Online Resource 1: DNeasy PowerSoil Pro Kit modified extraction protocol. Step 1 to 8 modified protocol ; step 9 to 18 manufacturer's protocol.

1. Gently crush a small piece of coral in a mortar and pestle (~100mg, in order not to overload the column though). And transfer in a PowerBead Pro Tube (or 2 ml Microcentrifuge).
2. Add 800 µl of buffer CD1 and vortex briefly.
3. Add 83 µl Proteinase K (e.g. from Qiagen Blood & Tissue Kit) and vortex briefly.
4. Incubate at 56 degrees for 60 minutes. Vortex for 10 seconds every 15 minutes,
   - Ideally, do this on a shaking heat block, with shaking on full).
5. Add 200ul buffer CD2 and vortex for 5 seconds.
6. Microfuge at full for 1 minute (at room temperature).
7. Transfer supernatant to new tube (Max 700 µl).
8. Add 600 µl of buffer CD3, vortex for 5 seconds
9. Load 650 µl of the lysate onto an MB Spin Column and centrifuge at 15,000 x g for 1 min.
10. Discard the flow-through and repeat step 8 to ensure that all of the lysate has passed through the MB Spin Column.
11. Carefully place the MB Spin Column into a clean 2 ml Collection Tube. Avoid splashing any flow-through onto the MB Spin Column.
12. Add 500 µl of Solution EA to the MB Spin Column. Centrifuge at 15,000 x g for 1 min.
13. Discard the flow-through and place the MB Spin Column back into the same 2 ml Collection Tube.
14. Add 500 µl of Solution C5 to the MB Spin Column. Centrifuge at 15,000 x g for 1 min.
15. Discard the flow-through and place the MB Spin Column into a new 2 ml Collection
16. Centrifuge at up to 16,000 x g for 2 min. Carefully place the MB Spin Column into a new 1.5 ml Elution Tube.
17. Add 50–100 µl of Solution C6 to the center of the white filter membrane.
18. Centrifuge at 15,000 x g for 1 min. Discard the MB Spin Column. The DNA is now ready for downstream applications.