or gp120-mediated oxidative stress. MA and/or gp120 increase the expressions of CYP2E1, which generates free electrons via conversion of NADPH to NADP⁺. The CYP2E1 reaction cycle produces ROS as a result of uncoupling of the reaction. In addition, the NADP⁺ generated during this process is recycled via NOX to produce superoxides and peroxides. The superoxides can further generate peroxides through Fenton chemistry. The ROS generated via this mechanism increases the caspase-3 activity, which then induces DNA fragmentation, ultimately leading to apoptosis in astrocytes. The combination of MA and gp120 thus exacerbates the toxicity through common molecular mechanism(s). R-H represents MA or physiological substrates, such as dopamine. R-H may be oxidized to R-OH, which may further be oxidized to R-(OH)₂ or R-CHO

further confirmed the toxic consequences of ROS production as a result of MA/gp120 interaction. In addition, inhibitors for CYP2E1, NOX, and FWH reaction abolished caspase-3 activity and apoptotic cell death, which clearly suggested that ROS, induced by gp120/MA interaction, was responsible for the apoptotic cell death.

In summary, results from our present study indicate that in the astrocytes, MA potentiates the gp120-mediated oxidative stress that may worsen the neurodegenerative complications in HIV-infected MA users. The involvement of CYP2E1 in interaction between gp120 and MA presents a novel opportunity to explore their further roles in neurotoxicity. Furthermore, it provides an opportunity to use this pathway as a target to find potentially novel pharmaceuticals. Specific inhibitors such as DAS, which is also a food additive,²² may potentially be used as food supplements/drugs in HIV-infected MA users to reduce neurotoxicity. The oxidative stress-mediated increased toxicity by MA/gp120 in astrocytic cells also provides an opportunity to potentially use antioxidants in

reducing MA-mediated HIV-1 pathogenesis in the patients who abuse MA. Finally, as peptide inhibitors against NOX have been shown to be protective in animal models, these inhibitors may potentially be used to reduce MA-induced HIV-1 pathogenesis in the humans.

Materials and Methods

Cell culture and reagents. All the experiments were performed either in SVGA astrocytes, a clone of SVG astrocytes, or primary HFAs. HFAs were obtained from aborted fetuses and >98% of the cells were found to be positive for GFAP (data not shown). Both the SVGA and the primary HFA cells were maintained at 37 °C in a humidified chamber with 5% CO₂. The cells were cultured in Dulbecco's modified Eagle's medium supplemented with 10% fetal bovine serum, 1% L-glutamine, 1% sodium bicarbonate, 1% non-essential amino acids, and 0.1% gentamicin. All cells were cultured in 12-well plates and allowed to adhere overnight before any treatment was performed.

MA was purchased from Sigma-Aldrich (St. Louis, MO, USA). The plasmid expressing gp120 (pSyngp120 JR-FL) was originally constructed by Drs. Park and Seed (Department of Molecular Biology, and Melvin and Barbara Nessel Gene Therapy Center, Massachusetts General Hospital, Boston, MA, USA). Recombinant protein gp120IIB and pSyngp120 JR-FL plasmids were obtained from the NIH

AIDS Research and Reference Program (Germantown, MD, USA). DAS was obtained from Fisher Scientific (Pittsburgh, PA, USA). DPI, DFO, OP, TROLOX, vit. E, fluoxetine, and paroxetine were purchased from Sigma-Aldrich. The treatments with antagonist were performed 1 h before the stimulus (MA and/or gp120) and maintained throughout the length of exposure. Specific antibodies against ferritin heavy chain (B12), CYP2E1 (H-21), and glyceraldehyde 3-phosphate dehydrogenase (GAPDH) (FL-335) were purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA), antibody against caspase-3, which detects procaspase3 (35 kDa) and cleaved caspase-3 (17 kDa) was purchased from Cell Signaling Technology (Danvers, MA, USA) and antibody against CYP2D6 and CYP2B6 were purchased from Abcam (Cambridge, MA, USA). Specific siRNA for CYP2E1, CYP2B6, CYP2D6, NOX2, NOX4, and control siRNAs were purchased from Dharmacon (Thermo Fisher Scientific Inc., Waltham, MA, USA).

Transfection. SVGA astrocytes were transfected with 2 μ g plasmid encoding gp120 (pSyn-gp120 JR-FL) using Lipofectamine 2000 (Invitrogen Inc., Carlsbad, CA, USA) as discussed previously.^{5,6,14,41} Briefly, 3×10^5 SVGA cells per well were seeded in a 12-well plate and incubated overnight to allow them to adhere. The following day, the complete medium was removed and the cells were washed two times with PBS, followed by incubation with serum-free medium. The transfection reagents were added to the cells that were transfected and incubated for 5 h. After 5 h, the cells were washed two times with PBS and incubated with the complete medium for a total duration of 24 h. Cells transfected with empty vector were used as controls in all the transfection experiments. For the treatments with inhibitor, SVGA astrocytes were pretreated with the antagonist 1 h before the transfection in serum-free medium and the antagonist was maintained throughout the duration of transfection.

In experiments involving siRNA transfections, SVGA cells were seeded at 8×10^5 cells per well in a six-well plate. The following day, cells were washed two times with PBS and transfected with 20 pmol of specific siRNA against NOX4, CYBB (NOX2), CYP2E1, CYP2B6, CYP2D6, or control siRNA for 48 h. The cells were then recounted and 2.75×10^5 cells per well were seeded in a 12-well plate. These gene-depleted cells were then exposed to MA and/or gp120 for 24 h and ROS was measured. The efficiency of gene knockdown was confirmed using western blotting.

Real-time reverse transcriptase-polymerase chain reaction. To measure the mRNA expression levels of various CYPs, the cells were either treated with 500 μ M MA and/or transfected with plasmid encoding gp120. Upon termination of the treatment, total RNA was isolated using Qiagen RNeasy Mini Kit (Qiagen, Valencia, CA, USA). The RNA (120 ng) was reverse transcribed into cDNA at 37 °C for 60 min using a 2-step TaqMan Gene Expression Kit (Applied Biosystems, Foster City, CA, USA). The cDNA obtained was amplified for GAPDH (4333764F), CYP1A1 (Hs01054794_m1), CYP2B6 (Hs03044636_m1), CYP2E1 (Hs00559367_m1), CYP3A4 (Hs00430021_m1), CYP2D6 (Hs00164385_m1), and CYP2A6 (Hs0071162_m1) using the probe mix obtained from Applied Biosystems as per the manufacturer's protocol. The expression values for various CYPs were normalized using GAPDH as a housekeeping gene. Relative fold expressions for various genes were analyzed using the $2^{-\Delta\Delta Ct}$ method.

Measurement of ROS production. To measure the oxidative stress produced in the astrocytes, the cells were treated with various agents for appropriate duration. After the termination of treatments, the cells were washed two times with PBS and incubated with 5 μ M 5-(and-6)-chloromethyl-2',7'-dichlorodihydrofluorescein diacetate (CM-H₂DCFDA) (Molecular Probes, Carlsbad, CA, USA) for 30 min in serum-free medium at 37 °C. The cells were then washed two times with PBS to remove unloaded dye and the images were taken using Leica fluorescent microscope (DMI 3000B; Leica Microsystems Inc., Buffalo Grove, IL, USA). Unstained cells were used as experimental controls and the fluorescent images were obtained using excitation and emission wavelengths at 485 and 535 nm, respectively.

To obtain the fluorescence intensity corresponding to ROS, the cells were collected and acquired using FITC wavelengths on FACSCanto II flow cytometer (BD Biosciences, San Jose, CA, USA). The ROS production was measured using unstained cells as negative controls and cells treated with 500 μ M H₂O₂ as positive controls. The mean fluorescence intensities were compared between different treatments.

Western blotting. The protein levels were measured by analyses of western blots using whole-cell lysates obtained from astrocytes treated with appropriate reagents. Briefly, the cells were lysed using radioimmunoprecipitation assay

(RIPA) buffer (Boston BioProducts, Ashland, MA, USA) supplemented with Halt™ Protease Inhibitor Cocktail (Pierce Biotechnology, Rockford, IL, USA). The protein concentrations in the lysates were measured using BCA Protein Assay Kit (Pierce Biotechnology). The protein samples (30 μ g) were electrophoresed on a 10% polyacrylamide gel and transferred to PVDF membrane. Finally, the membrane was probed with appropriate primary and secondary antibodies for caspase-3, ferritin heavy chain (Ferritin HC), NOX2, NOX4, CYP2B6, CYP2E1, and CYP2D6 to measure their expression levels. The bands were detected using BM Chemiluminescence Western Blotting Substrate (POD) (Roche Applied Sciences, Indianapolis, IN, USA). The bands were analyzed using FlourChem HD2 software (Alpha Innotech, San Leandro, CA, USA) and the intensities were normalized using GAPDH as loading control.

Cell survival assay (MTT assay). The cell viability was measured using a colorimetric 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. Briefly, 2×10^5 astrocyte cells were seeded in 12-well plates and treated with either the inhibitor or saline with subsequent treatment with MA and/or gp120 for 48 h. Upon termination of the treatments, the medium was replaced with 0.2 mM MTT containing medium and incubated at 37 °C for 4 h. The medium was carefully removed and purple formazan crystals were dissolved in 500 μ l DMSO containing 125 μ l of Sorenson's glycine buffer. The color formation was measured using Benchmark Microplate Reader (Bio-Rad Laboratories, Hercules, CA, USA) with absorbance at 570 nm and reference at 650 nm.

TUNEL assay. The measurement of cell viability was also correlated with DNA damage using TUNEL assay as per the manufacturer's protocol (GenScript, Piscataway, NJ, USA). Briefly, 2×10^5 cells per well were seeded in 12-well plates and the cells were exposed to the treatments with appropriate compounds. Upon treatments, the cells were air-dried for 5 min and fixed (4% paraformaldehyde in PBS) for 30 min at room temperature. After fixation, they were incubated overnight at 4 °C and permeabilized using 1% Triton X-100. The cells were then blocked in 3% H₂O₂ in methanol and labeled with TUNEL reaction mixture (biotin-11-dUTP and TdT) and streptavidin-FITC. The smear of cells were washed with PBS to remove excess of labeling reagents and incubated with POD-conjugated anti-FITC substrate. Finally, 3,3'-diaminobenzidine (DAB) working reagent was added and the cells were observed under the light microscope. The images were taken using Leica DMI 300B (Leica Microsystems Inc., Buffalo Grove, IL, USA).

Caspase-3 activity assay. The caspase-3 activity was measured using caspase-3 colorimetric assay kit (BioVision Inc., Milpitas, CA, USA) as per the manufacturer's protocol. Briefly, apoptosis was induced as per the treatments under investigation and the cells were lysed using lysis buffer. The protein concentration in the lysates was measured using BCA Protein Assay Kit (Pierce Biotechnology, Rockford, IL, USA) and 50 μ l of lysates containing 200 μ g protein was used for caspase-3 activity. The lysates were mixed with $2 \times$ reaction buffer containing 10 μ M DTT and DEVD-pNA substrate and incubated at 37 °C for 2 h. Finally, the absorbance was measured using Benchmark Microplate Reader (Bio-Rad Laboratories) at 405 nm and the caspase activity was calculated with relative standard curve obtained using substrate alone.

Protein carbonylation assay. The measurement of carbonylated protein was performed using protein carbonyl colorimetric assay kit (Cayman Chemicals, San Francisco, CA, USA) as per the manufacturer's protocol. Briefly, the cells were treated and lysed with RIPA lysis buffer. The protein samples were checked for the absence of nucleic acid contamination and 200 μ g proteins were used for the assay. The lysates were mixed with DNPH (2,4-dinitrophenylhydrazine) and incubated in the dark for 1 h followed by consecutive treatments with 20% and 10% trichloroacetic acid (TCA). This was followed by three washes with ethanol/ethyl acetate (1:1 mixture) and finally the pellet was resuspended in guanidine hydrochloride. The absorbance of the supernatants was measured at 370 nm using a microplate reader and carbonylated protein content was calculated.

Statistical analysis. The statistical analysis was performed to represent the data in mean \pm S.E. values. Results were based on at least three separate experiments unless specified with each experiment performed in triplicates. For the comparison between mock/control group, treatments were performed using two-tailed Student's *t*-test to calculate *P*-values, and *P*-value ≤ 0.05 was considered statistically significant. The *P*-value ≤ 0.05 was indicated by * and ≤ 0.01 was indicated by ** in the bar graphs. Experiments involving two variables

were analyzed using two-way ANOVA to address whether they interact with each other in a synergistic manner. To examine dose-dependent effect of the inhibitors, results were analyzed using *P*-value for trend test using general linear model. No adjustments were made in the statistical analysis for multiple comparisons.

Conflict of Interest

The authors declare no conflict of interest.

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