

Effect of quercetin on traits of the metabolic syndrome, endothelial function and inflammation in men with different *APOE* isoforms

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KEYWORDS

Metabolic syndrome; Quercetin; Endothelial function; Cardiovascular disease **Abstract** Background and aims: The polyphenol quercetin may prevent cardiovascular diseases due to its vasorelaxant and anti-oxidative properties. We investigated the effects of quercetin on risk factors of atherosclerosis, biomarkers of inflammation and oxidative stress, depending on the apolipoprotein E (APOE) genotype.

Methods and results: In a double-blind crossover study 49 healthy male subjects with APOE genotype 3/3 (n = 19), 3/4 (n = 22) and 4/4 (n = 8) consumed 150 mg/d quercetin or placebo for 8 weeks each, intermitted by a three-week washout phase. After each intervention, endothelial function, anthropometry, metabolic and inflammatory parameters were measured in the fasting and postprandial state following a standardized lipid-rich meal.

Endothelial function was not changed. In all subjects combined, quercetin significantly decreased waist circumference (P = 0.004) and postprandial systolic blood pressure (P = 0.044). Postprandial triacylglycerol concentrations were significantly decreased and HDL-cholesterol concentrations increased after quercetin as compared to placebo consumption (P = 0.025). Quercetin also moderately increased levels of TNF α (P = 0.024). There

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was a significant gene—diet interaction for waist circumference and for body mass index (BMI). *Conclusions:* Quercetin supplementation improved some risk factors of cardiovascular disease, yet exerted slightly pro-inflammatory effects. Genotype-dependent effects were seen only on waist circumference and BMI.

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Introduction

There is some evidence that the ingestion of flavonoids may be inversely correlated with the incidence of cardiovascular diseases and related risk factors [1]. Quercetin a major dietary flavonoid - occurs naturally in many plant foods, mainly in onions, broccoli, green cabbage and apples, and in lower concentrations in black tea and red wine. Quercetin may act through various anti-inflammatory mechanisms, e.g. regulation of the expression of cellular adhesion molecules and of the secretion of proinflammatory cytokines and chemokines [2,3]. Quercetin reduced also blood pressure both in human and animal studies [4-6] and inhibited the platelet activation pathway in man [7]. It improved endothelial function in rat models of hypertension [8]. Both short- and long-term black tea consumption improved endothelial function in patients with existing coronary artery disease [9].

The multifunctional and polymorphic protein apolipoprotein E (APOE) plays a key role in the metabolism of plasma lipids and affects the development of atherosclerosis. There are three isoforms designated E2, E3, and E4 in which carriers of the APOE4 allele are at a higher coronary risk [10]. APOE4 has arginine instead of cysteine at position 112 and thus one SH-group less than APOE3. This may contribute to impaired antioxidant activity. The fact that the anti-oxidative capacity of APOE4 is lower compared to the other isoforms could render them more susceptible to cardiovascular disease [11]. Furthermore APOE4 is associated with larger VLDL and potentially atherogenic remnant size triacylglycerol-rich lipoproteins (TRL), whereas APOE3 preferentially binds to HDL particles [12] which have antiinflammatory and antiatherogenic properties. Dysfunction of either the coronary or peripheral vascular endothelium constitutes an early and independent predictor of cardiovascular events [1,13]. The APOE4 allele was associated with impaired endothelium-dependent arterial dilation in the early stage of type 2 diabetes mellitus [14].

Both normolipidemic and hyperlipidemic subjects with the APOE4 genotype showed a stronger postprandial triacylglycerol response than respective subjects with APOE3/ 3 [15]. The protective effect of dietary compounds may be better visible in the postprandial state as well [9]. Furthermore, potentially atherogenic and proinflammatory remnant lipoproteins that may contribute to endothelial dysfunction are generated in the postprandial state [16]. Anti-oxidants such as ascorbic acid can enhance endothelial function [17]. Since quercetin consumption was expected to exert anti-oxidative effects the impact of an 8week quercetin intervention on endothelial function was investigated. In addition, metabolic, oxidative and inflammatory parameters were assessed. Carriers of the APOE4 allele were compared to APOE3 homozygotes from a prospectively genotyped study group in a double-blind randomized controlled clinical trial, both in the fasting and in part in the postprandial state, following a lipid-rich meal.

Methods

Study population

Forty-nine subjects (age 48-68 years) were recruited from the population-based cohort Metabolic Intervention Cohort Kiel (MICK, n = 1508) characterized previously [18]. Major exclusion criteria were known disorders that affect the digestion and metabolism of food components, and established diabetes mellitus (fasting glucose levels >6.9 mmol/ L after repeated determination). Further exclusion criteria were intake of lipid lowering and anti-hypertensive drugs, hormones, and other drugs with an impact on gastrointestinal motility, absorption or metabolism of nutrients. Eight APOE4/4 homozygotes were matched with 20 APOE3/3 homozygotes and 22 APOE3/4 heterozygotes. Right at onset two subjects with APOE3/3 and one with APOE3/4 dropped out of the study, because of a herniated vertebral disc, inability to swallow the capsules and night sweat. Only the latter condition may be related to treatment. They were replaced by other APOE3/3 and 3/4 genotype carriers. The study was approved by the local Ethics Advisory Committee and carried out according to the Helsinki declaration. Participants gave written informed consent prior to the study.

Study design

The study was single-centre, double-blind, randomized, crossover, placebo-controlled, and performed between May 2007 and May 2008. Two 8-week treatment periods were separated by a 3-week washout period. Subjects were randomly assigned to receive a total of either 150 mg quercetin dihydrate (Voigt Global Distribution Inc., Lawrence, KS, USA) or placebo respectively, provided in six capsules per day, which were consumed with the three principal meals, two capsules per meal. Verum and placebo capsules were identical in shape and taste. Compliance was checked by counting returned capsules. It was considered sufficient if >85% of the capsules were consumed. All participants achieved this goal. Participants were instructed not to change their eating habits and physical activity routine, and not to use any dietary supplements of vitamins, minerals, or oil preparations.

Anthropometric parameters and blood pressure were recorded following standard operation procedures according to WHO and Deutsche Hochdruckliga, respectively and blood samples were collected at baseline (day zero) and after every 8-week intervention. Fasting blood samples were collected between 0700 and 0900. A potato soup (as previously described [19]) containing 60 g sunflower oleic oil (Henry Lamotte Oils GmbH, Bremen, Germany), was then consumed within 15 min. After the meal blood samples were collected repeatedly, after 30 and 60 min and then hourly over a period of 8 h for the measurement of post-prandial triacylglycerol concentration, and over a period of 5 h for glucose and insulin concentrations. Plasma or serum aliquots were stored at -20 °C or -80 °C until analysis, depending on assay requirements.

Endothelial function

Vascular endothelial function was assessed at a separate visit two days after anthropometry and blood sampling, at the start of the study and after every 8-week intervention, by reactive hyperemia with finger plethysmographic methodology (Reactive Hyperemia peripheral arterial tonometry, PAT) using the Endo-PAT2000 system (Itamar, Caesarea, Israel), as described previously [20]. PAT index was determined in the fasting state and 4 h after consuming the meal described above. PAT index was calculated as the ratio of the average amplitude of the PAT signal post occlusion divided by the average baseline amplitude, and corrected to the non-occluded control arm. Low PAT readings predict adverse cardiovascular events [20] and show a significant association with adiposity and other cardiovascular risk factors [21]. PAT measurement has been approved by the FDA.

Biochemical analyses

Routine biochemical parameters were analyzed in an accredited laboratory (University Hospital Kiel). Glucose, total (TC), high density lipoprotein (HDL-C) and lowdensity lipoprotein (LDL-C) cholesterol, and triacylglycerols were analyzed in plasma by enzymatic methods (Thermo Fisher Scientific, Passau, Germany) using a Konelab 20i clinical analyzer (Thermo Fisher Scientific), and insulin by RIA (Adaltis, Bologna, Italy). HOMA-IR (homeostasis model assessment-insulin resistance) was calculated as [(fasting insulin $mU/L \times fasting$ glucose mmol/L)/22.5]. For the postprandial course of triacylglycerol, glucose and insulin concentrations the area under curve (AUC) was calculated according to the trapezoidal rule. Soluble adhesion molecules s-VCAM, s-ICAM, and s-E-selectin (Bender, Wien, Austria), highsensitive C-reactive protein (hs-CRP, DRG, Marburg, Germany), oxidized LDL (oxLDL, Mercodia, Uppsala, Sweden) and tumor necrosis factor alpha (TNF α , R&D, Wiesbaden-Nordenstadt, Germany) were determined by ELISA, the 8-epimer of prostaglandin F2 α (8-iso-PGF2 α) by an enzyme immunoassay (Assay Designs, Ann Arbor, USA). Total glutathione of washed packed erythrocytes, i.e. the sum of reduced and oxidized glutathione (GSH), was measured by enzymatic colorimetric procedure (Sigma--Aldrich, Steinheim, Germany). Plasma guercetin concentration was analyzed by HPLC/MS (HPLC: Agilent Technologies, Waldbronn, Germany; MS: 3200 QTrap; AB Sciex, Darmstadt, Germany) after treatment with a betaglucuronidase/sulfatase mixture (from Helix pomatia; Sigma—Aldrich) and extraction into ethylacetate as described previously [22].

Genotyping

Genomic DNA was purified using the NucleoSpin Blood Isolation kit (Macherey-Nagel, Düren, Germany). Genotyping of the variations in the *APOE* gene (rs429358, rs7412) were performed with the TaqMan system (ABI, Foster City, CA, USA), fluorescence was measured with ABI Prism 7900 HT sequence detection system.

Statistical analysis

The primary parameter defined prior to the study was the intervention effect on fasting endothelial function, i.e. the treatment-dependent change from baseline to 8weeks. Spearman's correlation coefficient was chosen for correlation analyses. The Mann-Whitney U-test was used to test for associations between genotypic groups and baseline values. The analysis of variance for repeated measures (RM-ANOVA) was chosen to test for effects of the APOE polymorphism, the dietary intervention and the interaction between diet and genotype on metabolic parameters. The probability plot of the standardized residuals is close to normal. APOE4 allele carriers (APOE3/ 4 and 4/4) were combined for statistical analysis, since it is the dominant allele and there were no differences in response between APOE3/4 and 4/4 subjects. Statistical analysis was performed with SPSS (SPSS for windows, Release 18, LEAD Technologies Inc.). Data are given as means \pm SEM.

Results

Characteristics of study subjects at baseline

Baseline body weight, body mass index (BMI) and waist circumference did not differ between *APOE* genotype groups (Table 1). The different genotypes showed similar concentrations for HDL-C, LDL-C, fasting triacylglycerols and glucose. In *APOE4* carriers, TC concentration tended to be higher than in *APOE3/3* homozygotes (P = 0.051). Fasting PAT indices were significantly higher in the *APOE4* group (P < 0.01), suggesting better endothelial function, but postprandial endothelial function did not differ between *APOE* isoforms. Postprandial PAT correlated with fasting PAT indices (r = 0.366; P < 0.01), but neither fasting nor postprandial PAT correlated with the respective systolic (SBP) or diastolic blood pressure (DBP). CRP concentration was significantly lower in the *APOE4* compared to the *E3/3* group (P < 0.05).

Anthropometry

Quercetin treatment as compared to placebo decreased waist circumference by 0.63 cm (P < 0.01) in all subjects. A significant diet-by-genotype interaction effect was

	All subjects	APOE3/3	APOE4	Р
	(n = 49)	(<i>n</i> = 19)	(n = 30)	
Anthropometrics				
Age (years)	$\textbf{59.4} \pm \textbf{0.9}$	59.5 \pm 1.4	$\textbf{59.4} \pm \textbf{1.2}$	0.967
Body weight (kg)	84.9 ± 1.1	$\textbf{83.7}\pm\textbf{1.7}$	$\textbf{85.7} \pm \textbf{1.4}$	0.601
BMI (kg/m ²)	$\textbf{26.3} \pm \textbf{0.3}$	$\textbf{26.0} \pm \textbf{0.5}$	$\textbf{26.4} \pm \textbf{0.4}$	0.594
Waist circumference (cm)	$\textbf{97.7}\pm\textbf{0.9}$	98.1 ± 1.3	$97.5\pm c1.3$	0.992
Fasting endothelial function				
Endo-PAT	$\textbf{2.12} \pm \textbf{0.08}$	$\textbf{1.89} \pm \textbf{0.13}$	$\textbf{2.27} \pm \textbf{0.1}$	0.003
SBP (mmHg)	$\textbf{138.4} \pm \textbf{2.3}$	137.1 \pm 2.3	139.3 ± 3.34	0.711
DBP (mmHg)	$\textbf{84.4} \pm \textbf{1.3}$	$\textbf{85.4} \pm \textbf{1.4}$	$\textbf{83.73} \pm \textbf{1.83}$	0.541
Postprandial (4 h) endothelial fu	Inction			
Endo-PAT	$\textbf{2.25} \pm \textbf{0.09}$	$\textbf{2.22} \pm \textbf{0.14}$	$\textbf{2.28} \pm \textbf{0.12}$	0.798
SBP (mmHg)	$\textbf{132.9} \pm \textbf{2.2}$	130.2 ± 2.7	134.7 ± 3.07	0.535
DBP (mmHg)	$\textbf{80.8} \pm \textbf{1.3}$	$\textbf{80.9} \pm \textbf{1.6}$	$\textbf{80.8} \pm \textbf{1.86}$	0.828
Fasting metabolic parameters				
Glucose (mmol/L)	$\textbf{5.60} \pm \textbf{0.08}$	$\textbf{5.55} \pm \textbf{0.12}$	$\textbf{5.61} \pm \textbf{0.10}$	0.918
Triacylglycerols (mmol/L)	$\textbf{1.20} \pm \textbf{0.07}$	1.08 ± 0.11	$\textbf{1.27} \pm \textbf{0.10}$	0.182
Cholesterol (mmol/L)			$\textbf{5.66} \pm \textbf{0.19}$	0.051
HDL-cholesterol (mmol/L)			$\textbf{1.41} \pm \textbf{0.07}$	0.277
LDL-cholesterol (mmol/L)			$\textbf{3.64} \pm \textbf{0.16}$	0.182
CRP (mg/L)	$\textbf{4.28} \pm \textbf{0.16}$	$\textbf{4.40} \pm \textbf{0.38}$	$\textbf{4.21} \pm \textbf{0.1}$	0.044

Bold indicates P-values less than 0.05.

PAT = peripheral arterial tonometry; SBP = systolic blood pressure; DBP = diastolic blood pressure; CRP = C-reactive protein.

Data are expressed as mean \pm SEM; p for Mann–Whitney U-test.

observed on BMI (P < 0.05), body weight (P < 0.05) and waist circumference (P < 0.01). Thus, the decrease in waist circumference was greater in APOE3 homozygotes than in APOE4 carriers. Ouercetin decreased BMI as well as body weight in APOE3/3 while it increased BMI slightly in APOE4 carriers. No other significant diet-by-genotype interactions were found.

Endothelial function

Quercetin did not change endothelial function, neither fasting nor 4 h after a fat-rich meal. It decreased postprandial SBP by 5.73 mmHg (P < 0.05), but did not change fasting SBP or fasting and postprandial DBP.

Lipid and carbohydrate metabolism

Quercetin increased HDL-C concentrations by 0.06 mmol/L (P < 0.05) and lowered the triacylglycerols significantly at all time points during the first 2 h following a lipid-rich meal. Accordingly, triacylglycerol AUC during the first 4 h were 11% lower following guercetin treatment as compared to placebo (P < 0.05), but AUC over 8 h was not (P = 0.085) (Fig. 1, Table 2). The postprandial glucose increase was also slightly but non-significantly attenuated following guercetin treatment (P = 0.066). None of these changes differed by genotype. Other lipid parameters, fasting glucose and insulin concentrations and postprandial insulin remained largely unchanged. Independently of the intervention, the APOE4 genotype was associated with significantly higher TC (P < 0.05) and non-significantly higher LDL-C concentrations (P = 0.075).

Lipid peroxidation and inflammation

Quercetin increased TNF α concentration by 0.11 pg/ml (P < 0.05) and decreased total GSH concentration in erythrocytes non-significantly (P = 0.067). Other inflammatory parameters (s-E-Selectin, s-VCAM, s-ICAM, oxLDL, hs-CRP) and the urinary isoprostane 8-iso-PGF2_{α} were not changed.

Plasma quercetin

Quercetin concentration in plasma was increased from 121.9 \pm 7.5 to 193.8 \pm 20.4 nmol/L following quercetin as compared to placebo consumption (n = 49, P < 0.01). There was no genotype-dependent difference of responsiveness (data not shown), as observed in another recent quercetin supplementation trial [6].

Discussion

Recently, the cardiovascular effects of quercetin were investigated in a group of men and women with a high cardio-metabolic risk phenotype where the APOE polymorphism was retrospectively genotyped [6]. In this study we prospectively genotyped members of the MICK cohort and matched different APOE genotypes for similar body shape, weight and age. Our male study subjects were of normal weight and without signs of the metabolic syndrome,

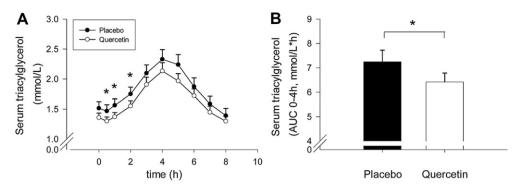


Figure 1 (A) Fasting and postprandial triacylglycerol concentrations following 8-week placebo or quercetin supplementation. (B) Area under the curve of the triacylglycerol response during the first 4 h postprandially. Means \pm SEM; *p < 0.05.

Table 2	Fasting and postprandial	parameters after	8-week placebo	and quercetin	supplementation	according to the	APOE
polymorpl	hism.						

	APOE3/3 ($n = 19$)		APOE4 ($n = 30$)		p	р	р
	Placebo	Quercetin	Placebo	Quercetin	intervention	genotype	interaction
BMI (kg/m ²)	$\textbf{26.3} \pm \textbf{0.5}$	$\textbf{26.1} \pm \textbf{0.5}$	$\textbf{26.4} \pm \textbf{0.4}$	$\textbf{26.5} \pm \textbf{0.3}$	0.320	0.638	0.028
Body weight (kg)	$\textbf{84.7} \pm \textbf{1.8}$	$\textbf{84.2} \pm \textbf{1.7}$	$\textbf{85.8} \pm \textbf{1.5}$	$\textbf{86.0} \pm \textbf{1.4}$	0.319	0.520	0.025
Waist circumference (cm)	100.1 ± 1.6	$\textbf{98.4} \pm \textbf{1.4}$	99.1 ± 1.5	99.1 ± 1.2	0.004	0.927	0.003
Fasting parameters							
Endo-PAT	$\textbf{1.95} \pm \textbf{0.19}$	$\textbf{1.96} \pm \textbf{0.19}$	$\textbf{2.06} \pm \textbf{0.09}$	$\textbf{2.00} \pm \textbf{0.07}$	0.821	0.513	0.673
SBP (mmHg)	$\textbf{135.8} \pm \textbf{3.6}$	$\textbf{133.4} \pm \textbf{3.9}$	$\textbf{133.7} \pm \textbf{3.2}$	$\textbf{132.2}\pm\textbf{3.3}$	0.094	0.835	0.588
DPB (mmHg)	$\textbf{82.7} \pm \textbf{1.3}$	$\textbf{81.7} \pm \textbf{2.1}$	$\textbf{79.1} \pm \textbf{1.5}$	$\textbf{80.1} \pm \textbf{1.7}$	0.358	0.161	0.259
Glucose (mmol/L)	$\textbf{5.74} \pm \textbf{0.13}$	$\textbf{5.64} \pm \textbf{0.12}$	$\textbf{5.64} \pm \textbf{0.13}$	$\textbf{5.66} \pm \textbf{0.10}$	0.535	0.830	0.307
Insulin (pmol/L)	$\textbf{109.0} \pm \textbf{12.5}$	$\textbf{103.5} \pm \textbf{9.8}$	$\textbf{93.9} \pm \textbf{6.3}$	$\textbf{89.2} \pm \textbf{5.6}$	0.228	0.192	0.928
HOMA-IR	$\textbf{3.94} \pm \textbf{0.53}$	$\textbf{3.66} \pm \textbf{0.42}$	$\textbf{3.33} \pm \textbf{0.27}$	$\textbf{3.18} \pm \textbf{0.25}$	0.228	0.261	0.716
Triacylglycerols (mmol/L)	$\textbf{1.45} \pm \textbf{0.11}$	$\textbf{1.42} \pm \textbf{0.12}$	$\textbf{1.56} \pm \textbf{0.16}$	$\textbf{1.32} \pm \textbf{0.10}$	0.138	0.996	0.242
Total cholesterol (mmol/L)	$\textbf{5.22} \pm \textbf{0.25}$	$\textbf{5.34} \pm \textbf{0.19}$	$\textbf{5.78} \pm \textbf{0.20}$	$\textbf{5.78} \pm \textbf{0.17}$	0.205	0.040	0.231
HDL-cholesterol (mmol/L)	$\textbf{1.23} \pm \textbf{0.07}$	$\textbf{1.31} \pm \textbf{0.09}$	$\textbf{1.34} \pm \textbf{0.06}$	$\textbf{1.39} \pm \textbf{0.06}$	0.025	0.346	0.450
LDL-cholesterol (mmol/L)	$\textbf{3.25} \pm \textbf{0.19}$	$\textbf{3.33} \pm \textbf{0.15}$	$\textbf{3.68} \pm \textbf{0.15}$	$\textbf{3.72} \pm \textbf{0.14}$	0.400	0.075	0.791
s-E-Selectin (ng/mL)	$\textbf{48.2} \pm \textbf{6.1}$	$\textbf{49.8} \pm \textbf{6.4}$	$\textbf{43.3} \pm \textbf{2.9}$	$\textbf{44.1} \pm \textbf{2.4}$	0.188	0.390	0.658
s-VCAM (ng/mL)	$\textbf{829.5} \pm \textbf{41.3}$	$\textbf{837.9} \pm \textbf{43.8}$	$\textbf{863.2} \pm \textbf{38.8}$	$\textbf{848.5} \pm \textbf{38.1}$	0.827	0.708	0.419
s-ICAM (ng/mL)	$\textbf{239.8} \pm \textbf{13.4}$	$\textbf{242.1} \pm \textbf{12.9}$	$\textbf{247.3} \pm \textbf{14.5}$	$\textbf{254.5} \pm \textbf{13.3}$	0.324	0.623	0.615
oxLDL (U/L)	$\textbf{81.1} \pm \textbf{5.1}$	$\textbf{80.3} \pm \textbf{4.1}$	$\textbf{84.2}\pm\textbf{3.9}$	$\textbf{87.0} \pm \textbf{3.8}$	0.681	0.737	0.481
GSH (nmol/mL erythrocytes)	1499 ± 71	1415 ± 66	1543 ± 68	1445 ± 50	0.067	0.652	0.883
CRP (mg/L)	$\textbf{3.30} \pm \textbf{0.97}$	$\textbf{4.07} \pm \textbf{1.41}$	$\textbf{2.18} \pm \textbf{0.42}$	$\textbf{3.31} \pm \textbf{0.85}$	0.174	0.402	0.793
$TNF\alpha$ (pg/mL)	$\textbf{1.76} \pm \textbf{0.15}$	$\textbf{1.98} \pm \textbf{0.17}$	$\textbf{1.99} \pm \textbf{0.20}$	$\textbf{2.03} \pm \textbf{0.24}$	0.024	0.329	0.505
8-iso-PGF2a (pg/mg creatinine)	$\textbf{191.5} \pm \textbf{164.3}$	$\textbf{143.6} \pm \textbf{81.2}$	155.6 ± 105.5	$\textbf{155.7} \pm \textbf{147.8}$	0.400	0.655	0.398
Postprandial parameters							
Endo-PAT (4 h pp)	2.10 ± 0.22	1.98 ± 0.18	$\textbf{2.32} \pm \textbf{0.10}$	2.27 ± 0.10	0.880	0.238	0.686
SBP (4 h pp, mmHg)	130.3 ± 2.4	$\textbf{125.7} \pm \textbf{3.2}$	134.5 ± 3.8	$\textbf{128.0} \pm \textbf{2.4}$	0.044	0.400	0.725
DBP (4 h pp, mmHg)	80.1 ± 1.6	79.8 ± 1.3	79.6 ± 1.9	79.9 ± 1.5	0.989	0.948	0.830
Glucose (AUC, mmol/L*h)	27.7 ± 0.5	26.9 ± 0.5	26.8 ± 0.5	26.7 ± 0.4	0.066	0.474	0.111
Insulin (AUC, mmol/L*h)	109 ± 12.5	103.5 ± 9.8	93.9 ± 6.3	89.2 ± 5.7	0.141	0.328	0.908
Triacylglycerols	14.8 ± 1.2	13.5 ± 1.2	14.9 ± 1.5	13.2 ± 1.0	0.085	0.967	0.804
(AUC, mmol/L*h)							

Bold indicates *P*-values less than 0.05.

PAT = peripheral arterial tonometry; SBP = systolic blood pressure; DBP = diastolic blood pressure; HOMA-IR = homeostasis model assessment of insulin resistance; CRP = C-reactive protein.

Data are expressed as mean \pm SEM; *p* for RM-ANOVA.

except for slightly increased SBP. Endothelial function appeared normal [21]. At entry *APOE4* subjects had just marginally higher TC concentration (which was confirmed during the intervention period) and only trends toward higher LDL-C and triacylglycerol concentrations, opposite to what might have been expected [6,10]. CRP concentrations were lower in *APOE4* than *APOE3/3* subjects, as observed before in a large population-based study. Those authors concluded that the serum CRP is independently determined by the common genetic polymorphisms within the APOE gene [23]. The nature of this unexpected association between *APOE* genotype and CRP is not clear.

Quercetin treatment tended to decrease fasting SBP and significantly decreased postprandial SBP by 5.7 mmHg, confirming previous observations in humans [4,6]. While Egert et al. [6] observed an improvement only in APOE3 homozygous subjects, both APOE3/3 and APOE4 subjects benefited in our study (Table 2). Despite favorable changes in postprandial SBP, endothelial function was not significantly improved, neither the fasting nor the postprandial values. This may be due to the fact that study participants showed fairly normal PAT indices to start with [21]. Fitting with this outcome endothelium-derived adhesion molecules E-selectin, ICAM and VCAM as well as oxLDL were not affected by guercetin administration. Contrary to expectations baseline endothelial function was better in APOE4 as compared to APOE3/3 genotype, but following placebo treatment the difference was no longer significant. In animal models quercetin improved endothelial function under hypertensive conditions only [8]. Furthermore quercetin decreased SBP, and improved dyslipidemia and insulin sensitivity only in obese Zucker rats, but not in lean rats [5]. Again baseline PAT indices did not correlate with blood pressure, emphasizing that factors regulating endothelial function differ at least in part from those regulating blood pressure.

In this study, quercetin did not change fasting, but decreased the postprandial triacylglycerol concentrations. This may result from a guercetin-induced reduction of fatty acid and triacylglycerol synthesis in the liver [24]. In some previous human studies guercetin or a guercetin-rich grape concentrate decreased triacylglycerol concentrations [25,26], but not in other studies [4,27,28]. In this study quercetin increased also HDL-C, as observed before for a grape concentrate [28]. Triacylglycerol concentrations are usually inversely related to HDL-C. TC and LDL-C were not decreased, consistent with two [4,27] but contrary to other previous reports [26,28]. None of these effects were APOE genotype-dependent, while Egert et al. [6] found adverse effects of quercetin on HDL-C and the LDL-C/HDL-C ratio only in APOE4 subjects, and adverse effects on apolipoprotein A-I only in APOE3 subjects. Their study subjects were obese, of a wide age range and both sexes [6] which might have resulted in gender or age bias.

The only adverse effect of quercetin in our study was an increased TNF α concentration, again with no difference by *APOE* genotype. But concentrations were very low under placebo and quercetin treatment alike. All other inflammatory parameters determined were, however, not significantly changed. In other human intervention studies of shorter duration quercetin improved only some of the inflammatory markers [6] or none at all [29]. Nevertheless

quercetin supplementation tended to decrease total glutathione, fitting with a previous observation in vitro [30]. Quercetin did not affect the concentration of 8-iso-PGF2_{α}, an established parameter of lipid peroxidation, again with no difference by *APOE* genotype. This indicates that *APOE3* is not superior to *APOE4* in its anti-oxidative capacity.

Quercetin moderately but significantly reduced BMI, body weight and waist circumference in *APOE3/3* but not in *APOE4* subjects. No such effects have been reported before in humans. But in lean and obese rats quercetin attenuated weight gain, which was attributed to its antiinflammatory effects on adipose tissue [5]. Other mechanisms may also be operative, like anti-adipogenic effects through the modulation of the AMPK, ERK and JNK signaling pathways [31].

The impact of quercetin effects on lipid concentrations in humans seem to be variable and it is not clear to what extent genetic background, diet or lifestyle habits may contribute. Our study in healthy, normal-weight men did not confirm the hypothesis of a beneficial impact of quercetin on endothelial function. Our data, however, provide evidence of moderate beneficial effects on blood pressure and lipidemia, and show for the first time that the postprandial state may emphasize quercetin effects that are less clear under fasting conditions. There were genotype-dependent effects on BMI and waist circumference which could not be explained by anti-inflammatory actions of quercetin. Quercetin exerted even slightly proinflammatory effects, independent of genotype.

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