

Fig. S1. Retraction fiber & migrasome (R&M) of glioblastoma cells and quality control of 2 purified R&Ms. A LN229 cell expressing EGFP-CD9 produced R&Ms (upper panel) and 3 4 produced few migrasomes (lower panel). Gray, EGFP-CD9. Scale bar, 20 µm. B Time-lapsed live-cell imaging (captured in every 30 sec) of U87MG EGFP-CD9 cells after treatment of 5 6 0.015% trypsin/EDTA. Green, EGFP-CD9. Scale bars, 20 µm. C Quality control of the R&M 7 purification stages in Fig. 1A. Each quality control sample were obtained and observed immediately. The samples designated in each image ("A" ~ "E") are paralleled samples 8 described in R&M purification stages of Fig. 1A. The image "F" represents the debris obtained 9 10 after trypsinization procedure and the image "G" represents the purified R&M obtained by using Exosome Purification Reagent. Green, EGFP-CD9. Scale bars, 20 µm. D Scanning 11

electron microscope images of purified R&M. From "a" to "f", irregular and atypical purified
R&M samples are shown. Yellow arrows represent the retraction fiber-like structures. Scale
bars are indicated in each figure. E Transmission electron microscope images of attached cells
and purified R&M. From "a" to "e", transmission electron microscope imaging of attached
cells and purified U87MG-R&M and LN229-R&M were presented. R, retraction fiber; M,
migrasome; Cb, Cell body. Scale bars, 200 µm.



19 Fig. S2. Inhibition of autophagosome/lysosome fusion by chemical drugs and genetic ablation. A Western blotting of SQSTM1, LC3B, and β-actin proteins in and LN229 cells. Cells were 20 treated with chloroquine (CQ; 50 µM, 12 h) or bafilomycin A1 (BafA1; 50 nM, 12 h). W(M), 21 molecular weight. **B** qRT-PCR experiment for quantify mRNA levels of STX17 and SNAP29 22 in LN229 cells transfected with STX17 or SNAP29 siRNAs. \*\*\* indicates p < 0.001. Data are 23 expressed as mean  $\pm$  SEM. Student's t-test was used to analyze the statistical significance 24 between each group (n = 3). C Quantification of the number of RFs per a cell. Image analyses 25 were performed using results from Fig. 3H. \*\* indicates p < 0.01. Data are expressed as mean 26 ± SEM. The unpaired nonparametric Mann-Whitney U test was used to analyze the statistical 27 significance between each group (n = 10). **D** Western blotting of SQSTM1, LC3B, and  $\beta$ -actin 28 proteins in and LN229 cells with siSTX17 or siSNAP29 (left panel). gRT-PCR experiment set 29 of cells used in Western blotting for quantify mRNA levels of STX17 and SNAP29. \*\* indicates 30 p < 0.01. \*\*\* indicates p < 0.001 (right panel). Data are expressed as mean  $\pm$  SEM. Student's 31 t-test was used to analyze the statistical significance between each group (n = 3). E Tandem-32 fluorescent LC3B expressed in LN229 cells were used for performing knockdown of STX17 33 and SNAP29. GFP<sup>-</sup>/RFP<sup>+</sup> vesicles represent autolysosomes and GFP<sup>+</sup>/RFP<sup>+</sup> vesicles represent 34 autophagosomes (upper panel). Scale bars, 20 µm. Colocalization analysis was performed for 35 investigating colocalized pattern of GFP and RFP (lower panel). F Tandem-fluorescent LC3B 36 37 was expressed in LN229 cell. Live-cell imaging performed in NaAsO<sub>2</sub> (AS; 10 µM, 12 h) treatment condition. Scale bars, 20 µm. 38



Fig. S3. Reinforcement of autophagy under the disturbed autophagic flux induces R&M 40 formation. A Live-cell imaging of LN229 cells. Cells were treated with Torin-1 (250 nM, 2 h) 41 and/or chloroquine (CQ; 50 µM, 12 h). The number of cells imaged for quantifying retraction 42 fibers (RFs) and migrasomes are as follows: n = 20 for DMSO, n = 18 for Torin-1, n = 17 for 43 CQ, and n = 18 for Torin-1/CQ. Scale bars, 20 µm. **B** Western blotting of SQSTM1, LC3B, 44 and β-actin under treatment with Torin-1 and/or CQ. C Quantification of the number of RFs 45 per a cell. Image analyses were performed using results from Fig. S3A. D Quantification of the 46 number of migrasomes per RF (100 µm). Image analyses were performed using the results 47 from Fig. S3A. E qRT-PCR experiment set of cells used in Fig. 5 and 6 for determining the 48 knockdown efficiency of both *ITGA5* and *TSPAN4*. \* indicates p < 0.05. \*\* indicates p < 0.01. 49 \*\*\* indicates p < 0.001 (n = 3). 50

39

## A Uncropped blots (Fig. 2C)

Integrin a5



SQSTM1 (p62)



LC3B



51

## CD63



α-tubulin



## RPL4



## RPS13



GAPDH



B Uncropped blots (Fig. 4C)



C Uncropped blots (Fig. S2A)

SQSTM1 (p62)

- 53
- D Uncropped blots (Fig. S2D)

SQSTM1 (p62)



LC3B (short expose)



E Uncropped blots (Fig. S3B)

SQSTM1 (p62), β-actin



Fig. S4. Uncropped blots for Western blot experiments. A Uncropped blots of Fig. 2C. Western 55 blotting was conducted by immunoblotting against the following antibodies: integrin  $\alpha 5$ , 56 SQSTM1 (p62), LC3B, α-tubulin, RPL4, RPS13, and GAPDH. **B** Uncropped blots of Fig. 4C. 57 Western blotting was performed by immunoblotting against the following antibodies: 58 SQSTM1 (p62), LC3B, p-eIF2α (Ser51), and β-actin. C Uncropped blots of Fig. S2A. Western 59 blotting was conducted by immunoblotting against the following antibodies: SQSTM1 (p62), 60 LC3B, and β-actin. **D** Uncropped blot images of Fig. S2D. Western experiments was performed 61 by using the following antibodies: SQSTM1 (p62), LC3B, and  $\beta$ -actin. E Uncropped blot 62 images of Fig. S3B. Western blotting was performed by using the following antibodies: 63 SQSTM1 (p62), LC3B, and  $\beta$ -actin. 64

Gene	Species	Direction	Sequence (5'-3')
18S rRNA	Homo	Forward	CAGCCACCCGAGATTGAGCA
	Sapiens	Reverse	TAGTAGCGACGGGGGGGTGTG
ITGA5	Homo	Forward	AGCAAGAGCCGGATAGAGGA
	Sapiens	Reverse	TCAGGGCATTCTTGTCACCC
TSPAN4	Homo	Forward	TGGGTGCCATCAAGGAGAAC
	Sapiens	Reverse	CTTGTCCGTGTAGGCGAAGA
STX17	Homo	Forward	GGGGAATGGTGTGGTGCTAA
	Sapiens	Reverse	TAGACATGCAGTTGGGCTGG
SNAP29	Homo	Forward	CTGGCCCTCATGTACGAGTC
	Sapiens	Reverse	AGGGTGCCATTCTGTTCAGG
XBP1s	Homo	Forward	GCTGAGTCCGCAGCAGGT
	Sapiens	Reverse	CTGGGTCCAAGTTGTCCAGAAT

Table S1. The primers used in this study.