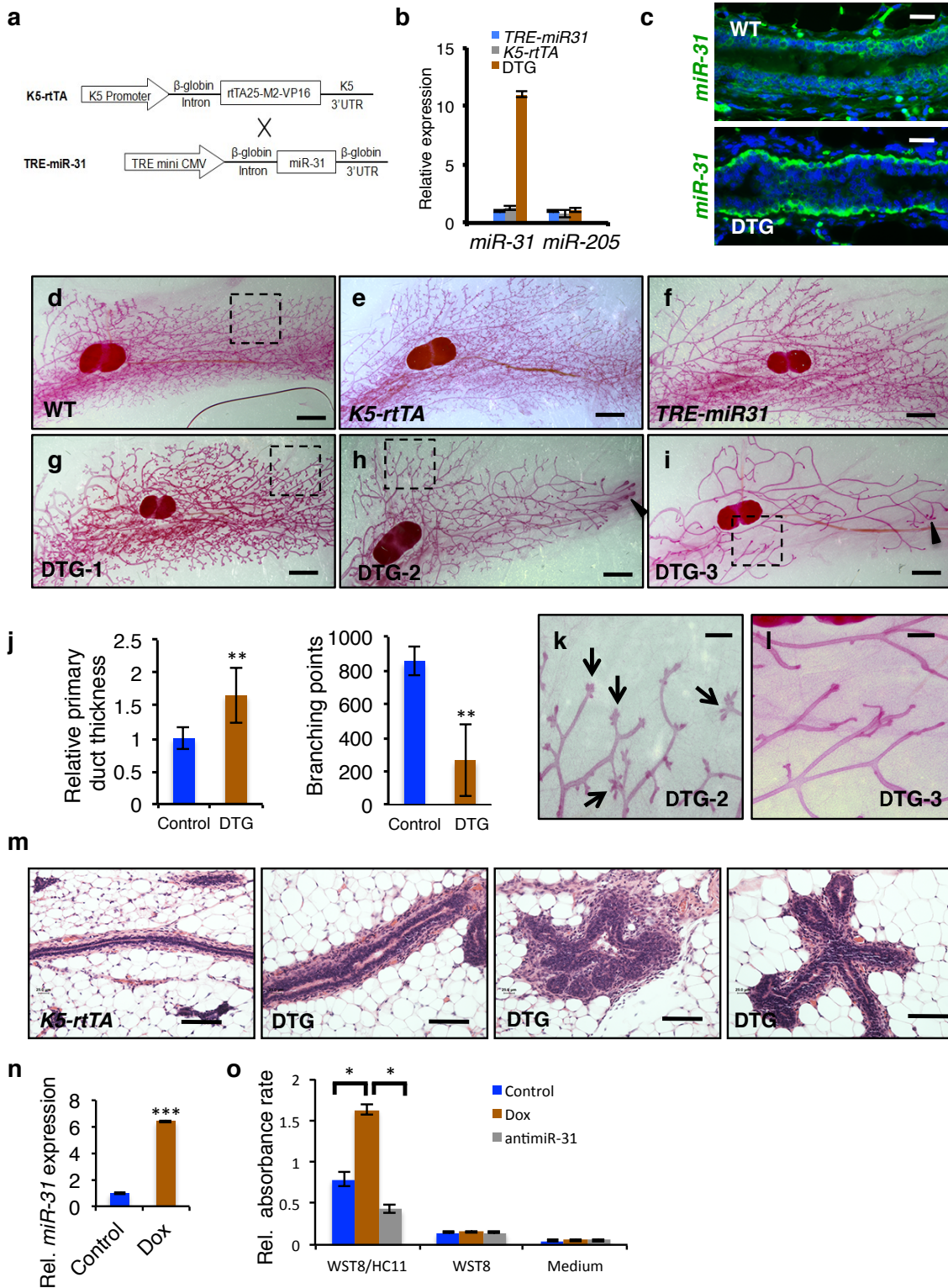


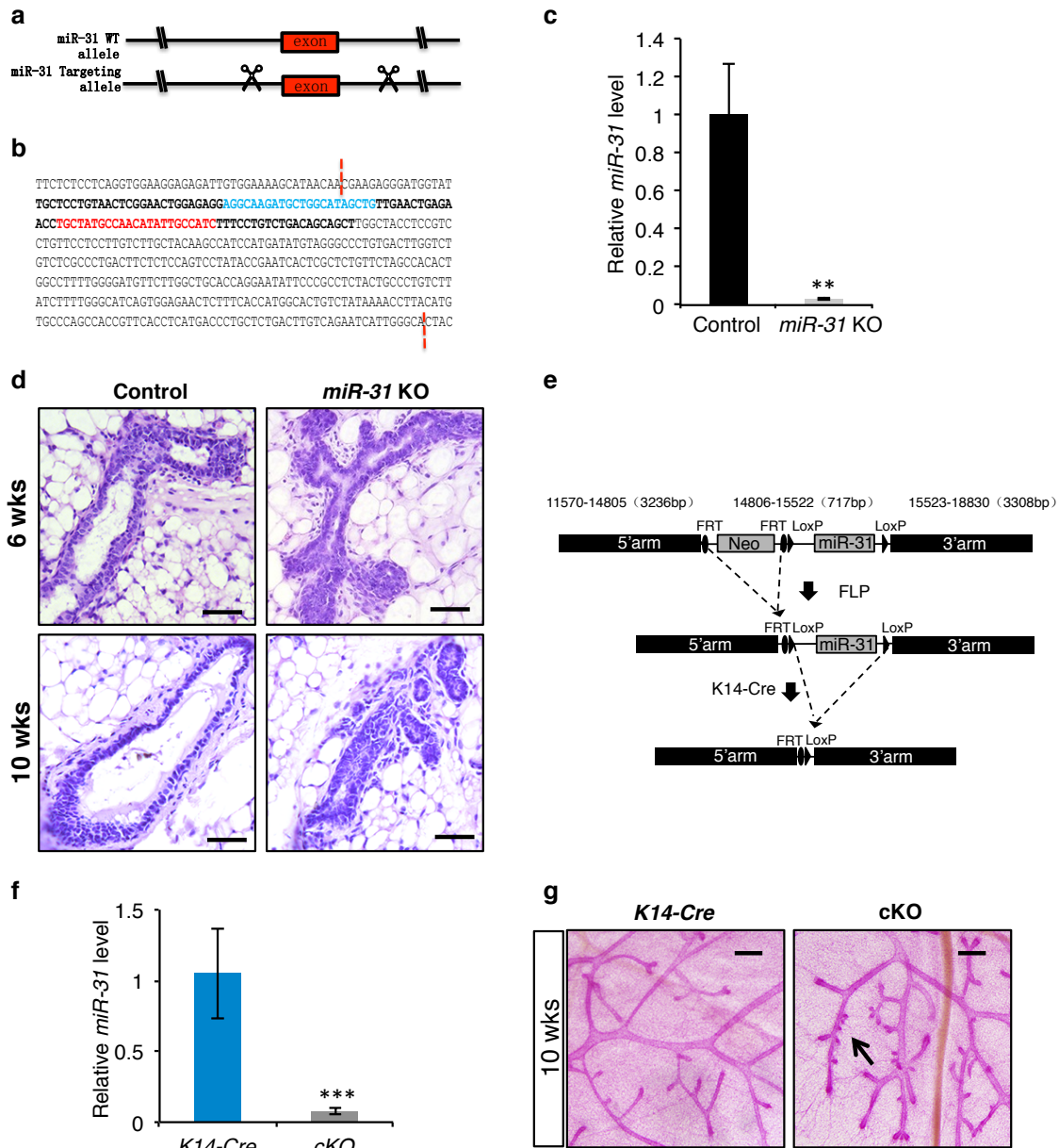
**Supplementary Figure 1. *MiR-31* is regulated by progesterone.** (a) Representative cell sorting profiles using CD24-PE-Cy7 and CD29-FITC in cell suspensions of WT mammary glands. qRT-PCR showing expression of basal marker gene, K14, and of luminal marker gene, K18, in CD24<sup>-</sup>CD29<sup>-</sup>, CD24<sup>-</sup>CD29<sup>+</sup>, CD24<sup>+</sup>CD29<sup>low</sup> and CD24<sup>+</sup>CD29<sup>high</sup> subpopulations. n = 3 biological replicates. (b) qRT-PCR analysis for *miR-31* in HC11 mouse mammary epithelial cells under conditions of vehicle, Estradiol, E+P1 (10nM Estradiol and 100 nM progesterone, n = 9 technicalbiological replicates), and E+P2 (10nM Estradiol and 1μM progesterone, n = 4 technicalbiological replicates). (c) qRT-PCR analysis for *miR-31* in HC11 mouse mammary epithelial cells treated with E+P

(10nM Estradiol and 100 nM progesterone) and/or Mifepristone. n = 3 technical replicates. **(d)** Immunofluorescence for RANKL in HC11 mouse mammary epithelial cells treated with Mifepristone, Estradiol and Progesterone (E+P) or vehicle control. n = 3 technical replicates. **(e)** Western blotting for p65 and p-p65 in HC11 mouse mammary epithelial cells treated with vehicle control or Estradiol and Progesterone (E+P). GAPDH was used as a loading control. n = 3 technical replicates. **(f)** Luciferase activity in lysates of HC11 mammary epithelial cells transfected with luciferase reporter plasmid of pGL3-basic, *miR-31* promoter and *miR-31* mutant promoter with mutation at the -1375 binding site, treated with Vehicle or Estradiol and Progesterone (E+P). n = 3 technical replicates. **(g)** Chromatin immunoprecipitation (ChIP) assay carried out on HC11 mammary epithelial cells using antibodies against p65 under indicated conditions. The enrichment of p65 binding to the site of -1375 bp in *miR-31* promoter was quantified using qPCR. **(h)** Immunohistochemistry for ER and PR in *PyVT* tumors and mammary ducts at 8 and/or 12 weeks of age. Scale bar: 50  $\mu$ m. n = 3 biological replicates. **(i)** Expression of *miR-31* across different breast tumors subtypes in TCGA RNA-Seq. **(j)** Pearson correlation analysis on *miR-31* and RANKL ( $P$  value =  $1.55e-14$ ;  $r = 0.3156$ ), as well as *miR-31* and  $TNF\alpha$  ( $P$  value =  $0.667e-07$ ;  $r = 0.21$ ) in breast cancer TCGA RNA-seq. Data represented as mean  $\pm$ S.D. Two tailed unpaired  $t$ -test for b, c, f, g (\* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.001$ ).



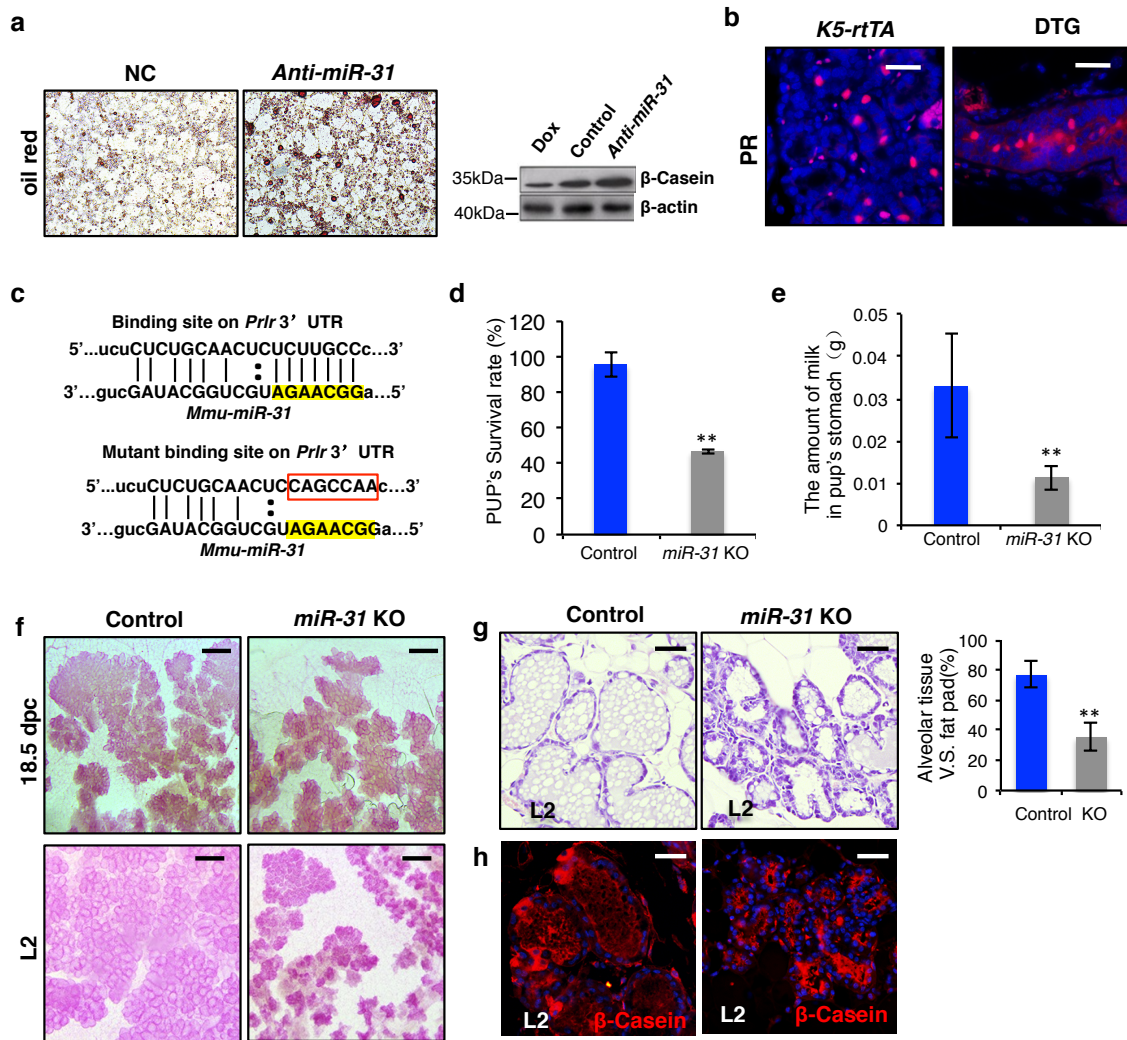
**Supplementary Figure 2. *MiR-31* induction causes impaired branching and mammary epithelial hyperplasia.** (a) Schematic maps of constructs used to generate *K5-rtTA/TRE-miR31* (DTG) double transgenic mice. (b) qRT-PCR for *miR-31* and *miR-205* in CD24<sup>+</sup>CD29<sup>high</sup> cells of *TRE-miR31*, *K5-rtTA* and DTG

transgenic mammary glands at 12 weeks of age following Dox treatment from 1 week of age. *n* = 3 technical replicates. **(c)** *In situ* Hybridization for *miR-31* in WT and DTG mammary ducts at 12 weeks of age following Dox treatment from 1 week of age. **(d-i)** Whole-mount staining of WT (*n* = 3), *K5-rtTA* (*n* = 3), *TRE-miR31* (*n* = 3) and DTG mammary glands at 12 weeks of age. DTG-1(**g**, *n* = 3), DTG-2 (**h**, *n* = 3) and DTG-3 (**i**, *n* = 3) represent 3 classes of DTG mammary gland phenotypes. The areas outlined by dashed boxes in panel **d** and **g** are shown at higher magnification in main Fig. 2a. Those dashed boxes in panels **h** and **i** are shown in **k** and **l**, respectively. Arrowheads indicate TEBs. Scale bars: 1mm. **(j)** Quantification of primary duct thickness (Left panel) and mammary branch points (Right panel) in Control (*n* = 3) and DTG (*n* = 3) mice at 12 weeks of age. Y axis in Left panel represents fold change of diameter in primary duct. **(k,l)** The higher images indicated by dashed boxes in (**h**) and (**i**). Scale bar: 0.2 mm. **(m)** H&E staining showing histology of *K5-rtTA* (*n* = 3) and DTG (*n* = 3) mammary glands at 12 weeks of age following Dox treatment at 1 week of age. Scale bar: 50  $\mu$ m. **(n)** qRT-PCR analysis for *miR-31* in *miR-31* inducible HC11 mammary epithelial cells following treatment of Dox. *n* = 3 technical replicates. **(o)** WST-8 cell proliferation assay showing that *miR-31* overexpression promotes cell proliferation, whereas *anti-miR-31* has no significant effect. *n* = 3 technical replicates. Data represented as mean  $\pm$  S.D. *n* = 3. Two tailed unpaired *t*-test for *j*, *n*, *o* (\**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.001).



**Supplemental Figure 3. Generation of constitutive *miR-31* null and a conditional *miR-31* null allele.** (a) Strategy to generate *miR-31* KO mice using CRISPR/Cas9 RNA-guided nucleases. (b) The 402 bp DNA fragment containing *miR-31* indicated by dash lines was deleted in the KO allele. The *miR-31* exon indicated by bold; mature *miR-31* indicated by blue, *miR-31\** indicated by red. (c) qRT-PCR analysis for *miR-31* in Control (n = 3) and *miR-31* KO (n = 3) mice. (d) H&E staining of Control (n = 3) and *miR-31* KO (n = 3) mammary gland at 6 weeks and 10 weeks of age. Scale bar: 50  $\mu$ m. (e) Strategy to generate *miR-31* conditional KO allele in mammary epithelium. (f) qRT-PCR analysis for *miR-31* in

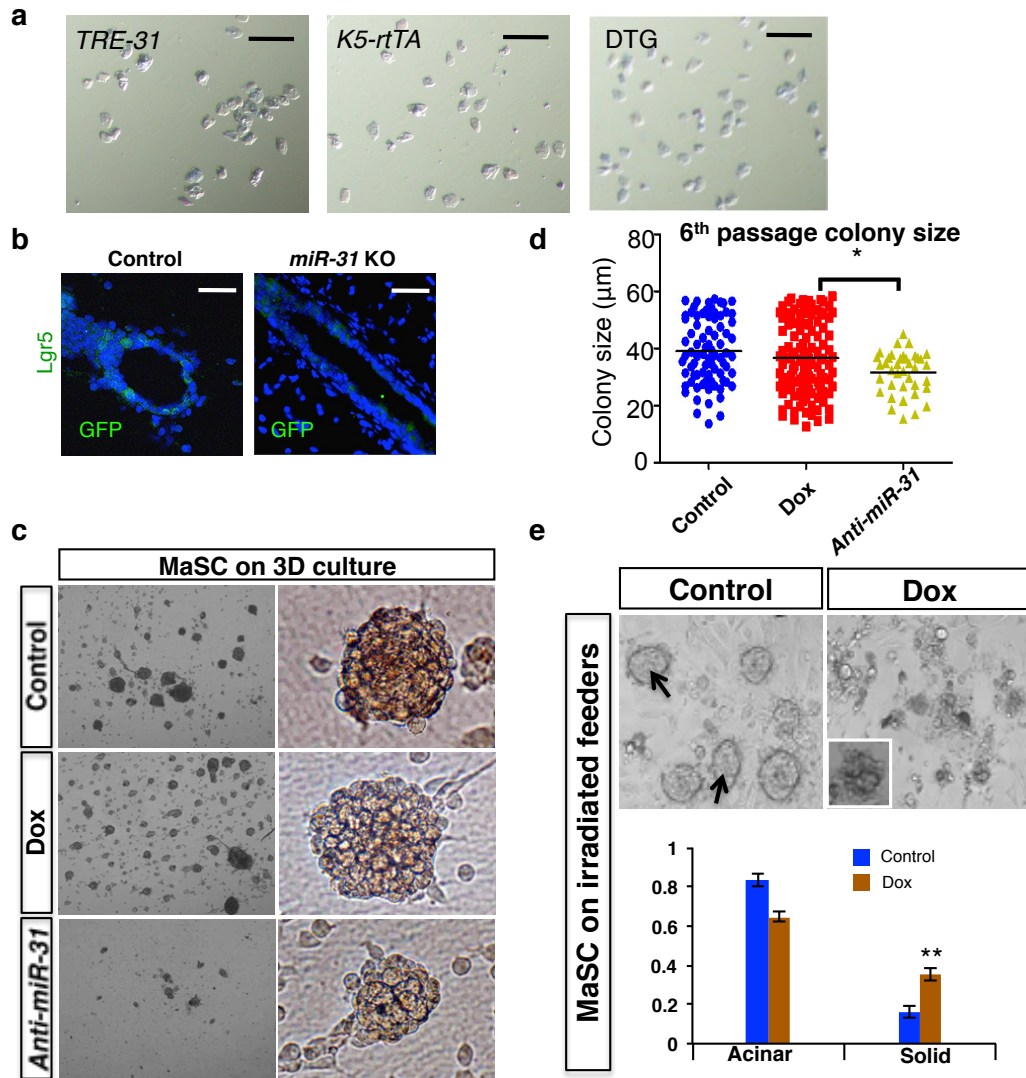
*K14-Cre* (n = 3) and cKO (*K14-Cre;miR-31<sup>f/f</sup>*, n = 3) mice. (g) Whole-mounts of mammary gland from *K14-Cre* (n = 3) and cKO (n = 3) mice at 10 weeks of age. Scale bar: 0.2 mm. Data represented as mean  $\pm$  S.D. n = 3. Two tailed unpaired *t*-test for c, f (\*\**P* < 0.01; \*\*\**P* < 0.001). Note: The schematic depiction of the general strategy for generating *miR-31* mutant mice (**Supplementary Figure 3a and b**) was also used in another unrelated study on the role of *miR-31* in intestinal stem cells and colorectal cancer, and the manuscript is currently accepted in *eLife*.



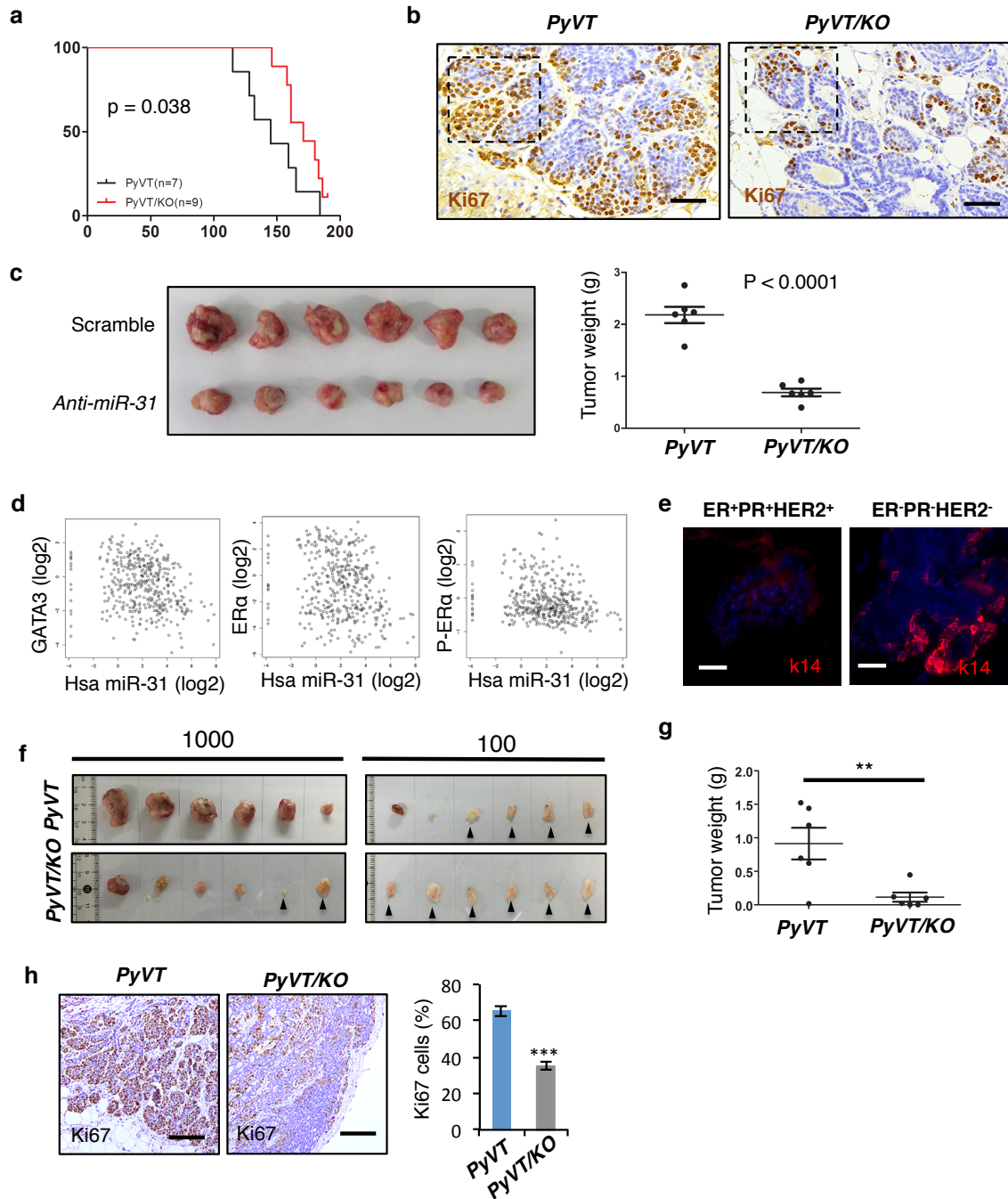
**Supplementary Figure 4. *MiR-31* is important for normal alveogenesis during pregnancy.** (a) Red Oil staining in HC11 mouse mammary epithelial cells infected with scramble RNA (NC) and *anti-miR-31* after lactogenic hormone-induced cell differentiation (Left panel). Western blot analysis showing reduced  $\beta$ -casein protein levels in *miR-31* overexpressing HC11 cells (Dox induced *miR-31* over-expression) and increased  $\beta$ -casein protein in *anti-miR-31* infected HC11 cells after lactogenic hormone-induced cell differentiation (Right panel).  $\beta$ -actin was used as a loading control. (b) Immunofluorescence for PR in *K5-rtTA* (n = 3) and DTG (n = 3) mammary glands at 14.5 dpc following Dox treatment at 3 weeks of age. Scale bar: 25  $\mu$ m. (c) Top panel indicates *miR-31* binding site in wild type *Prlr* 3'UTR regions. Bottom panel depicts mutant *miR-31* binding site. Seed sequences are marked by yellow. Red box indicates the mutated *miR-31* binding sites in the *Prlr* 3'UTR. (d) Statistical analysis of survival rate of pups raised by WT (Control, n = 3) and *miR-31* KO (n = 3) mothers at weaning age.

KO, n = 21; WT, n = 22. **(e)** Quantification of milk amount from pup's stomach at lactation day 2, in which pups raised by WT (Control, n = 14) and *miR-31* KO (n = 12) mothers. **(f)** Whole mount analysis for Control (n = 3) and *miR-31* KO (n = 3) mammary glands at 18.5 dpc and lactation day 2 (L2). Scale bar: 0.2 mm. **(g)** H&E staining for Control (n = 3) and *miR-31* KO (n = 3) mammary gland at lactation day 2 (L2). Statistics of proportion of mammary alveolar tissue v.s. fat pad. Scale bar: 25  $\mu$ m. **(h)** Immunofluorescence for  $\beta$ -Casein in Control (n = 3) and *miR-31* KO (n = 3) mammary glands at lactation day 2 (L2). Scale bar: 25  $\mu$ m. Data represented as mean  $\pm$  S.D. n  $\geq$  3. Two tailed unpaired *t*-test for d, e, g (\*\**P* < 0.01).





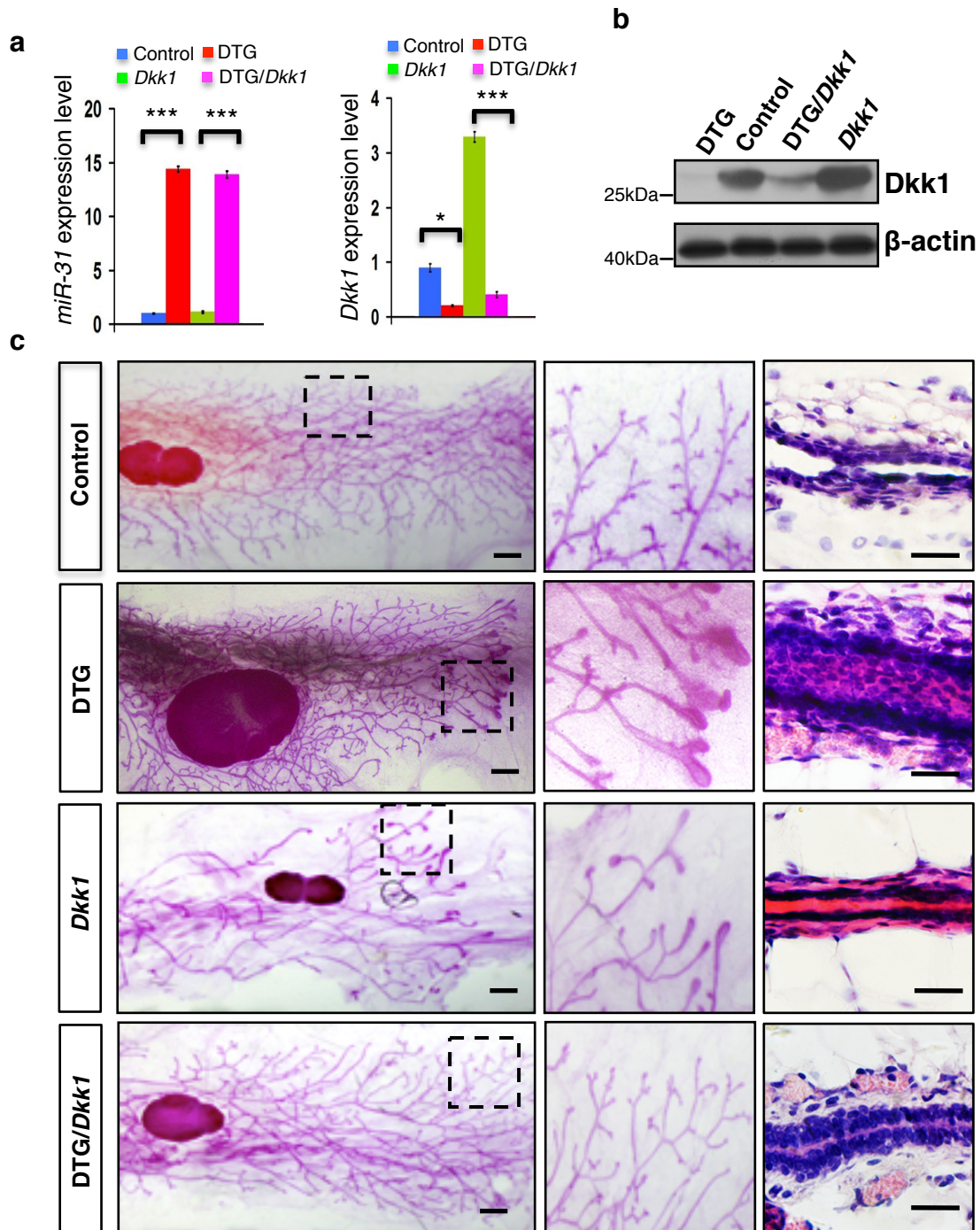
**Supplementary Figure 5. *MiR-31* induces mammary basal stem cell expansion.** (a) Images of vaginal smears taken from *TRE-miR31*, *K5-rtTA* and DTG mice at proestrus phase, related to Fig. 4a. (b) GFP immunofluorescence in mammary ducts from *Lgr5-eGFP-CreER* (Control, n = 3) and *Lgr5-eGFP-CreER;miR-31* KO (*miR-31* KO, n = 3) mice. Scale bar: 50 µm. (c) Representative images of colonies formed on culture inserts under conditions of Control (Scramble RNA), Dox treatment (Dox induced *miR-31* overexpression) and *anti-miR-31* transfection. High magnification images of single colonies are shown in the right panels. (d) Colony sizes at 6<sup>th</sup> passage under the indicated conditions. n = 3 technical replicates. (e) Representative images of colonies formed on irradiated feeder layers in the presence or absence of Dox. Arrows indicate acinar colonies. Inset shows a solid colony. The graph shows statistical results of solid colony and acinar colony formation in the presence or absence of Dox. Data represented as mean ± S.D. n = 3. Two tailed unpaired *t*-test for d, e (\**P* < 0.05; \*\**P* < 0.01).



**Supplementary Figure 6. Loss of *miR-31* resulted in compromised tumor growth, reduced cancer stem cells and impaired lung metastasis.** (a) The statistical analysis on overall survival rate of *PyVT* ( $n = 7$ ) and *PyVT/KO* ( $n = 9$ ) mice. (b) Immunohistochemistry for Ki67 in *PyVT* ( $n = 3$ ) and *PyVT/KO* ( $n = 3$ ) tumors at 12 weeks of age. The higher magnification images indicated by dashed boxes were shown in Fig. 5d. Scale bar: 50  $\mu\text{m}$ . (c) Gross appearance of engrafted tumors with  $5 \times 10^4$  4T1 ( $n = 6$  mice) and *anti-miR-31*-treated 4T1 ( $n = 6$  mice) mouse breast cancer cells engrafted 4 weeks post transplantation.

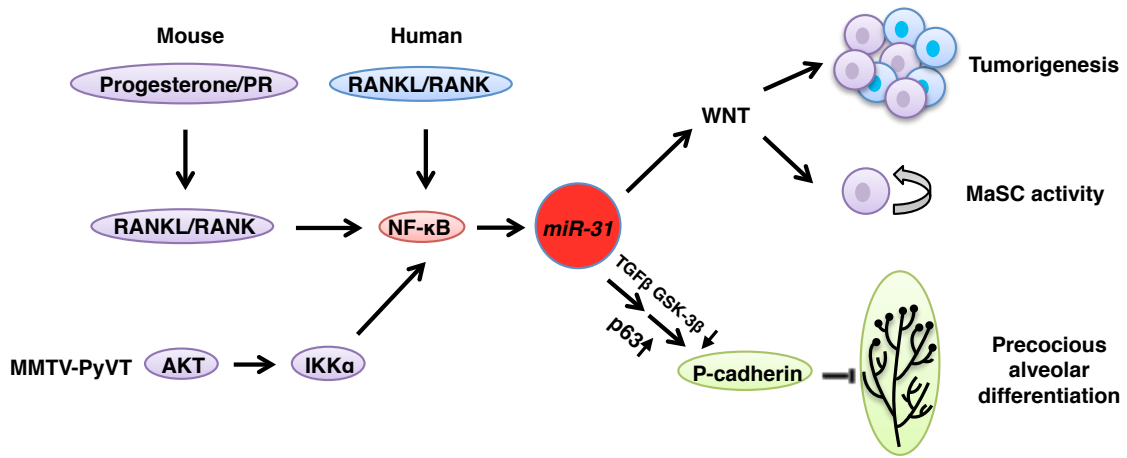
Quantification of tumor weight of the engrafted tumors. **(d)** Pearson correlation analysis on *miR-31* and GATA3 ( $P = 8.99e-08$ ,  $r = -0.26$ ), ER $\alpha$  ( $P = 1.74e-06$ ,  $r = -0.23$ ), p-ER $\alpha$  ( $P = 1.88e-05$ ,  $r = -0.21$ ) in TCGA breast cancer RNA-Seq, respectively. **(e)** Immunofluorescence for K14 in ER<sup>+</sup>PR<sup>+</sup>HER2<sup>+</sup> and ER<sup>-</sup>PR<sup>-</sup>HER<sup>-</sup> human breast tumors. Scale bar: 25  $\mu$ m. **(f)** Gross appearance of tumors transplanted with 1000 ( $n = 6$  mice) and 100 ( $n = 6$  mice) of *PyVT* and *PyVT/KO* tumor cells, engrafted 8 weeks post transplantation. Arrowheads point to fat pads (not tumors). Related to Fig. 5j. **(g)** Quantification of tumor weight of the engrafted tumors transplanted with 1000 *PyVT* and *PyVT/KO* tumor cells in Panel **f**. **(h)** Immunohistochemistry for Ki67 and quantification of Ki67 positive cells in the xenografted tumors transplanted with 1000 *PyVT* and *PyVT/KO* tumor cells in Panel **f**. Scale bar: 100  $\mu$ m.  $n = 3$  biological replicates. Data represented as mean  $\pm$  S.D.  $n \geq 3$ . Two tailed unpaired *t*-test for c, g, h ( $*P < 0.05$ ;  $**P < 0.01$ ;  $***P < 0.001$ ). Survival curves were estimated by the Kaplan-Meier method and compared using the Wilcoxon test for panel **a**.



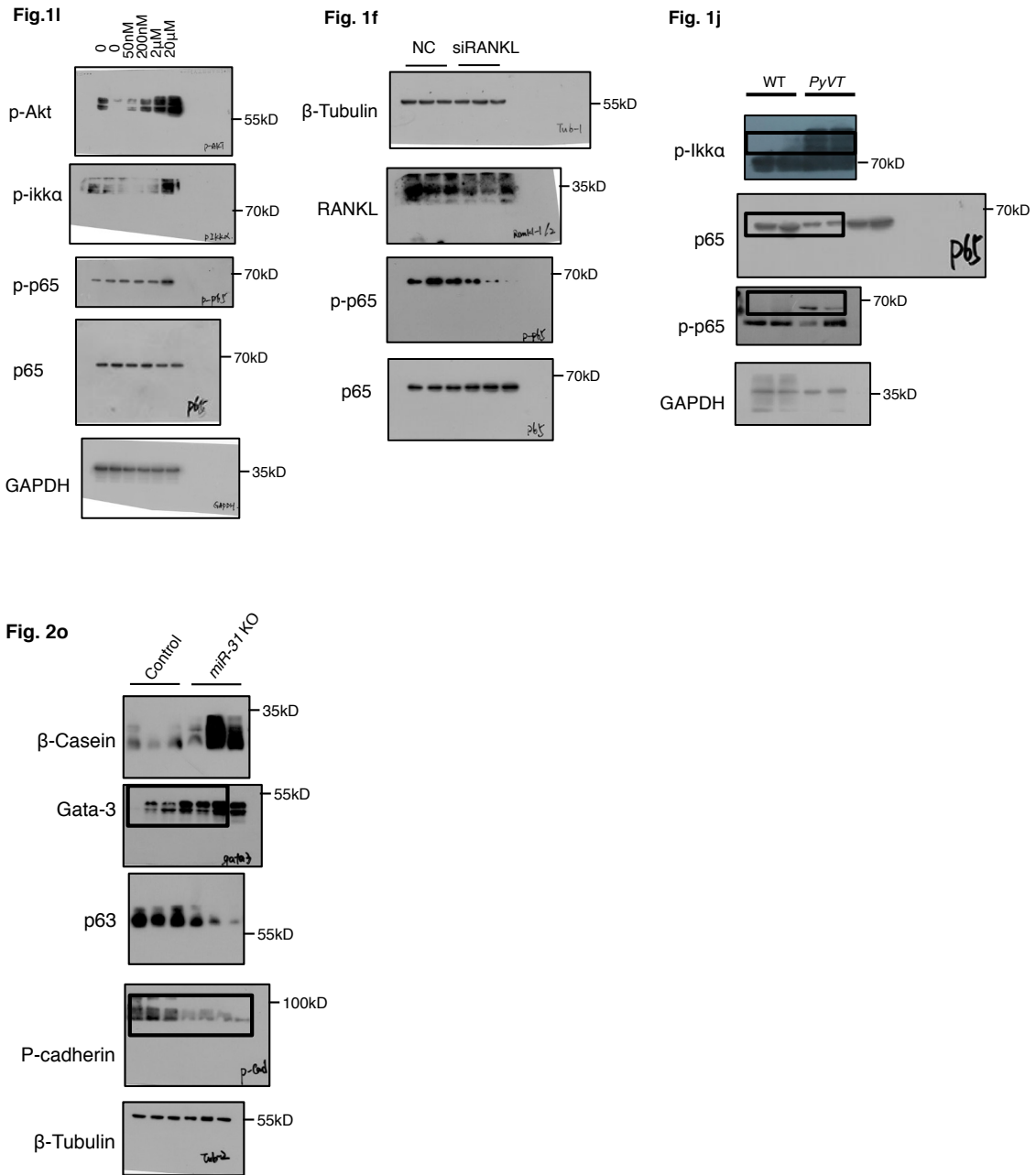


**Supplementary Figure 8. *Dkk1* induction can partially rescue DTG mammary phenotype.** (a,b) qRT-PCR for *miR-31* and *Dkk1* (a), and Western blotting for *Dkk1* (b) in Control (*K5-rtTA*), DTG (*K5-rtTA;TRE-miR31*), *Dkk1* (*K5-rtTA;TRE-Dkk1*), and DTG/*Dkk1* (*K5-rtTA;TRE-miR31;TRE-Dkk1*) mammary glands at 12 weeks of age following Dox induction at 1 week of age.  $\beta$ -actin was used as a loading control. (c) Whole-mount staining of mammary glands from *K5-rtTA*, DTG, *Dkk1* and DTG/*Dkk1* mice at 12 weeks of age following Dox induction

at 1 week of age. Regions outlined by dashed boxes are shown at higher magnification in the respective middle panels. Histology of the corresponding mammary ducts is shown in the right panels. 3 *K5-rtTA*, 3 DTG, 3 *Dkk1* and 3 DTG/*Dkk1* mice were analyzed. Scale bar: Left panels, 1 mm; Right panels: 25  $\mu\text{m}$ . Data represented as mean  $\pm$ S.D.  $n = 3$ . Two tailed unpaired *t*-test for a (\* $P < 0.05$ ; \*\*\* $P < 0.001$ ).



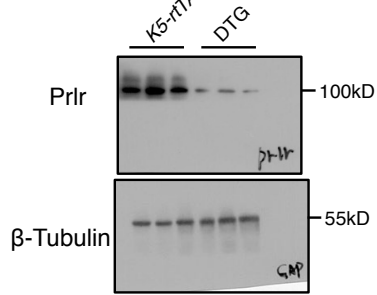
**Supplementary Figure 9. The working model of *miR-31* in regulating mammary gland development, MaSC activity and breast tumorigenesis.**



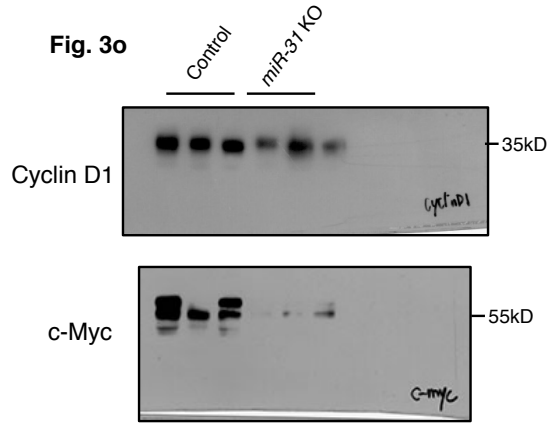
**Supplementary Figure 10. Uncropped blots for western blot analysis in the main figures.**



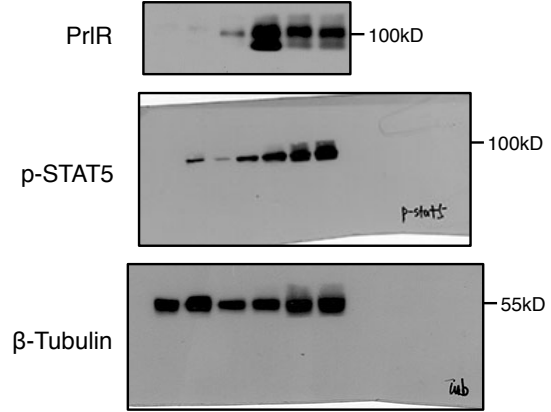
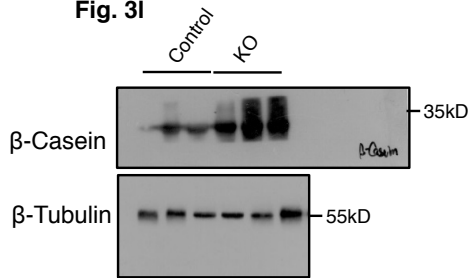
**Fig. 3g**



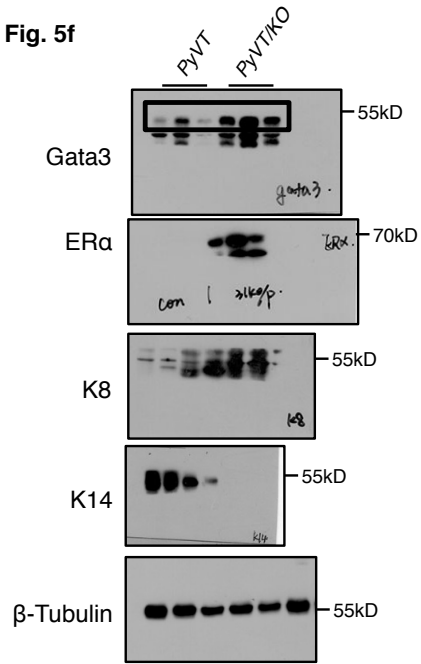
**Fig. 3o**



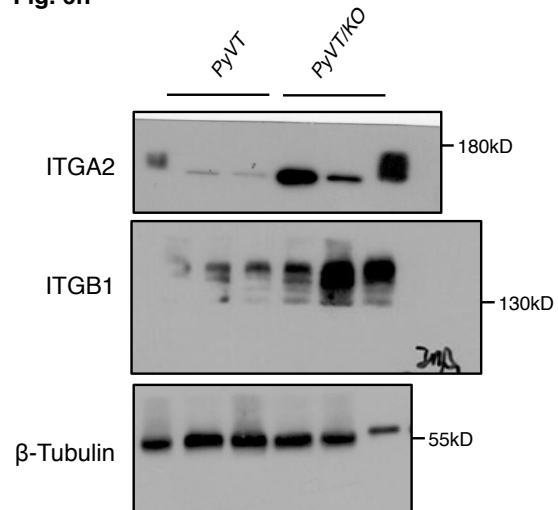
**Fig. 3l**



**Fig. 5f**



**Fig. 6h**



Supplementary Fig. 10 continued

Fig. 7c

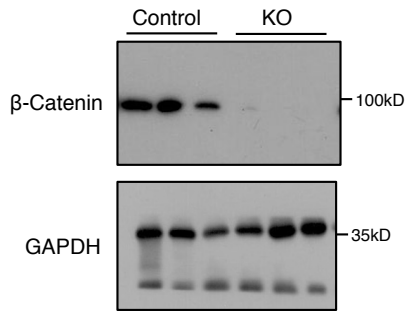


Fig. 7g

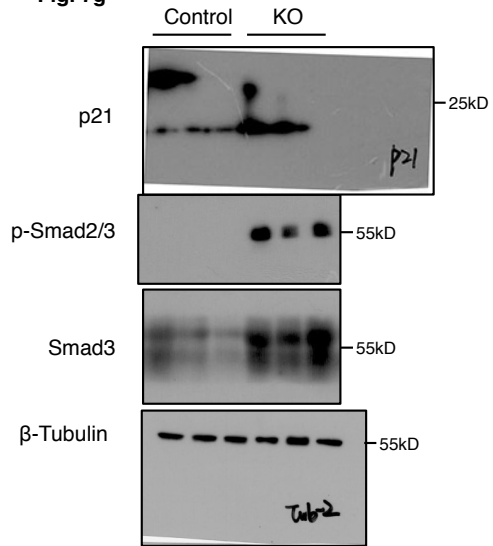


Fig. 8c

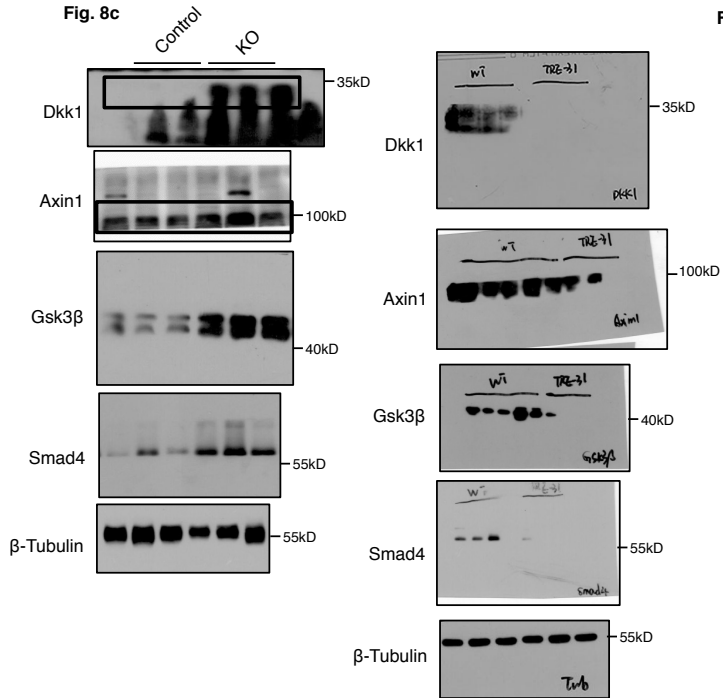
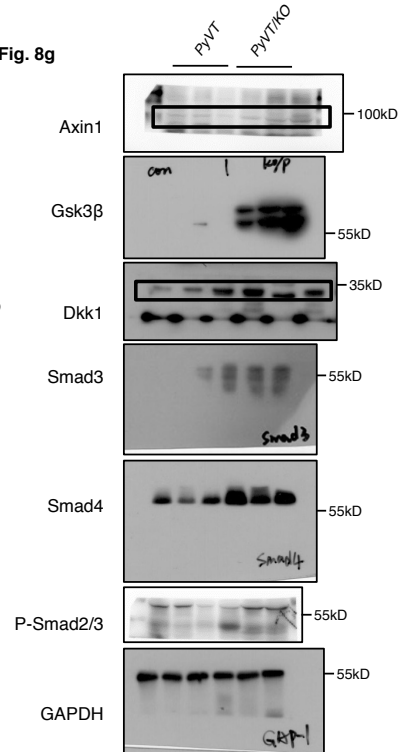


Fig. 8g



Supplementary Fig. 10 continued