1	The establishment of species-specific primers for the molecular identification of ten
2	stored-product psocids based on ITS2 rDNA
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4	Zi-Hua Zhao ^{1,§} , Bing-Yi Cui ^{1,§} , Zhi-Hong Li ^{1,*} , Fan Jiang ^{1, 5} , Qian-Qian Yang ^{1,6} ,
5	Zuzana Kučerová ² , V áclav Stejskal ² , George Opit ³ , Yang Cao ⁴ , Fu-Jun Li ⁴
6	
7	¹ Department of Entomology, College of Plant Protection, China Agricultural
8	University, Beijing 100193, China;
9	² Crop Research Institute, Drnovsk á 507, 161 06 Prague 6, Czech Republic;
10	³ Department of Entomology and Plant Pathology, 127 Noble Research Center,
11	Oklahoma State University, Stillwater, OK 74078, USA;
12	⁴ Academy of State Administration of Grain, Beijing 100037, China;
13	⁵ Institute of Plant Quarantine Research, Chinese Academy of Inspection and
14	Quarantine, Beijing 100176, China;
15	⁶ College of Life Science, China Jiliang University, Hangzhou 310018, China;
16	

[§]These two authors (Zi-Hua Zhao and Bing-Yi Cui) contributed equally to this research. Address: No. 2 Yuanmingyuan West Road, Haidian District, Beijing 100193, China *Corresponding author: <u>lizh@cau.edu.cn</u> (Zhi-Hong Li)

17 Supplement Material Appendix

Table S1 The primers for the ITS2 gene amplification of 10 stored-product psocids
 18 Table S2 The ITS2 gene sequence of 10 stored-product psocids (The sequences 19 highlighted in red indicate species-specific primers for ten stored-product psocids) 20
Table S3 The number of 53 specific sequences submitted to Genebank
 21 Figure S1 The electrophoresis strip for specificity estimation of species-specific 22 primers (LBF and LBR) of L. bostrychophila (M: D2000 DNA Marker; 1: L. 23 bostrychophila (Guangxi); 2: L. bostrychophila (Henan); 3: L. bostrychophila 24 (Chongqing); 4: L. bostrychophila (Berlin); 5: L. bostrychophila (USA); 6: L. 25 bostrychophila (Prague); 7 L. bostrychophila (Beijing); 8: L. bostrychophila 26 (Guangzhou); 9: L. entomophila (Shandong); 10: L. decolor (Yunnan); 11: L. paeta 27 28 (Zhejiang); 12: L. corrodens (USA); 13: L. brunnea (Prague); 14: L. rufa (USA); 15: L. pearmani (USA); 16: L. mendax (Jiangsu); 17: L. tricolor (Shandong); CK: 29 Negative control) 30 31 Figure S2 The electrophoresis strip for specificity estimation of species-specific primers (21LEnF and 208LEnR-2) of L. Entomophila (M: DNA Marker II; 1: L. 32 entomophila (Beijing); 2: L. entomophila (Guangxi); 3: L. entomophila (Prague); 4: L. 33 entomophila (Chongqing); 5: L. bostrychophila (Guangxi); 6: L. bostrychophila 34

- 35 (Yunnan); 7: L. Decolor (Yunnan); 8: L. corrodens (USA); 9: L. brunnea (Prague); 10:
- 36 L. rufa (USA); 11: L. pearmani (USA); 12: L. mendax (Jiangsu); 13: L.
- 37 *tricolor*(Shandong); CK: Negative control)
- 38 Figure S3 The electrophoresis strip for specificity estimation of species-specific

39	primers (164LDeF and 319LDeR) of L. decolor (M: DNA Marker II; 1 and 2: L.
40	Decolor (Yunnan); 3 and 4: L. Decolor (Chongqing); 5: L. Entomophila (Shandong);
41	6: L. bostrychophila (Chongqing); 7: L. paeta (Shandong); 8: L. corrodens (USA); 9:
42	L. Brunnea (Prague); 10: L. Rufa (USA); 11: L. mendax (Jiangsu); 12: L. Pearmani
43	(USA); 13: L. Tricolor (Shandong); CK: Negative control)
44	Figure S4 The electrophoresis strip for specificity estimation of species-specific
45	primers (LPa15F and LPa180R) of L. paeta (M: DNA Marker II; 1: L. paeta
46	(Caoxian, Shandong); 2: L. paeta (Taian, Shandong); 3: L. paeta (Zhejiang); 4: L.
47	paeta (Hubei); 5: L. paeta (USA); 6: L. entomophila (Shandong); 7: L. bostrychophila
48	(Guangxi); 8: L. decolor (Yunnan); 9: L. corrodens (Prague); 10: L. brunnea (USA);
49	11: L. Rufa (USA); 12: L. pearmani (USA); 13: L. mendax (Jiangsu); 14: L. tricolor
50	(Shandong); CK: Negative control)
51	Figure S5 The electrophoresis strip for specificity estimation of species-specific
52	primers (LC170F and LC277R) of L. corrodens (M: D2000 DNA Marker; 1: L.
53	corrodens (Danmark); 2: L. corrodens (Prague); 3: L. corrodens (USA); 4: L.
54	entomophila (Shandong); 5: L. bostrychophila (Guangzhou); 6: L. decolor
55	(Chongqing); 7: L. paeta (Hubei); 8: L. brunnea (USA); 9: L. rufa (USA); 10: L.
56	tricolor (Shandong); 11: 11: L. mendax (Jiangsu); 12: L. pearmani (USA); CK:
57	Negative control)

Figure S6 The electrophoresis strip for specificity estimation of species-specific
primers (LBr350F and LBr577R) of *L. brunnea* (M: D2000 DNA Marker; 1: *L. brunnea* (Prague); 2: *L. brunnea* (USA); 3: *L. entomophila* (Beijiing); 4: *L.*

- 61 bostrychophila (Guangzhou); 5: L. decolor (USA); 6: L. paeta (Shandong); 7: L.
- 62 corrodens (USA); 8: L. rufa (USA); 9: L. pearmani (USA); 10: L. tricolor (Shandong);
- 63 11: *L. mendax* (Jiangsu); CK: Negative control)
- 64 Figure S7 The electrophoresis strip for specificity estimation of species-specific
- primers (78LRuF-3 and 276LRuR-3) of L. rufa (M: D2000 DNA Marker II; 1, 2,
- and 3: L. rufa (USA); 4: L. entomophila (Beijiing); 5: L. bostrychophila (Guangxi); 6:
- 67 L. decolor (Yunnan); 7: L. paeta (Prague); 8: L. corrodens (USA); 9: L. brunnea
- 68 (USA); 10: L. mendax (Jiangsu); 11: L. pearmani (USA); 12: L. tricolor (Shandong);
- 69 CK: Negative control)

71	Figure S8 The electrophoresis strip for specificity estimation of species-specific
72	primers (186LPeF and 436LPeR) of L. pearmani (M: D2000 DNA Marker; 1 and 2: L.
73	pearmani(USA); 3: L. entomophila (Beijiing); 4: L. bostrychophila (Guangxi); 5: L.
74	decolor (Yunnan); 6: L. paeta (Prague); 7: L. corrodens (USA); 8: L. brunnea (USA);
75	9: L. rufa (USA); 10: L. mendax (Jiangsu); 11: L. tricolor (Shandong); CK: Negative
76	control)
77	Figure S9 The electrophoresis strip for specificity estimation of species-specific

- primers (LM60F and LM224R) of *L. mendax* (M: D2000 DNA Marker; 1 and 2: *L.*
- 79 mendax (Jiangsu); 3: L. entomophila (Beijiing); 4: L. bostrychophila (Guangxi); 5: L.
- 80 decolor (Yunnan); 6: L. paeta (Prague); 7: L. corrodens (USA); 8: L. brunnea (USA);
- 9: *L. rufa* (USA); 10: *L. pearmani* (USA); 11: *L. tricolor* (Shandong); CK: Negative
 control)

83	Figure S10 The electrophoresis strip for specificity estimation of species-specific
84	primers (LTri20F and LTri249R) of L. tricolor (M: D2000 DNA Marker; 1 and 2: L.
85	tricolor (Shandong); 3: L. entomophila (Beijiing); 4: L. bostrychophila (Guangxi); 5:
86	L. decolor (Yunnan); 6: L. paeta (Prague); 7: L. corrodens (USA); 8: L. brunnea
87	(USA); 9: L. rufa (USA); 10: L. mendax (Jiangsu); 11: L. pearmani (USA); CK:
88	Negative control)

- Figure S11 The sensitivity estimation of species-specific primers with different
 concentration of template DNA of *L. bostrychophila* (M: DNA Marker II; 1: 40ng; 2:
 20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8: Negative control)
- 92 Figure S12 The sensitivity estimation of species-specific primers with different

93	concentration of template DNA of L. entomophila (M: D2000 DNA Marker; 1: 40ng;
94	2: 20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8: Negative control)
95	Figure S13 The sensitivity estimation of species-specific primers with different
96	concentration of template DNA of L. decolor (M: D2000 DNA Marker; 1: 40ng; 2:
97	20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8: Negative control)
98	Figure S14 The sensitivity estimation of species-specific primers with different
99	concentration of template DNA of L. paeta (M: D2000 DNA Marker; 1: 40ng; 2: 20ng;
100	3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8: Negative control)
101	Figure S15 The sensitivity estimation of species-specific primers with different
102	concentration of template DNA of L. corrodens (M: D2000 DNA Marker; 1: 40ng; 2:
103	20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8: Negative control)
104	Figure S16 The sensitivity estimation of species-specific primers with different
105	concentration of template DNA of L. brunnea (M: D2000 DNA Marker; 1: 40ng; 2:
106	20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8: Negative control)
107	Figure S17 The sensitivity estimation of species-specific primers with different
108	concentration of template DNA of L. rufa (M: D2000 DNA Marker; 1: 40ng; 2: 20ng;
109	3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8: Negative control)
110	Figure S18 The sensitivity estimation of species-specific primers with different
111	concentration of template DNA of L. pearmani (M: D2000 DNA Marker; 1: 40ng; 2:
112	20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8: Negative control)
113	Figure S19 The sensitivity estimation of species-specific primers with different
11/	concentration of template DNA of L mendar (M: D2000 DNA Marker: 1: 40ng: 2:

- 115 20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8: Negative control)
- 116 Figure S20 The sensitivity estimation of species-specific primers with different
- 117 concentration of template DNA of *L. tricolor* (M: D2000 DNA Marker; 1: 40ng; 2:
- 118 20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8: Negative control)

Primer	名称	sequence
ITS2 rDNA	Upstream primer	5'- TGTGAACTGCAGGACACATG-3'
ITS2 rDNA	Downstream primer	5'- GTCTTGCCTGCTCTGAG-3'
ITS2 rDNA	Downstream primer	5'-GTCTTGTCTGATCTGAG-3'

121Table S1 The primers for the ITS2 gene amplification of 10 stored-product psocids

 Table S2 The ITS2 gene sequence of 10 stored-product psocids
 123 124 (1) L. entomophila 125 TCTGAGGGTCGGTGATCAATATTAAACCGAACGTTCCGTGTGATACGGAACTGATGGG 126 127 AAGCGAGTGCACGCACAAATTACAGAGTTCGGAAGAACTGTGCTTGCAGTGGTTACA 128 TTTTTAACGTCACGTTAGTAAGCGTAAAACTTTACCACGGACGTATAAAAGAACAACG 129 AATTCTATATCTACTCTTCATCGTC 130 131 132 (2) L. bostrychophila AACATCGATATTTCGAACGCACATTGCCGAGCTAGGATTTTTTCCGGCTCGACGTCTG 133 134 TCTGAGGGTCGATAATTTTATTAAAATAAAGTTTTCGTTGAGAATAAAATCTCACGGAC 135 TGTGGAAGTGTCGAAAGATTTGAGTTTTGCTCGAAACTTCGTCTTCTGAAAGTTAAGA **ACGAGAATACAAGGAGCG**AGCTCGTATTTTGCGGTTTTCTGCTTCACGTTAGTTTTGT 136 137 TTTACTGAGCGGATTGTCAGAAAAATCGTAAGAGTTCCTTTTGTAATCTTTTCATCGTC 138 139 (3) L. decolor 140 AACATCGATATTTCGAACGCACATTGCGCTCTGGGGGAACTCCGTTCCCGGAGCACGTCT GTCTGAGGGTCGGTGCTTAAATATAAATTTCGGAACGATTCTTTTTAACGGAAGGTCGT 141 GATCGAAACGTTGAAAGCTTCGGGTCGTAAGAAAACCCGTCTTTCGAAATGCACTAG 142 143 **AACCGAGA**ATCTTCGTAAATGAAGAAGGCGAGAGCTGTAAAGAGCAGCGATACGTGA ATCACGTTTACCGACAGGCGAACGATAACAAGCATGCCCTAAGCACGTCTTGTAAAAA 144 145 GGGCGAGAGTGCTTTAGGGGAAGCGGTTCGCATATATTTTTATAACTGTCGTATACGCG 146 147 ACGAAAAAGATTCTTTTCATCGTC 148 149 (4) *L. paeta* 150 AACATCGATATTTCGAACGCACATTGCCGAGTTTTTGGATTTATCCTGACTCGACGTCT 151 152 GTCTGAGGGTCGGTGATATCTATTGAAAGATATTTCGTTGTTACCGACGATTTTGAGAG TGTCAGATTCTGCATTGCAGAATTTGTCTTTTGAAATACGATGTCACGAGGTTCTTGCA 153 **AAAGGGTAACTGCATCGGACGGT**ATTTGGTGTCCAATGCTTTCGACTTTCGGAGGTC 154 ACGTTAGTATGATTTTACGAAACGGATTTGGAAAGTCGAGATATTTGCAAAATCTCATCA 155 TCGTC 156 157 158 (5) L. corrodens AACATCGATATTTCGAACGCACATTGCCGAGCTAGGATTTTTTCCGGCTCGACGTCTGT 159 CTGAGGGTCGATGATTTCTAATAAACAGATTTTCCGTTAGAGTCAATCTATGGAAAAATT 160

- 166 (6) *L. brunnea*

AACATCGATATTTCGAACGCACATTGCGCTCTGGGGGAACTCCGTTCCCGGAGCACGTCT 167 GTCTGAGGGTCGATGCTTACTTGTAAAATTTAGAACGCTGGTTATTTTTTCGAATCGC 168 169 AAACGATTGAAAAAAAAATCTTTTTAACCGGCGTGATAAAATGATGAAAGCTTCGGGT CGCTCTCCGCGTAAGCGGAGGGTTCCGTCTTTCGAAATGCACGATCCCGAGATATCGCT 170 CGGAAATAAAACGAGTTCGAAGAAGACCGATACGTATAATAGTCTCGAGCCGCCAGCG 171 172 **AGATCCTTGTTACGAA**TGCACAAACGTAATAATAATAATAAGGATCGAAGAAGGTCTT 173 174 CCTCTCGTCACGTTAGTGGTTATTTTGATATGTAACGTGAAACGGTCGAGATGGAAAAC 175 GTATCGCGGGTAAAGACGAGAAAAAGAGGAGGCTTTCGAACCGTGTTTTTATCGCGGC TCTCGGAAACGAAGATCGGTTTTCGTGTACAAGCGTTTGTTAGTAGTACTCGTAAAAC 176 **AGAATTCCGGC**GATTTCTTTCATCGTC 177

- 178
- 179 (7) *L. rufa*

180 AACATCGATATTTCGAACGCACATTGCGCTCTGGGGGAACTCCGTTCCCGGAGCACGTCT GTCTGAGGGTCGGTGCTTAATGTTAAATGTCGGAACGCTTGACGGCGTGATCGAAAC 181 GTTGAAAGCTTCGGGTCGTGAAAATTAAGACCCGTCTTTCCAAATGCACGAAAGACCG 182 183 AGAATCTTATGAGGAAGAAGAACACTATAGAGAACGATACGTCAATCTCACGGCTGCT 184 CGTAAAGCAAATTAAAAGTACAT**TAGCGTACCGATAGGTCGTAA**ATTTTTTTAAAG CGAGCCCGTAAGTAAACGAAAAAGAAAAGATCGCACGCATCGTCGTCGATCTTCCTCG 185 TCACGTTCATGATGATACATAGCAAATAAATAAATAAAAGAAAAACGGTCGAGGGTACT 186 GCGGGGATTCGGTATAGCGTGAGGAAACGAAAATCGTTACTCGTCGAAATTGTGTTTTT 187 TATTTTTTTTCACCTCAAGAAAAGATTCTTTTCATCGTC 188

189

190 (8) *L. pearmani*

AACATCGATATTTCGAACGCACATTGCGCTCTGGGGGAACTCCGTTCCCGGAGCACGTCT 191 GTCTGAGGGTCGATGCTTAATTGTAAAATCTAGAACGCCGTCTCTCGCACGATTAAGTT 192 CGGGGGGGGGGGCGTCGTGATAAAAGACTCCGTTGAATGCTTCGAGCTCGCGAGAGCCC 193 GTCATTCCAAATGCACGAAACGAGATTCTCACAAGAATTTCGAACGAGTCGGAAGAT 194 GAGAGAAACGATACGTAAATCGTACTTGCTATCTACGGAAGCCAATATTTGCCCGTGAT 195 ACGAAACATGTCTTCCTCGTCACGTTAGTCAGTTTGTTAATAATAAGCAAAATAACGGG 196 197 CGAGACGTTGATGTATCGCCGCGGCGGCTAGCAGTAGATCGTACGGAAACGAAAATCG TTTACTCGTTTGAAGAATGGTAGCCGTTTTTCGTTGGTAGCCTTTCGACCAAACCGAA 198 GTGAGATTCTTTTCATCGTC 199

201 (9) *L. mendax*

201	() L. menuux
202	AACATCGATATTTCGAACGCACATTGCGGTCCTTGGAACTTTGTTCCCGGACCACGTCT
203	<u>GTCTGAGGGTCGGTGTTTGCT</u> ATTAGAACATTTTTATTCGTTTCGCTTCGGCACGAA
204	TAATGTGGAGGTTTCGAGTATCGGCTCGTCTTCCTAAACAGAATAATGCGAGAATTCCG
205	TAAGCGGGAATTTATGGGTTTTGTCCTCGCGTCACGTTTCTGAAATTAAGAGCAGCGT
206	$\underline{\mathbf{TGACGGACTT}} \mathbf{GCGAGGTAAAGCTCTGTTCGGCGAAATAGACGGAAATTCTAATTCATC}$
207	GTC
208	
209	(10) L. tricolor
210	AACATCGATATTTCGAACG <mark>CACATTGCCGAACTTTGAATT</mark> TATTCTGGTTCGACGTCT
211	GTCTGAGGGTCGATGAAAATTATTGAAATATTTACCGTTAGATGTAATGTCTACGGTTAT
212	TGAAAGTTACGAATTTCCTCGGAAATTCGTCTTTCGAAATTCATATGTCCCCGAGAATC
213	GTACATGAAAGAGAATGTCTCAGTAAATGGTATGTTATTTGCTGCATTGGGCATATTTCT
214	GCTTCACGTTAGTAAACCAAACGGATAGTAGAAAATGTCTTCGTGGAACGATTTCTC
215	ATCTTCGTCCTCAGATCAGGAATCACTAGTGAATTCGCGGCCGCCTGCAGGTC
216	

Population	Locations	Authors	No. GenBank
L. entomophila_BJ- P. R. China	Beijing, P. R. China	Cui Bingyi, Li Zhihong	KC707588
L. entomophila_HuB-P. R. China	HuBei,P. R. China	Cui Bingyi, Li Zhihong	KC707590
L. entomophila_GX- P. R. China	Guangxi, P. R. China	Cui Bingyi, Li Zhihong	KC707589
L. entomophila_SD- P. R. China	Shandong,P. R.China	Cui Bingyi, Li Zhihong	KC707591
L. bostrychophila_BJ- P. R. China	Beijing, P. R. China	Cui Bingyi, Li Zhihong	JQ966091
L. bostrychophila_GX- P. R. China	Guangxi, P. R. China	Cui Bingyi, Li Zhihong	JQ966093
L. bostrychophila_GZ- P. R. China	Guangzhou,P.R.China	Cui Bingyi, Li Zhihong	JQ966096
L.bostrychophila_HeN- P. R. China	Henan, P. R. China	Cui Bingyi, Li Zhihong	JQ966097
L. bostrychophila_CQ- P. R. China	Chongqing, P. R. China	Cui Bingyi, Li Zhihong	JQ966099
L. bostrychophila_P-CZ	Prague,CZ	Cui Bingyi, Li Zhihong	JQ966092
L. bostrychophila_CZ	Central Bohemia, CZ	Cui Bingyi, Li Zhihong	JQ966095
L. bostrychophila_USA	Manhattan,USA	Cui Bingyi, Li Zhihong	JQ966094
L. bostrychophila_GER	Berlin ,GER	Cui Bingyi, Li Zhihong	JQ966098
L. decolor_CQ- P. R. China	Chongqing, P. R. China	Cui Bingyi, Li Zhihong	KC707592
L. decolor_YN- P. R. China	Yunnan, P. R.China	Cui Bingyi, Li Zhihong	KC707593
L. decolor_P-CZ	Prague,CZ	Cui Bingyi, Li Zhihong	KC707594
L. decolor_P-CZ	Prague,CZ	Cui Bingyi, Li Zhihong	KC707595
L. decolor_P-CZ	Prague,CZ	Cui Bingyi, Li Zhihong	KC707596
L. decolor_USA	USA	Cui Bingyi, Li Zhihong	KC707597
L. decolor_USA	USA	Cui Bingyi, Li Zhihong	KC707598
L. decolor_USA	USA	Cui Bingyi, Li Zhihong	KC707599
L. paeta_USA	USA	Cui Bingyi, Li Zhihong	GU563533
L. paeta_HeB-P. R.China	HebeiP,R.China	Cui Bingyi, Li Zhihong	KC707604
L. paeta_SDT-P. R.China	Taian,Shandong,P.R.China	Cui Bingyi, Li Zhihong	KC707605
L. paeta_SDT-P. R.China	Taian,Shandong,P.R.China	Cui Bingyi, Li Zhihong	KC707606
L. paeta_SDT-P. R.China	Taian,Shandong,P.R.China	Cui Bingyi, Li Zhihong	KC707607
L. paeta_SDC-P. R.China	Caoxian,Shandong,P.R.China	Cui Bingyi, Li Zhihong	KC707619
L. paeta_ZJ-P. R.China	Zhejiang, P. R. China	Cui Bingyi, Li Zhihong	KC707612
L. paeta_ZJ-P. R.China	Zhejiang, P. R. China	Cui Bingyi, Li Zhihong	KC707613
L. paeta_HuB-P. R.China	Hubei, P. R. China	Cui Bingyi, Li Zhihong	KC707614
L. paeta_HuB-P. R.China	Hubei, P. R. China	Cui Bingyi, Li Zhihong	KC707615
L. paeta_HeN-P. R.China	Henan, P. R.China	Cui Bingyi, Li Zhihong	KC707616
L. paeta_HeN-P. R.China	Henan, P. R. China	Cui Bingyi, Li Zhihong	KC707617
L. paeta_HeN-P. R.China	Henan, P. R. China	Cui Bingyi, Li Zhihong	KC707618
L. paeta_CZ	Prague,CZ	Cui Bingyi, Li Zhihong	KC707603

Table S3 The number of 53 specific sequences submitted to Genebank

L. paeta_USA	USA	Cui Bingyi, Li Zhihong	KC707608
L. paeta_USA	USA	Cui Bingyi, Li Zhihong	KC707609
L. paeta_USA	USA	Cui Bingyi, Li Zhihong	KC707610
L. paeta_USA	USA	Cui Bingyi, Li Zhihong	KC707611
L. corrodens_CZ	Prague,CZ	Cui Bingyi, Li Zhihong	KC707623
L. corrodens_DMK.	Danmark	Cui Bingyi, Li Zhihong	KC707620
L. corrodens_DMK.	Danmark	Cui Bingyi, Li Zhihong	KC707621
L. corrodens_DMK.	Danmark	Cui Bingyi, Li Zhihong	KC707622
L. corrodens_USA	USA	Cui Bingyi, Li Zhihong	KC707624
L. corrodens_USA	USA	Cui Bingyi, Li Zhihong	KC707625
L. brunnea_P-CZ	Prague,CZ	Cui Bingyi, Li Zhihong	KC707626
L. brunnea_USA	USA	Cui Bingyi, Li Zhihong	KC707626
L. mendax_JS-P. R. China	Jiangsu, P. R. China	Cui Bingyi, Li Zhihong	KC707629
L. pearmani_USA	USA	Cui Bingyi, Li Zhihong	KC707628
L. tricolor_SD-P. R. China	Shandong, P. R.China	Cui Bingyi, Li Zhihong	KC707630
L. tricolor_SD-P. R. China	Shandong,P. R.China	Cui Bingyi, Li Zhihong	KC707631
L. tricolor_SD-P. R. China	Shandong,P. R.China	Cui Bingyi, Li Zhihong	KC707632
L. rufa_USA	USA	Cui Bingyi, Li Zhihong	KC707629



Figure S1 The electrophoresis strip for specificity estimation of species-specific primers (LBF and LBR) of *L. bostrychophila* (M: D2000 DNA Marker; 1: *L. bostrychophila* (Guangxi); 2: *L. bostrychophila* (Henan); 3: *L. bostrychophila* (Chongqing); 4: *L. bostrychophila* (Berlin); 5: *L. bostrychophila* (USA); 6: *L. bostrychophila*(Prague); 7 *L. bostrychophila* (Beijing); 8: *L. bostrychophila* (Guangzhou); 9: *L. Entomophila* (Shandong); 10: *L. Decolor* (Yunnan); 11: *L. paeta* (Zhejiang); 12: *L. corrodens* (USA); 13: *L. Brunnea* (Prague); 14: *L. Rufa* (USA);
15: *L. Pearmani* (USA); 16: *L. mendax* (Jiangsu); 17: *L. Tricolor* (Shandong); CK: Negative control)



Figure S2 The electrophoresis strip for specificity estimation of species-specific primers (21LEnF and 208LEnR-2)
of *L. Entomophila* (M: DNA Marker II; 1: *L. entomophila* (Beijing); 2: *L. entomophila* (Guangxi); 3: *L. entomophila* (Prague); 4: *L. entomophila* (Chongqing); 5: *L. bostrychophila* (Guangxi); 6: *L. bostrychophila*(Yunnan); 7: *L. Decolor* (Yunnan); 8: *L. corrodens* (USA); 9: *L. brunnea* (Prague); 10: *L. rufa* (USA); 11: *L. pearmani* (USA); 12: *L. mendax* (Jiangsu); 13: *L. tricolor*(Shandong); CK: Negative control)



Figure S3 The electrophoresis strip for specificity estimation of species-specific primers (164LDeF and 319LDeR)
of *L. decolor* (M: DNA Marker II; 1 and 2: *L. Decolor* (Yunnan); 3 and 4: *L. Decolor* (Chongqing); 5: *L. Entomophila* (Shandong); 6: *L. bostrychophila* (Chongqing); 7: *L. paeta* (Shandong); 8: *L. corrodens* (USA); 9: *L. Brunnea* (Prague); 10: *L. Rufa* (USA); 11: *L. mendax* (Jiangsu); 12: *L. Pearmani* (USA); 13: *L. Tricolor*(Shandong); CK: Negative control)



Figure S4 The electrophoresis strip for specificity estimation of species-specific primers (LPa15F and LPa180R)
of *L. paeta* (M: DNA Marker II; 1: *L. paeta* (Caoxian, Shandong); 2: *L. paeta* (Taian, Shandong); 3: *L. paeta* (Zhejiang); 4: *L. paeta* (Hubei); 5: *L. paeta* (USA); 6: *L. entomophila* (Shandong); 7: *L. bostrychophila*(Guangxi); 8: *L. decolor* (Yunnan); 9: *L. corrodens* (Prague); 10: *L. brunnea* (USA); 11: *L. Rufa* (USA); 12: *L. paetani* (USA); 13: *L. mendax* (Jiangsu); 14: *L. tricolor* (Shandong); CK: Negative control)

Figure S5 The electrophoresis strip for specificity estimation of species-specific primers (LC170F and LC277R)

of L. corrodens (M: D2000 DNA Marker; 1: L. corrodens (Danmark); 2: L. corrodens (Prague); 3: L. corrodens

256 (USA); 4: L. entomophila (Shandong); 5: L. bostrychophila (Guangzhou); 6: L. decolor (Chongqing); 7: L. paeta

257 (Hubei); 8: L. brunnea (USA); 9: L. rufa (USA); 10: L. tricolor (Shandong); 11: 11: L. mendax (Jiangsu); 12: L.

258 *pearmani* (USA); CK: Negative control)

- **262** Figure S6 The electrophoresis strip for specificity estimation of species-specific primers (LBr350F and LBr577R)
- 263 of L. brunnea (M: D2000 DNA Marker; 1: L. brunnea (Prague); 2: L. brunnea (USA); 3: L. entomophila (Beijiing);
- 264 4: L. bostrychophila (Guangzhou); 5: L. decolor (USA); 6: L. paeta (Shandong); 7: L. corrodens (USA); 8: L. rufa
- 265 (USA); 9: L. pearmani (USA); 10: L. tricolor (Shandong); 11: L. mendax (Jiangsu); CK: Negative control)

Figure S7 The electrophoresis strip for specificity estimation of species-specific primers (78LRuF-3 and 276LRuR-3) of *L. rufa* (M: D2000 DNA Marker II; 1, 2, and 3: *L. rufa* (USA); 4: *L. entomophila* (Beijiing); 5: *L.*

272 bostrychophila (Guangxi); 6: L. decolor (Yunnan); 7: L. paeta (Prague); 8: L. corrodens (USA); 9: L. brunnea

273 (USA); 10: *L. mendax* (Jiangsu); 11: *L. pearmani* (USA); 12: *L. tricolor* (Shandong); CK: Negative control)

277	Figure S8 The electrophoresis	strip for specificit	v estimation of species	s-specific primers	(186LPeF and 436LPeR)
_,,	i iguie bo ine electrophotesis	sup for specificit	y countration of species	, specific primers	(TOOLI OF and TOOLI OR)

- 278 of L. pearmani (M: D2000 DNA Marker; 1 and 2: L. pearmani(USA); 3: L. entomophila (Beijiing); 4: L.
- bostrychophila (Guangxi); 5: L. decolor (Yunnan); 6: L. paeta (Prague); 7: L. corrodens (USA); 8: L. brunnea
- 280 (USA); 9: L. rufa (USA); 10: L. mendax (Jiangsu); 11: L. tricolor (Shandong); CK: Negative control)
- 281

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283	

- 284 Figure S9 The electrophoresis strip for specificity estimation of species-specific primers (LM60F and LM224R) 285 of L. mendax (M: D2000 DNA Marker; 1 and 2: L. mendax (Jiangsu); 3: L. entomophila (Beijiing); 4: L. 286 bostrychophila (Guangxi); 5: L. decolor (Yunnan); 6: L. paeta (Prague); 7: L. corrodens (USA); 8: L. brunnea 287 (USA); 9: L. rufa (USA); 10: L. pearmani (USA); 11: L. tricolor (Shandong); CK: Negative control)
- 288

- 290
- **Figure S10** The electrophoresis strip for specificity estimation of species-specific primers (LTri20F and LTri249R)
- of L. tricolor (M: D2000 DNA Marker; 1 and 2: L. tricolor (Shandong); 3: L. entomophila (Beijiing); 4: L.
- bostrychophila (Guangxi); 5: L. decolor (Yunnan); 6: L. paeta (Prague); 7: L. corrodens (USA); 8: L. brunnea
- 294 (USA); 9: L. rufa (USA); 10: L. mendax (Jiangsu); 11: L. pearmani (USA); CK: Negative control)
- 295

Figure S12 The sensitivity estimation of species-specific primers with different concentration of template
 DNA of *L. entomophila* (M: D2000 DNA Marker; 1: 40ng; 2: 20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001
 ng; 8: Negative control)

- Figure S13 The sensitivity estimation of species-specific primers with different concentration of template
- DNA of *L. decolor* (M: D2000 DNA Marker; 1: 40ng; 2: 20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8:
- Negative control)

Figure S14 The sensitivity estimation of species-specific primers with different concentration of template DNA of *L. paeta* (M: D2000 DNA Marker; 1: 40ng; 2: 20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8: Negative control)

Figure S15 The sensitivity estimation of species-specific primers with different concentration of template DNA of L. corrodens (M: D2000 DNA Marker; 1: 40ng; 2: 20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8: Negative control)

Figure S16 The sensitivity estimation of species-specific primers with different concentration of template
 DNA of *L. brunnea* (M: D2000 DNA Marker; 1: 40ng; 2: 20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8:
 Negative control)
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Figure S17 The sensitivity estimation of species-specific primers with different concentration of template
 DNA of *L. rufa* (M: D2000 DNA Marker; 1: 40ng; 2: 20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8:
 Negative control)

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335 Figure S18 The sensitivity estimation of species-specific primers with different concentration of template

336 DNA of *L. pearmani* (M: D2000 DNA Marker; 1: 40ng; 2: 20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng;

8: Negative control)

339

Figure S19 The sensitivity estimation of species-specific primers with different concentration of template
DNA of *L. mendax* (M: D2000 DNA Marker; 1: 40ng; 2: 20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8:
Negative control)

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Figure S20 The sensitivity estimation of species-specific primers with different concentration of template
DNA of *L. tricolor* (M: D2000 DNA Marker; 1: 40ng; 2: 20ng; 3: 10ng; 4: 1ng; 5: 0.1ng; 6: 0.01ng; 7:0.001 ng; 8:
Negative control)